

Generation of *In-vitro* Spermatogonial Stem Cells following Genetic Manipulation of Primordial Germ-like Cells

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Abstract

Research about potential use of stem cells for the development of germ line cells *in vitro* had been challenged. In the present study, we reported a novel protocol consisting of cocktail growth factor addition for germ cell differentiation followed by transfection. The cells were purified based on the expression on the cell surface of a protein. This protein is not present in normal cells of mice and does not interfere with cellular function. This cell surface marker is efficiently recognized by monoclonal antibodies. Bone marrow mesenchymal stem cells derived primordial germ like cells were differentiated to spermatogonial stem like cells by inducer cocktail including Retinoic acid (RA)+Leukemia inhibitory factor (LIF)+Basic fibroblast growth factor (bFgf). Co-culture system was used as a feeder under differentiated cells. A 400 bp fragment of spermatogonia-specific *Stra-8* locus was enough to direct gene expression to the germ line stem cells. *Stra8-CD4HAglo* construct was used for purification of pre-meiotic differentiated cells. Expression of pluripotency (*Pou5F1*, *Nanog*, *c-Myc*) and specific germ cell (*Mvh*, *PiwiL2*, *Stra-8*) genes in each stage were analyzed. The purified cells expressed the known molecular markers of PGC-like cells such as *Mvh*, *PiwiL2* & *Stra-8*. The outcomes of qPCR showed that ratio pluripotency of genes expression in selective group significantly decreased ($p \leq 0.05$) in the initial differentiation process. This results showed that ratio of *Pou5F1*, *Nanog*, *c-Myc*, *Mvh*, *PiwiL2* & *Stra-8* expression to purified PGC-like cells were 0.41, 0.204, 1.1, 0.003, 0.184 and 2.276, respectively. Treatment of cells with RA affected up regulation of *Stra-8*. Although, *c-Myc* gene as an oncogenic gene had significantly increased ($p \leq 0.05$) at the end of differentiation stage compared to initial phase of study, this level of expression could not be tumorigenic. qPCR results of the differentiation stage showed higher expression of *Stra-8* in co-culture+cocktail and co-culture groups. Also, there was a significant difference ($p \leq 0.05$) in the expression of *Pou5F1* & *Nanog*. Our results suggest that selection and purification of PGC-like cells based on *Stra-8* as a pre-meiotic marker is a useful tool for getting *in vitro* spermatogonial stem cell. This method facilitates identification of safely differentiated germ cells *in vitro*.

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Introduction

The development of Spermatogonia Stem Cells (SSCs) into gametes is an essential feature of sexual reproduction, although the development of SSCs and their differentiation

into sperm is well understood on the genes controlling gamete formation, particularly during the early stages of SSCs development ⁽¹⁾. Murine SSC originate from a founder

population of Primordial Germ Cells (PGCs) formed just after embryonic day 7 (E7.0) in the extra embryonic mesoderm posterior for the primitive streak. During migration a proliferation of PGC is regulated by multiple growth factors as well as cell-cell and cell-matrix interactions. Once they arrive in the genital ridge, the PGCs are enclosed by somatic Sertoli cells and become gonocytes⁽²⁾.

One of the major problems in the *in vitro* differentiation of germ cells is the lack of appropriate molecular markers for the characterization of the differentiated germ cells to distinguish them from the somatic cells⁽³⁾. Most of the markers used for PGCs are present in Stem Cells (SCs) as well: Fragilis, Mvh, SSEA1, c-Kit⁽⁴⁾. Therefore, it is very difficult to distinguish early germ cells from undifferentiated SCs. Meiotic and post-meiotic markers are more reliable markers, but it has been demonstrated that the progression through the meiotic process is still a challenge in the *in vitro* differentiation of gametes⁽³⁾. The transfection of germ line cells with marked or fluorescent proteins linked to specific gene promoters (genes implicated in pluripotency or germ cell line development) enables the visualization of the cells in which the specific gene of interest is expressed during the differentiation process⁽⁵⁻⁷⁾.

Because stem cells have unlimited potential to self-renew and produce differentiating and undifferentiating cells, heterogenous SSC transplantation offers the possibility of long-term restoration of pluripotential capability⁽⁸⁾. Cells stimulated by growth factors, such as the remaining undifferentiated cells, showed a dramatically increase in the expression of *c-Myc*, and over expression of *c-Myc* protein is observed in a wide variety of human cancers. However, *c-Myc* is also expressed as part of normal cellular functions, particularly during proliferation and differentiation⁽⁹⁾.

In this study, we introduced a novel method of *in vitro* SSC development. The differentiated PGC-like cells were purified by selection based on the expression of a pre-meiotic germ cells marker. The *Stra-8*, expressed in SSC,

appeared to be a useful candidate. We used a vector with a surface reporter maker. This vector based on the expression on the cell surface of a protein that is not present in the normal mouse, does not interfere with cellular function and is efficiently recognized by monoclonal antibodies. This method facilitates identification of safely differentiated germ cells *in vitro*.

Materials and Methods

Stem cell preparation, expansion, and differentiation

Bone Marrow Stromal Cells (BMSCs) were aspirated from the tibia and femur of 5 male NMRI mice (6-8 weeks old) and cultured according to the method described by Jamous et al.⁽¹⁰⁾. Bone Morphogenic Protein (BMP)-4 (25 ng/ml; Chemicon, USA) was added to the fourth-passage BMSCs daily for 4 days based on the method described by Mazaheri et al.⁽¹¹⁾. PGC-like cells were enriched using a co-culture system with 3 μ M retinoic acid (RA) [Sigma, Germany]. The inserts [Grainer, Germany] were placed in a 24-well plate [Grainer, Germany] containing mitomycin-treated MEF cell line (STO: C537, NCBI) and cultured for 7 days (unpublished data). CD4H Aglo construct was used for purification of premeiotic differentiated cells by magnetic-activated cell sorting (MACS: Meltenyi Biotec, Germany) based on *Stra-8* expression.

To induce purified *Stra-8*⁺ cells to SSCs, four groups were designed: a control (C; Dulbecco's Modified Eagle's Medium [DME D: Gibco, UK] supplemented 10% Fetal Bovine Serum [FBS: Gibco, South USA]), an inducer cocktail (IC; basic fibroblast growth factor [bFGF: Millipore, UK] 1 ng/ml + RA 3 μ M + Leukemia Inhibitory Factor [LIF: Millipore, UK] 10³ unit/ml), a co-culture system (CO; a feeder layer of Sertoli cells) and an inducer cocktail was followed by the addition of a feeder layer of Sertoli cells (IC + CO). 1 \times 10⁵ cells/ml were seeded into a 24 well tissue culture plate and treated with above experimental supplements during 2 days (Figure 1).

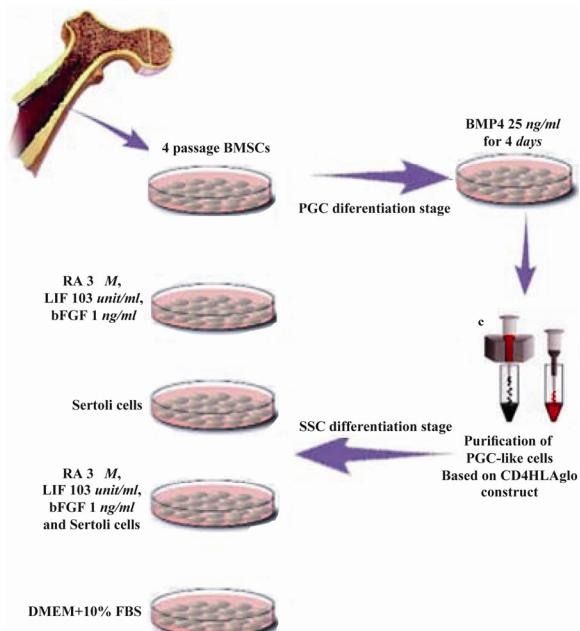


Figure 1. Schematic representation of experimental design during SSC-like cells differentiation stage from purified PGC-like cells

The expression of pluripotency genes (POU domain class 5 transcription factor 1 [*Pou 5F1*], *Nanog*, *c-Myc*) and germ line genes (mouse vasa homolog [*Mvh*], piwi-like homolog 2 [*Piwil2*], stimulated by retinoic acid [*Stra*]-8) in purified Stra-8⁺ cells and four experimental groups (C, IC, CO & IC + CO) were analyzed by quantitative polymerase change reaction (qPCR).

Cell transfection using a retroviral system

Plasmid construction: The expression of a foreign neutral surface marker to purify cells that express a transgene was performed using the Stra8-CD4HAglo construct. This construct was previously described⁽¹²⁾ and was a kind gift from Dr. Cuzin (University of Nice Sophia-Antipolis). Transfection with lipofectamine 2000 (Invitrogen, USA) was performed according to the manufacturer's guidelines. Initially, cells were plated in 96-well plates at 5×10^3 cells/well and allowed to grow overnight to achieve 70-80% confluence. Cells were transfected at different DNA: lipofectamine 2000 ratios ranging from 1-3 μ l lipofectamine 2000 and 0.5-1.5 μ g DNA. The reagent/DNA mixture was added to OptiMem medium (Invitrogen, USA) and the cells in-

cubated for 4-6 hr. Colonies resistant to G418 (400 mg/ml: Invitrogen, UK) were selected. Resistant colonies were analyzed by immunocytochemistry and flow cytometry to detect CD4 (Abcam, UK); colonies that contained the Stra8-CD4HAglo construct were isolated by MACS and cultured in a differentiated state.

Cultures were proliferated in the above described medium for differentiation of PGC-like cells into SSC-like cells. For optimization of the transfection protocol, the expression of GFP in mBMSCs was analyzed by fluorescence microscopy using a pCMV-Myc-Cyto-GFP plasmid (Addgene, USA) as a positive control for efficiency of transfection.

Magnetic activated cell sorting (MACS): Immunomagnetic isolation of CD4⁺ cells from transfected cells was performed using a CD4 Positive Isolation kit (130-045-101; Miltenyi Biotec, Germany) according to the manufacturer's instructions. Purified Stra-8⁺ cells were analyzed by immunocytochemistry and flow cytometry to detect CD4.

Immunocytochemistry: Purified Stra-8⁺ cells were fixed in 4% neutral buffered paraformaldehyde. Human T cells lymphocyte were isolated for immunocytochemical analysis as a control positive. Anti-CD4 antibody conjugated with FITC (0.1 μ g/10 μ l; AB86886, Chemicon, UK) was used for immunocytochemical staining. The images were captured using a Zeiss LSM 5 fluorescent microscope.

Quantitative analysis of gene expression: Total RNA was extracted from the purified PGC-like cells (Stra-8⁺ cells) and SSC-like cells in four experimental groups using RNX-Plus™ (Cinnagen, Iran) according to the manufacturer's recommendations. To eliminate genomic contamination, RNA was treated with DNase I using a kit (EN0521; Fermentas). Concentrations of RNA were determined by UV spectrophotometry (Eppendorff, Germany). The cDNAs were synthesized from 500 ng DNase-treated RNA samples with a RevertAid™ First Strand cDNA Synthesis kit (K1622; Fermentas, Germany) using oligo

(dT) primers. For PCR reactions, primers were adapted from other primers (designed by the NCBI website)^(6,7,13,14), and synthesized by Cinnagen (Table 1).

PCRs were performed using Master Mix and SYBR Green I in an Applied Biosystems, StepOne™ thermal cycler (Applied Biosystems, USA). The PCR program started with an initial melting cycle for 5 min at 95°C to activate the polymerase, followed by 40 cycles of melting (30 sec at 95°C), annealing (30 sec at 58°C) and extension (30 sec at 72°C). The quality of the PCR reactions was confirmed by melting curve analyses. Efficiency was determined for each gene using a standard curve (logarithmic dilution series of cDNA from the testes). For each sample, the reference gene (β_2M and β -actin) and target gene were amplified in the same run. Reference genes were approximately equal. The target genes were normalized to a reference gene and expressed relative to a calibrator (purified PGC-like cells).

Statistical analysis

Data were presented as mean \pm SD (standard deviation) and were analyzed using One-way repeated measure analysis of variance (ANOVA) followed by Tukey's post hoc test. The statistics software package SPSS Version 16.0 was used to perform the calculations. If $p<0.05$ then the result was judged to be significant. For the real time PCR data the log-

arithmic values had to be converted to real values by raising 2 to the power of the $\Delta\Delta Ct$ value before statistical analysis was performed. Flow cytometric assays (Becton Dickinson) was done and data were analyzed with the WinMDI 2.9 software.

Results

In vitro differentiation of bone marrow-derived PGC-like cells into SSC-like cells

We sorted Stra-8⁺ cells based on the cell surface expression of CD4. When purified PGC-like cells differentiated into SSC-like cells, the expression of the *Pou5F1* and *Nanog* were significantly decreased ($p\leq0.05$) in the all of the experimental groups except CO group, whereas the expression of *c-Myc* was significantly increased ($p\leq0.05$) compared to purified PGC-like cells. The expression of *Mvh* and *Piwil2* was decreased in the all of the experimental groups and that of *Stra-8* was significantly increased ($p\leq0.05$) in IC+CO group compared to purified PGC-like cells. Our results indicated that in the IC and C (without treatment or co-culture) groups, the expression of *Stra-8* was significantly ($p\leq0.05$) decreased, whereas there was no change in the CO group compared to purified PGC-like cells (Figure 2).

Our results indicated that the expression of *Pou5F1* and *Nanog* was significantly increased ($p\leq0.05$) in CO group, whereas there

Table 1. Primer used for real-time PCR

Genes	Primer sequence	GenBank code	Melt temperature (°C)
<i>Ddx4/Vasa</i> ^a	FOR: 5'- TGCCTTTGCTCCGCACCAT-3' REV: 5'- GGGTGAAGCACTTCAGGACC-3'	NM_001145885	74.2
β_2M ^b	FOR: 5'-TGACCGGCCTGTATGCTATC-3' REV: 5'-CACATGTCTCGATCCCAGTAG-3'	NM_009735	77.6
β -actin	FOR: 5'- TCCCTGGAGAAGAGCTACG- 3' REV: 5'- GTAGTTCTGTGGATGCCACA- 3'	NM_001101	79.2
<i>Stra-8</i> ^c	FOR: 5'-TCACAGCCTCAAAGTGGCAGG-3' REV: 5'-GCAACAGAGTGGAGGAGGAGT-3'	NM_009292	77.4
<i>Piwil2/milt</i> ^d	FOR: 5'- GCACAGTCCACGTGGTGGAAA -3' REV: 5'- TCCATAGTCAGGACCGGGAGGG -3'	NM_021308	81.8
<i>Oct-4/Pou5F1</i>	FOR: 5'- AGCACGAGTGGAAAGCAAC -3' REV: 5'-AGATGGTGGCTGGCTGAAC-3'	NM_013633	73
<i>Nanog</i>	FOR: 5'-CTGGGAACGCCTCATCAA-3' REV: 5'-CGCATCTTCTGCTTCCTGG-3'	ABO93574	75.4
<i>c-Myc/Myc</i>	FOR: 5'-CCCTCAGTGGCTTTCCCTAC-3' REV: 5'-CCACAGACACCACATCAATTTC-3'	NM_010849	78.9

a: *Vasa*, b: β_2M , c: *Stra-8* and d: *Piwil2* primers were reported by Toyooka et al (2003), Boroujeni et al (2008), Nayernia et al (2004) and Lee et al (2006), respectively

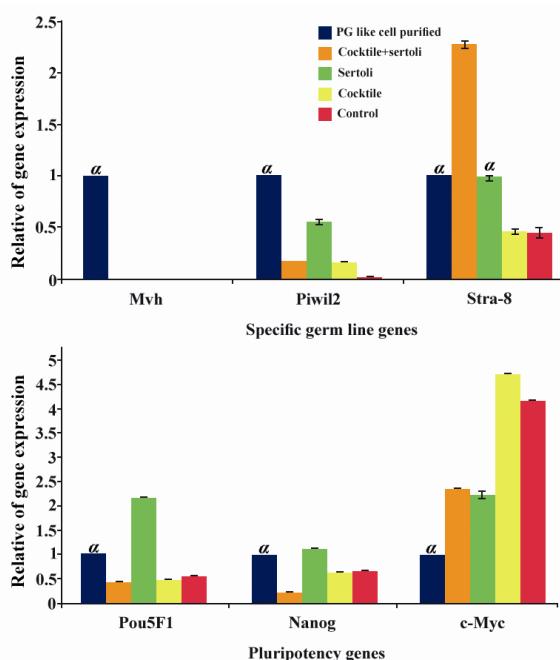


Figure 2. Real time-PCR analysis. The profile of mean calibrated specific germ line and pluripotency genes expression (y-axis) was shown in designed different groups (x-axis) for derivation of SSC-like cells. mRNA levels were normalized with respect to β actin, chosen as an internal control. Perimordial germ cell (PGC)-like cell purified was calibrator. Differentiation stage started with PGC-like cells. Histograms show mean expression values (\pm SD, n=3; p<0.05). a: significant difference with other groups. Cockticle component of LIF, bFGF and RA. Sertoli was used as feeder cells

was no change compared with other experimental groups. The expression of *Mvh*, *Piwil2* and *Stra-8* were significantly increased (p≤0.05) in C, CO and IC+Co groups, whereas these genes were significantly decreased (p≤0.05) in CO and IC groups, respectively. Also, the expression of *Stra-8* was down regulated in IC and C groups compared with other experimental groups (Figures 3A and 3B).

In further investigations, we sorted *Stra-8*⁺ cells based on the cell surface expression of CD4. At this stage, all pluripotency genes as well as in *Piwil2* were significantly decreased (p≤0.05) whereas the expression of *Stra-8* was significantly increased (p≤0.05) and that of *Mvh* was unchanged compared to the PGC-like cells (unpublished data).

Purification of *Stra-8*⁺ cells by MACS

We used a *Stra8-CD4HALglo* construct, harboring coding regions of human CD4 pro-

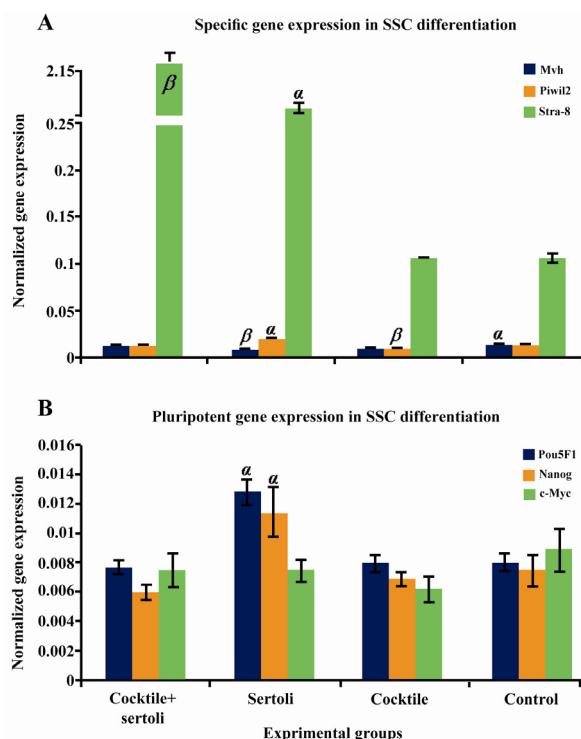


Figure 3. Real time-PCR analysis. The profile of mean normalized specific germ line and pluripotency genes expression (y-axis) was shown in designed different groups (x-axis) for derivation of SSC-like cells. mRNA levels were normalized with respect to β actin & β 2m, chosen as internal controls. Histograms show mean expression values (\pm SD, n=3; p<0.05). a, β: significant difference with other stages in same genes. SSC: Spermatogonial stem cells. Cockticle component of LIF, bFGF and RA. Sertoli was used as feeder cells

tein under the control of promoter regions of murine *Stra-8*, for transfection to induce the expression of human CD4 in PGC-like cell enrichment suspensions. We purified and cultured *Stra-8*⁺ cells using a procedure that supports expansion of isolated cells. After induction with RA (final concentration 3×10^6 M for 7 days with feeder cells), we identified a small population of cells (22%) within cultured adherent PGC-like cells that expressed human CD4. However, we purified approximately 80% *CD4*⁺ cells using MACS. Our observations are shown in figure 4.

Discussion

Efficient derivation of SSCs from different sources of stem cells *in vitro* has been challenging in the treatment of male infertility^(6,15-17). In this study, we reported a protocol of growth factor addition for differentiation fol-

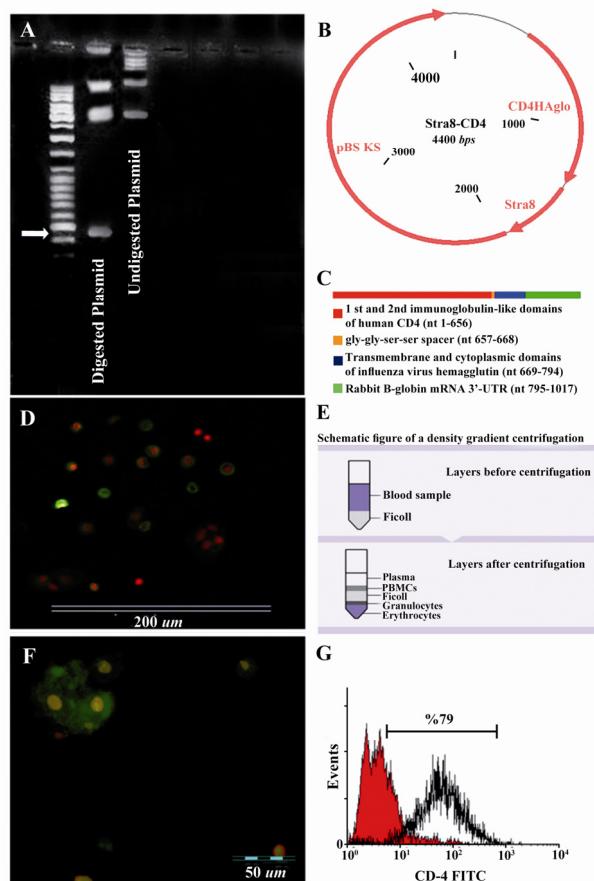


Figure 4. Schematic representation of the Stra8-CD4HLAgo fusion gene, A) RT-PCR analysis 1.4 kb promoter region of mouse Stra8 gene linked to the coding region of human CD4 in positive bacterial colony. B) Schematic representation map of vector. C) Map of CD4HLAgo construct. D) A fluorescent microscopic picture of CD4 positive cells (lymphocyte cells). E) Schematic figure of isolation of human lymphocyte cells as a CD4 positive cells. F) A fluorescent microscopic picture of CD4 positive cells after purification. G) Flow cytometric of purified CD4 positive cells

lowed by transfection and sorting for purification and a co-culture mimicking *in vivo* conditions to obtain SSC-like cells. The heterogeneity of the cells led us to sort them based on the expression of *Stra8*, which is regulated by RA. Adult primary cells are, in general, difficult to transfect with DNA, and BMSCs derived PGC-like cells are no exception. Importantly, for *in vitro* analysis of BMSCs derived PGC-like cells, the introduction of DNA has to be mediated by a technique that does not affect their proliferation and differentiation⁽¹⁸⁾.

Nayernia et al showed that after RA induction (final concentration of $10^{-5} M$), ex-

pression of Stra8-EGFP was detected in approximately 60% of all cells; they observed the expression of Stra8-EGFP in 90% of all cells after twice FACS purification⁽¹⁹⁾. Furthermore, they revealed an indication of permanent activation of the Stra-8 promoter as well as differentiation into male germ cells. Our results showed that the transfection efficiency of lipofectamine 2000 after optimization was approximately 40% using a pCMV-GFP vector as a control, only 22% mBMSC-derived PGC-like cells could express the human CD4 marker. This point is important as the structure of the DNA in ESCs is very loose and could be more easily affected by inductive factors than adult mesenchymal stem cells⁽²⁰⁾. Thus, the source of stem cells used for transfection is the reason for the difference in the results obtained from our study and that from Nayernia et al. However, ESCs raise ethical issues and are more affected by transfection method than BMSCs; these problems have been solved or minimized in our study. Our cells exhibited 80% purity in the expression of *Stra8* gene, which is activated by RA.

In the differentiation step we used inducer cocktail with and without Sertoli cell co-cultures which significantly increased the expression of *Stra8*. Sertoli cells play an essential role in germ cell development *in vivo* and *in vitro*, forming niches for germ cells which allow a certain number of germ cells to reside or repopulate the seminiferous tubules, limiting the expansion of spermatogonial population and providing essential factors for growth/proliferation and differentiation of germ cells into spermatozoa^(21,22). Sertoli cells support the differentiation of germ cells into SSCs by producing BMP-4 and Activin A. Receptors of LIF and bFGF are expressed in normal mature mice SSCs^(23,24). These growth factors as well as SCF could functionally differentiate PGCs into SSCs in gelatin-coated plates^(7,16,17). The combination of SCF and LIF induces the proliferation and viability of PGCs^(25,26). The addition of LIF, RA, and forskolin differentiated PGCs from ESCs⁽²⁷⁾, and the co-culture with Sertoli cells with LIF

and RA led to the formation of germ cell colonies⁽²⁸⁾. bFGF triggers the bFGF-1 receptor, making Sertoli cells to secrete growth factors that in turn affect the PGCs, inducing expression of cell adhesion molecules such as α 6 and β 1 integrins to suppress migration and initiate differentiation⁽²⁷⁾. The expression of *Stra-8* is significantly increased because of the effects of RA.

Our data revealed a significant increase in *Piwil2* expression in Sertoli cells group compared to another groups. *Piwil2* is sharply expressed in spermatids and partially in primary spermatocytes on day 14 post-fertilization and acts as a germ line-sertoli-germ line signaling loop. *Piwil2* knockout mice have defective spermiogenesis initiation⁽²⁹⁻³¹⁾. The up regulation of *Piwil2* occurred with *Nanog* & *Pou5F1* in Sertoli cell group compared to another experimental differentiation groups. Synergistic *Nanog* and *Pou5F1* control each other expression, regulating self renewal in stem cells as well as the development of germ cells⁽³²⁾. *Nanog* is expressed in PGCs until the end of migration to the genital ridge, at which point it is suppressed while differentiation is initiated⁽³³⁾. At the differentiation stage, the expression of *Nanog* was increased because of the inhibition of *P53* bonding to the gene promoter⁽³⁴⁾.

Furthermore, Sertoli cells could secrete Kit-ligand (steel factor)⁽³⁵⁾, which affect the proliferation, and enrichment of PGCs after cultured in media supplemented with 10% fetal bovine serum is estimated to contain 3.6×10^8 M RA⁽³⁶⁾. These cells could release LIF, which is important for the proliferation, enrichment (in a dose-dependent manner), and self-renewal of PGCs⁽²⁸⁾. The increase in the expression of *Piwil2* is attributed to its role in the regulation of self-renewal in germ cells and SSCs^(29,30). On the other hand, Sertoli cells could support the stemness of PGC-like cells without differentiation to SSC-like cells compared to group of Sertoli cells following cocktail. *Piwil2*, *Pou5F1* & *Nanog* were expressed during proliferation of the germ cell development. During the differentiation stage

of PGC-like cells to SSC-like cells, there was a significant decrease in *Piwil2*, *Pou5F1* & *Nanog*. Our results revealed that the germ line-sertoli-germ line signaling loop in the IC+Co group could support differentiation of PGC-like cells to SSC-like cell.

The expression of *Mvh* and *Piwil2* in all of the experimental groups was lower than that in the purified PGC-like cells whereas, increased the expression levels of *Stra-8* occurred toward the end of the study in IC+Co group relative to PGC-like cell. This gene is expressed in the early gonad of both male and female embryos, preventing meiosis in the germ cells when they first arrive at the gonad and come into contact with RA. Moreover, the expression of *Nanog* and *Pou5F1* was significantly lower than that observed in Purified PGC-like cells except Sertoli cell group, but the *c-Myc* expression was significantly increased compared with purified PGC-like cells at the end of study in the all of experimental groups. However, a significantly decrease was observed in the expression of *c-Myc* after purification, while that of *c-Myc* increased at the differentiation step.

Conclusion

Based on our findings, we can conclude that sorting of stem cells derived germ line cells based on promoter of specific gene could be high and efficiently safe method for cell therapy. We can eliminate undifferentiated cells with high expression of *Pou5F1*, *Nanog* and *c-Myc* as pluripotency genes of population cells during differentiation stage. Also, these newly differentiated cells could differentiate with LIF, bFGF, RA addition, and Sertoli cell co-culture cause increase in the expression levels of SSCs specific genes such as *Stra-8*.

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References

1. de Rooij DG. Regulation of spermatogonial stem cell behavior in vivo and in vitro. *Anim Reprod* 2006;3(2):130-134.
2. Anderson A, Copel TK, Scholer H, Heasman J, Wylie C. The onset of germ cell migration in the mouse embryo. *Mech Dev* 2000;91(1-2):61-68.
3. Marques-Mari AI, Lacham-Kaplan O, Medrano JV, Pellicer A, Simón C. Differentiation of germ cells and gametes from stem cells. *Hum Reprod Update* 2009;15(3):379-390.
4. Clark AT, Bodnar MS, Fox M, Abeyta MJ, Firpo MT, Pera RA. Spontaneous differentiation of germ cells from human embryonic stem cells in vitro. *Hum Mol Genet* 2004;13(7):727-739.
5. Hubner K, Fuhrmann G, Christenson LK, Kehler J, Reinbold R, De La Fuente R, et al. Derivation of oocytes from mouse embryonic stem cells. *Science* 2003;300(5623):1251-1256.
6. Toyooka Y, Tsunekawa N, Akasu R, Noce T. Embryonic stem cells can form germ cells in vitro. *Proc Natl Acad Sci USA* 2003;100(20):11457-11462.
7. Nayernia K, Li M, Jaroszynski L, Khusainov R, Wulf G, Schwandt I, et al. Stem cell based therapeutic approach of male infertility by teratocarcinoma derived germ cells. *Hum Mol Genet* 2004;13(14):1451-1460.
8. Kubota H, Brinster RL. Technology insight: in vitro culture of spermatogonial stem cells and their potential therapeutic uses. *Nat Clin Pract Endocrinol Metab* 2006;2(2):99-108.
9. Gardner L, Lee LA, Dang CV. c-myc Protooncogene. In: Bertino JR (eds). *Encyclopedia of Cancer*. San Diego, California: Academic Press; 2002,555-561.
10. Jamous M, Al-Zoubi A, Khabaz MN, Khaledi R, Al Khateeb M, Al-Zoubi Z. Purification of mouse bone marrow-derived stem cells promotes ex vivo neuronal differentiation. *Cell Transplant* 2010;19(2):193-202.
11. Mazaheri Z, Movahedin M, Rahbarizadeh F, Amanpour S. Different doses of bone morphogenetic protein 4 promote the expression of early germ cell-specific gene in bone marrow mesenchymal stem cells. *In Vitro Cell Dev Biol Anim* 2011;47(8):521-525.
12. Giuli G, Tomljenovic A, Labrecque N, Oulad-Abdelghani M, Rassoulzadegan M, Cuzin F. Murine spermatogonial stem cells: targeted transgene expression and purification in an active state. *EMBO Rep* 2002;3(8):753-759.
13. Boroujeni MB, Salehnia M, Valojerdi MR, Mowla SJ, Forouzandeh M, Hajizadeh E. Comparison of gene expression profiles in erythroid-like cells derived from mouse embryonic stem cells differentiated in simple and co-culture systems. *Am J Hematol* 2008;83(2):109-115.
14. Lee JH, Schütte D, Wulf G, Füzesi L, Radzun HJ, Schweyer S, et al. Stem-cell protein Piwil2 is widely expressed in tumors and inhibits apoptosis through activation of Stat3/Bcl-XL pathway. *Hum Mol Genet* 2006;15(2):201-211.
15. Kee K, Gonsalves JM, Clark AT, Pera RA. Bone morphogenetic proteins induce germ cell differentiation from human embryonic stem cells. *Stem Cells Dev* 2006;15(6):831-837.
16. Geijsen N, Horoschak M, Kim K, Gribnau J, Eggan K, Daley GQ. Derivation of embryonic germ cells and male gametes from embryonic stem cells. *Nature* 2004;427(6970):148-154.
17. Nayernia K, Lee JH, Drusenheimer N, Nolte J, Wulf G, Dressel R, et al. Derivation of male germ cells from bone marrow stem cells. *Lab Invest* 2006;86(7):654-663.
18. Esposito M.T. In vitro and in vivo characterization of murine bone marrow stromal stem cells: self renewal, differentiation, tumorigenic potential [Ph.D's thesis]. [Sede di Napoli]: University of Degli Studi Di Napoli "Federico II"; 2009. 103 p.
19. Nayernia K, Nolte J, Michelmann HW, Lee JH, Rathskirchen K, Drusenheimer N, et al. In vitro-differentiated embryonic stem cells give rise to male gametes that can generate offspring mice. *Dev Cell* 2006;11(1):125-132.
20. Ber S. Bone marrow derived adult stem cells: characterization and application in cell therapy [Ph.D's thesis]. [Göttingen]: Georg-August-University of Göttingen; 2006. 77 p.
21. Sofikitis N, Ono K, Yamamoto Y, Papadopoulos H, Miyagawa I. Influence of the male reproductive tract on the reproductive potential of round spermatids abnormally released from the seminiferous epithelium. *Hum Reprod* 1999;14(8):1998-2006.
22. Huleihel M, Lunenfeld E. Regulation of spermatogenesis by paracrine/autocrine testicular factors. *Asian J Androl* 2004;6(3):259-268.
23. Huleihel M, Abuelhija M, Lunenfeld E. In vitro culture of testicular germ cells: Regulatory factors

- and limitations. *Growth Factors* 2007;25(4):236-252.
24. Van der Wee KS, Johnson EW, Dirami G, Dym TM, Hofmann MC. Immunomagnetic isolation and long-term culture of mouse type A spermatogonia. *J Androl* 2001;22(4):696-704.
 25. Creemers LB, den Ouden K, van Pelt AM, de Rooij DG. Maintenance of adult mouse type A spermatogonia in vitro: influence of serum and growth factors and comparison with prepubertal spermatogonial cell culture. *Reproduction* 2002;124(6):791-799.
 26. Matsui Y, Toksoz D, Nishikawa S, Nishikawa SI, Williams D, Zsebo K, et al. Effect of steel factor and leukemia inhibitory factor on murine primordial germ cells in culture. *Nature* 1991;353(6346):750-752.
 27. De Felici M, Dolci S. Leukemia inhibitory factor sustains the survival of mouse primordial germ cells cultured on TM4 feeder layers. *Dev Biol* 1991;147(1):281-284.
 28. Matsui Y, Zsebo K, Hogan BL. Derivation of pluripotential embryonic stem cells from murine primordial germ cells in culture. *Cell* 1992;70(5):841-847.
 29. Cox DN, Chao A, Lin H. Piwi encodes a nucleoplasmic factor whose activity modulates the number and division rate of germline stem cells. *Development* 2000;127(3):503-514.
 30. Cox DN, Chao A, Baker J, Chang L, Qiao D, Lin H. A novel class of evolutionarily conserved genes defined by piwi are essential for stem cell self-renewal. *Genes Dev* 1998;12(23):3715-3727.
 31. Lin H, Spradling AC. A novel group of pumilio mutations affects the asymmetric division of germline stem cells in the *Drosophila* ovary. *Development* 1997;124(12):2463-2476.
 32. Western PS, van den Bergen JA, Miles DC, Sinclair AH. Male fetal germ cell differentiation involves complex repression of the regulatory network controlling pluripotency. *FASEB J* 2010;24(8):3026-3035.
 33. Yamaguchi S, Kurimoto K, Yabuta Y, Sasaki H, Nakatsuji N, Saitou M, et al. Conditional knockdown of Nanog induces apoptotic cell death in mouse migrating primordial germ cells. *Development* 2009;136(23):4011-4020.
 34. Hoffman B, Amanullah A, Shafarenko M, Liebermann DA. The proto-oncogene c-myc in hematopoietic development and leukemogenesis. *Oncogene* 2002;21(21):3414-3421.
 35. Aflatoonian B, Moore H. Human primordial germ cells and embryonic germ cells, and their use in cell therapy. *Curr Opin Biotechnol* 2005;16(5):530-535.
 36. Fuchs E, Green H. Regulation of terminal differentiation of cultured human keratinocytes by vitamin A. *Cell* 1981;25(3):617-625.