Evaluation of Recombinant Human Growth Hormone Secretion in E. coli using the L-asparaginase II Signal Peptide

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Abstract

Background: In the recent years, there has been an increasing interest in secretory production of recombinant proteins, due to its various advantages compared with intracellular expression. Signal peptides play a critical role in prosperous secretion of recombinant proteins. Accordingly, different signal peptides have been assessed for their ability to produce secretory proteins by trial-and-error experiments. The aim of this study was to evaluate the effect of L-asparaginase II signal peptide on the recombinant human Growth Hormone (hGH) protein secretion in the Escherichia coli (E. coli) host.

Methods: Cloning and expression of a synthetic hGH gene, containing L-asparaginase II signal sequence was performed in E. coli BL21 (DE3) using 0.1mM IPTG as an inducer at 23°C overnight. Periplasmic protein extraction was performed using three methods, including osmotic shock, osmotic shock in the presence of glycine and combined Lysozyme/EDTA osmotic shock. Afterwards, the hGH expression was determined by SDS-PAGE.

Results: Based on experimentally obtained results, hGH protein is expressed as inclusion body even in the presence of L-asparaginase II signal peptide.

Conclusion: Therefore, this signal peptide is not effective for secretory production of the recombinant hGH.

Keywords: Escherichia coli (E. coli), Human growth hormone, L-asparaginase II, Recombinant proteins, Signal peptide

Introduction

Human Growth Hormone (hGH) or somatotropin, a single chain polypeptide contains 191 amino acid residues with a molecular mass of 22 kDa, is synthetized and secreted by the anterior pituitary gland 1,2. The mature form of the hGH is created from its 217 amino acid precursor after removal of the signal peptide (26 amino terminal residues). Owing to a substantial role of the hGH in various biological functions, it has a wide range of therapeutic applications like hormone therapy of hypopituitary dwarfism, skin burns, bone fractures, bleeding ulcers, HIV wasting syndrome and genetic disorders such as Turner’s and Down’s syndromes 3-5. Since this protein does not need post-translational modifications like glycosylation, Escherichia coli (E. coli) host is considered a prokaryotic powerhouse for industrial production of recombinant hGH 6,7,8. Notwithstanding different advantages of E. coli, including appropriate growth on low-cost media, fast biomass accumulation 6,9 and extensive knowledge of its genetics and physiology 10, there are some obstacles in acquiring considerable yields of correctly folded recombinant proteins. Failure of protein to rapidly reach a correct folding and disulfide bond formation leads to protein degradation by proteases and accumulation as inactive aggregates in the cytoplasm called inclusion bodies 6,11,12. Transferring the protein into the bacterial periplasmic region of E. coli or directly into the extracellular medium, using an appropriate signal peptide at the N-terminal of the protein, is an applicable approach to solve these problems 11. Efficient secretion of the recombinant protein is achieved using an optimal signal sequence, which is compatible with the secretory protein. Unfortunately, there is no general rule for selecting a suitable signal peptide to guarantee successful protein secretion. Therefore, trial-and-error experimental approach is universally applied to evaluate different signal peptides 11. Uptill now, various signal peptides have been evaluated for secretory production of hGH in E. coli, like OmpA 13,14, PhoA 15, pelB 14,16, LTB 17, npr 18, Stt 19,20, DsbA 18, penicillinase 21 and natural hGH signal peptide 22.

In a continuing effort, the purpose of the current...
study was to investigate the influence of a bacterial signal peptide, L-asparaginase II, on the recombinant hGH protein secretion for the first time.

Materials and Methods

Bacterial strain, plasmid, and culture growth conditions

The BL21 (DE3) E. coli host strain (Novagen, Germany) carrying T7 RNA polymerase gene and pET15b plasmid (Pasteur Institute, Iran) carrying T7 promoter were applied for cloning and expression steps. Cells were grown at 37 °C in Luria-Bertani (LB) broth medium consisting of 10 g/L tryptone, 5 g/L yeast extract, and 5 g/L NaCl with shaking at 200 rpm. LB medium was supplemented with 100 μg/ml ampicillin for the growth of recombinant cells 23.

Design and construction of hGH secretory expression plasmid

Codon optimization of the hGH gene (Accession no. NM_000515) was performed by Codon Usage Wrangler server at http://www.mrc-lmb.cam.ac.uk/ms/methods/codon.html to apply the E. coli preferred codons and consequently, achieve high-level expression in this prokaryotic expression host. In order to evaluate the effect of L-asparaginase II signal peptide on secretion of hGH, its nucleotide sequence from E. coli K-12 (Accession no. NC_000913.3) was obtained from database resources of the National Center for Biotechnology Information (NCBI) at www.ncbi.nlm.nih.gov and inserted in the N-terminal of the optimized hGH gene. The signal peptide cleavage site was predicted using SignalP server (version 4.0), which is based on neural network method 24. Codon-optimized synthetic hGH containing L-asparaginase II signal sequence was prepared from Generay Biotech (Shanghai, China), after embedding the NdeI and BamHI restriction endonuclease cut sites in the N- and C-terminal of the gene construct, respectively. Synthetic gene and the bacterial expression vector (pET-15b) were digested with NdeI and BamHI enzymes (Fermentas, EU), and the ligation step was performed using Rapid DNA Ligation Kit (Fermentas, EU). Competent E. coli BL21 (DE3) cells were transformed with ligation mixture using TransformAid TM Bacterial Transformation Kit (Fermentas, EU). After extraction of recombinant plasmids from transformed E. coli, PCR analysis was applied to check the proper insertion of the gene in the recombinant plasmid using T7 promoter-specific primers. The nucleotide sequence of recombinant insert was verified by automated DNA sequencing (SinaClon BioScience Co., Iran).

Induction of protein expression and periplasmic protein extraction

Transformed cells were cultivated in LB medium containing 100 μg/ml ampicillin at 37 °C until a cell density of OD600=0.5-0.7 was achieved before induction. 0.1 mM IPTG (isopropyl-β-d-thiogalacto-pyranoside), purchased from Fermentas, EU, was applied to induce protein expression and cultivation was continued at 23 °C with shaking (200 rpm) overnight. The cell pellets were then harvested for periplasmic protein extraction by centrifugation at 4,000×g for 10 min and the supernatant was saved for analysis by SDS-PAGE. Periplasmic extraction of the protein was performed using three methods. According to the first method, osmotic shock, cell pellets were resuspended in sucrose/EDTA solution (20%). After incubation for 15 min at room temperature, the osmotically fragile cells were harvested by centrifugation at 4000×g for 10 min. The supernatant was removed and the cell pellet was resuspended in 1 ml cold distilled water (4°C) and incubated with gentle agitation at 4°C for 15 min. The resulting spheroplasts were harvested by centrifugation (13000×g at 4°C) and the supernatant, referred to as the periplasmic fraction, was recovered. The pellet was saved for analysis by SDS-PAGE. The second method was the osmotic shock in the presence of glycine, in which 1% glycine was added together with IPTG in the induction time. In the third procedure, combined lysosome/EDTA osmotic shock, the cell pellets were resuspended in sucrose/lysozyme solution containing 20% sucrose, 0.2 M Tris-HCl, 0.05% lysozyme, and 1 mM EDTA. The cell suspension was then incubated statically for 15 min at room temperature. Thereafter, equal volume of cold water was added and the suspension was incubated on ice for a further 15 min. Finally, the periplasmic fraction was recovered by centrifugation (13000×g at 4°C). The supernatant was removed and labeled as periplasmic fraction 25.

Expression analysis using SDS-PAGE

SDS-PAGE was performed in accordance with Laemmli 26 under reducing conditions, and protein bands were stained by Coomassie Brilliant Blue. Resolving and stacking gels were prepared in final concentrations of the 15 and 5%, respectively.

Results

High-level cytoplasmic expression of recombinant hGH in E. coli frequently leads to aggregation of misfolded protein 8,27. To overcome this obstacle, an alternative expression approach can be applied to secrete the protein into the periplasmic space of the E. coli by inserting a signal sequence to the N-terminal of the hGH gene. It has been demonstrated that naturally secreted proteins like hGH, are universally considered to be appropriate candidates for the secretory production in heterologous systems 8. An optimal balance between all stages of the secretory cascade is required for a prosperous secretion process. One of the most important factors that has a considerable effect on all steps of secretion pathway and finally, on the yield of the recombinant secreted protein is the signal peptide 28. This short amino-terminal extension is then proteolytically removed by specific enzymes called signal peptidases during translocation to yield the correctly
Evaluation of hGH Secretion with L-Asparaginase II Signal Peptide

It has been demonstrated that substantial difference of eukaryotic and prokaryotic codon usages is one of the principal reasons for non-efficient expression of the foreign genes in E. coli. Previous reports also indicated that codon optimization for E. coli considerably enhances the mammalian protein expressions in this host.

The native sequence of L-asparaginase II signal peptide from E. coli K-12 was fused to the N terminal of the optimized hGH gene to assist in protein secretion across the cytoplasmic membrane. The cleavage site of this signal peptide was evaluated using SignalP server (version 4.0). Although several computational tools are available for prediction of the signal sequences and location of their cleavage site, SignalP ranks among the most precise and reliable tools, which provides high-throughput processing of protein sequences with accuracy of 87%. Figure 1 schematically demonstrates SignalP outputs, including various scores (C, S, Y, S-mean and D), and the cleavage site. Predicted D-score for L-asparaginase II signal peptide in connection with hGH gene was 0.61 (>0.5), which indicated an acceptable probability of acting as a signal peptide. The sequence was flanked by NdeI/BamHI restriction sites and successfully inserted into the corresponding sites on the expression vector pET15b to transform E. coli BL21 (DE3) cells. The structure of the synthetic gene is illustrated in figure 2. As shown in figure 3, PCR analysis using T7 promoter-specific primers verified the exact insertion of the gene in the recombinant plasmid. Automated DNA sequencing (SinaClon BioScience Co., Iran) also confirmed the correct insertion of the synthetic nucleotide sequence (data not shown).

**Induction of protein expression and periplasmic protein extraction**

It has been demonstrated that, reduction of culture temperature and IPTG concentration can be helpful to decrease the probability of protein aggregation in the form of inclusion bodies. Therefore, protein expression was induced by 0.1 mM IPTG at 23 °C overnight. Different strategies have been employed to enhance the permeability of the outer membrane and promote extracellular secretion of recombinant proteins from E. coli.
coli. These procedures include mechanical methods (ultrasound), physical methods (osmotic shock, freezing and thawing), chemicals (addition of magnesium, calcium, EDTA, glycine, and Triton X-100), and enzymatic (lysozyme) treatments 11,39-41. In our study, three methods were applied to extract the periplasmic proteins, after protein expression. The first applied method for this purpose was the osmotic shock, as one of the most widely used methods of periplasmic release, which leads to the outer membrane of the cells to become permeable using hypertonic solutions 25,42,43. Previous studies reported that osmotic shock alone did not efficiently release the Streptomyces thermoviolaceus (S. thermoviolaceus) α-amy-lase from the periplasm of E. coli 42. It has been also demonstrated that glycine supplementation may slightly disrupt peptidoglycan cross-linkages and cell membrane integrity 44. On the other hand, glycine supplemented to the medium retarded formation of inclusion bodies and enhanced the efficiency of extracellular production of recombinant proteins 44,45. Therefore, as the second method in this study, osmotic shock was performed on cells, which were cultured in the glycine-supplemented medium. In 1996, French et al demonstrated that in comparison with other periplasmic extraction methods, combined lysozyme/osmotic shock was the most appropriate method for recovery of a recombinant protein from the periplasm of E. coli cells 25. Therefore, this method was also performed for periplasmic protein extraction in our study. The SDS-PAGE analysis of protein expression and extraction using these three methods are described in figure 4.

Discussion

Based on the obtained results, no hGH protein band was observed in the supernatant and periplasmic fractions. In contrast, a major band of hGH protein was seen in the cytoplasmic fraction. Consequently, hGH protein is expressed as inclusion body.

Based on the obtained results, almost all of the recombinant hGH was expressed in the cytoplasm as inclusion body even in the presence of L-asparaginase II signal sequence, which indicates no efficient role for this signal peptide in the hGH protein secretion in E. coli BL21 (DE3). There are several interpretations that could explain why this problem arose. A prosperous secretion process generally needs an optimal balance between all stages of the secretory cascade. Inefficient secretion might be due to several factors like the structural characteristics of the signal peptide, ineffective cleavage of signal peptide from hGH protein, incompatibility between the mature protein and the signal peptide, inability to cross over the membrane and reach the periplasmic space or insufficient balance between the capacity of the export machinery and rate of protein synthesis 25. One of the most important concerns in secretory production is to understand the characteristics of the target protein that permits the secretion process to occur. This still remains unsolved and makes scientists apply the trial-and-error approach rather than rational evaluation 46. Taken together, although native and mutant L-asparaginase II signal peptide were efficient for recombinant hirudin III and cyclodextrin glucanotransferase secretion in E. coli, respectively 33,34, hGH secretion was not achieved in the presence of this signal peptide. The experiences gained from this study...
and other previous investigations, which reported low secretion yields, prompted us to apply bioinformatics tools as an alternative approach instead of time-consuming, costly and labor-intensive trial-and-error strategy. Although our in silico study was conducted after the present study, its results were published prior to the current experimental report. Results of both studies were in agreement and indicated that L-asparaginase II signal peptide is not suitable for hGH secretion.

Conclusion
Considering the significant advantages of secretory production of recombinant proteins, this study was designed to evaluate the possible role of a bacterial signal peptide, L-asparaginase II, in the hGH protein secretion in the E. coli host. The results of the current experimental study and our earlier in silico published study have demonstrated that successful secretion of hGH is not achieved using L-asparaginase II signal sequence. This study provides confirmatory evidence that although one signal peptide can be suitable for secretion of one protein, it is not necessarily appropriate for secretion of other proteins.

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Conflict of Interest
The authors declare that they have no conflict of interest.

Ethical Approval
This article does not contain any studies with human participants or animals performed by any of the authors.

References


