



## Targeted Overexpression of NDRG2 using Survivin Promoter Reduces Viability and Invasiveness of A549 Cell Line

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### Abstract

**Background:** Anti-tumor effects of N-myc Downstream Regulated Gene2 (NDRG2) have been demonstrated in many tumors. In the present study, NDRG2 was specifically overexpressed in lung cancer cell line using Survivin Promoter (Sur-P). Then, the effects of NDRG2 overexpression on viability, apoptosis, migration, and invasion of A549 cells were evaluated.

**Methods:** Recombinant pAdenoVator-Sur-P-NDRG2-IRES-GFP plasmid harboring *NDRG2* gene under transcriptional control of Sur-P and mock plasmid were constructed. A549 lung tumor cells and LX-2 cells (non-tumor cell line) were transfected with pAdenoVator-Sur-P-NDRG2-IRES-GFP, pAdenoVator-CMV-NDRG2-IRES-GFP, or mock plasmids. Tumor specificity of Sur-P was evaluated using fluorescent microscopy for GFP expression. The effects of *NDRG2* overexpression on cell viability, apoptosis, and migration of A549 cells were measured using MTT, annexinV/7-AAD flow cytometry, and transwell migration assay, respectively. *NDRG2* and matrix metalloproteinase-2 (*MMP-2*) expression were measured using real time-PCR.

**Results:** pAdenoVator-Sur-P-NDRG2-IRES-GFP transfection resulted in a huge GFP expression in A549 cells, but not in LX-2 cells. The results of real time-PCR analysis also showed that pAdenoVator-Sur-P-NDRG2-IRES-GFP transfection led to an abundant *NDRG2* expression in A549 cells. *NDRG2* overexpression decreased A549 cell viability through increasing cell apoptosis. In addition, migration, invasion, and *MMP-2* expression decreased following *NDRG2* overexpression in A549 cells.

**Conclusion:** The findings indicate that the targeted overexpression of *NDRG2* using Sur-P can reduce the viability and invasiveness of A549 cells, suggesting possible benefits of this approach in lung cancer therapy.

**Keywords:** A549 cells, Apoptosis, Lung cancer, Matrix metalloproteinase 2, Metastasis, Survivin

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### Introduction

According to GLOBOCAN 2020 statistics, lung cancer is the second in incidence (11.4%) and the first in mortality rate (18%) among 36 prevalent cancers worldwide<sup>1</sup>. Histologically, lung cancer is classified as Small-Cell Lung Carcinoma (SCLC) and Non-SCLC (NSCLC), which account for about 15% and 85% of lung cancer cases, respectively<sup>2</sup>. Surgical resection, radiotherapy, chemotherapy, targeted therapy, and combination therapy are available therapeutic strategies for lung cancer. Four classes of chemotherapeutic

drugs are used for the treatment of patients with NSCLC which include alkylating agents (cisplatin and carboplatin) microtubule-targeting drugs (paclitaxel and docetaxel), antimetabolites (pemetrexed and gemcitabine), and topoisomerase inhibitors (etoposide)<sup>3</sup>. Although chemotherapy still plays an important role in the treatment of patients with lung cancer, drug toxicity and resistance are the main challenges in this issue. Previous studies have revealed the role several mechanisms in developing drug resistance including reduced

drug uptake to tumor cells, increased drug efflux from the tumor cells, genetics and epigenetic modifications, apoptosis dysfunction, and acquisition of cancer stem cell-like properties<sup>4</sup>. Therefore, finding an efficient and safe therapeutic method that specifically targets cancer cells has been the focus of many studies. In this context, gene therapy has emerged as a promising approach<sup>5</sup>.

Tumor Specific Promoters (TSPs), transcriptional regulatory elements that ensure targeted and efficient expression of genes in tumor cells, are key elements in the success of gene therapeutic approaches<sup>6</sup>. Survivin, an apoptosis inhibitory protein, is expressed in numerous types of cancer including SCLC and NSCLC<sup>7</sup>. Numerous studies have revealed tumor specificity and strength of Survivin promoter (Sur-P) in the expression of various genes in several tumor cells<sup>8</sup>. Moreover, its transcriptional activity is increased in hypoxia, a condition that usually found in tumor cells<sup>9</sup>. Specific overexpression of SP1 protein, a transcription factor that directly binds to Sur-P, results in high transcriptional activity of Sur-P in lung tumor cells<sup>10</sup>.

N-myc Downstream Regulated Gene 2 (*NDRG2*) is a multifunctional protein with crucial roles in the regulation of cell growth and differentiation. *NDRG2* expression is downregulated in many cancers including lung cancer. Furthermore, its expression has been found to be inversely associated with tumor stage, tumor size, and invasion of tumor cells to circulation and directly associated with patient's survival time<sup>11,12</sup>. Moreover, Faraji *et al* showed that *NDRG2* upregulation in A549 cells suppressed cell viability and invasion of tumor cells through decreasing expressions of Matrix Metalloproteinases-2 and -9 (*MMP-2* and *MMP-9*)<sup>13</sup>.

The present study aimed to construct a recombinant plasmid harboring *NDRG2* cDNA under the transcriptional control of Sur-P to evaluate the tumor specificity and strength of this promoter in enhancing the *NDRG2* expression. The results revealed specific overexpression of *NDRG2* in A549 cell lines using this plasmid. Furthermore, *NDRG2* overexpression was associated with increased apoptosis and decreased invasiveness of the tumor cells.

## Materials and Methods

### Construction of plasmids

Three types of recombinant plasmids were used in this study including pAdenoVator-CMV-*NDRG2*-IRES-GFP (*NDRG2* under transcriptional control of CMV promoter), pAdenoVator-Sur-P-*NDRG2*-IRES-GFP (*NDRG2* under transcriptional control of Sur-P), and pAdenoVator-Sur-P-IRES-GFP (Mock plasmid). CMV-*NDRG2* was constructed in our previous studies<sup>24,26</sup>. To construct Sur-P-*NDRG2* plasmid, Sur-P<sup>14</sup> was amplified from genomic DNA (prepared from peripheral blood mononuclear cells) using forward (5'-CCG GAGTCGACGTTCTTTGAAAGCAG-3'; contained

SalI restriction site) and reverse (5'-TTAGCTCTAGA CAACGTCGG GGCACC-3'; contained XbaI restriction site) primers. Sur-P fragment was digested by XbaI and SalI restriction enzymes (Thermo Scientific, USA) and cloned into the pAdenoVator-IRES-GFP vector using T4 DNA ligase (Thermo Scientific, USA). The ligation mixture was then transformed into competent *Escherichia coli* (*E. coli*) DH5 $\alpha$ . The formation of Sur-P-*NDRG2* was validated using colony PCR and sequencing.

### Cells and cell culture conditions

A549 cell line (ATCC ref. No. CLL-185) was purchased from Pasture Institute, Iran. LX-2 (human hepatic stellate cell line) was kindly provided by Dr. Younes Hoseini. A549 cells and LX-2 cells were cultured in RPMI 1640 (Gibco/Invitrogen, Carlsbad, CA) supplemented with 10% and 5% Fetal Bovine Serum (FBS) (Gibco, USA) and 1% penicillin-streptomycin (Gibco, USA). Both cell lines were incubated at 37°C with 5% CO<sub>2</sub> atmosphere.

### Overexpression of the *NDRG2* gene in A549 cells

To evaluate tumor specific activity of Sur-P, A549 and LX-2 cell lines were transfected with pAdenoVator-CMV-*NDRG2*-IRES-GFP, pAdenoVator-Sur-P-*NDRG2*-IRES-GFP, and mock plasmids using lipofectamine™ 2000 (Invitrogen, Carlsbad, CA) according to the manufacturer's instructions. After 48 hr, the transfected cells were detached using 0.25% Trypsin-EDTA, washed, and resuspended in cold PBS buffer. Fluorescence microscopy and flow cytometry analyses were used to monitor the cellular expression of GFP and to measure the efficiency of transfection. Fluorescence microscopy was performed on an inverted microscope (Hund, Germany) with filter sets designed for GFP. Flow cytometric analysis was conducted using BD FACSCalibur flow cytometer (BD Biosciences, USA). For each sample, 10,000 events were collected. The expression of *NDRG2* was evaluated using real-time PCR as described in the following section.

### Cell viability assay using the MTT method

The effect of *NDRG2* overexpression on cell viability was evaluated using MTT assay. Briefly, 6×10<sup>3</sup> A549 cells were seeded on each well of a 96-well plate and were transfected with either pAdenoVator-Sur-P-*NDRG2*-IRES-GFP plasmid (*NDRG2* group) or mock plasmid (pAdenoVator-Sur-P-IRES-GFP) using Lipofectamine 2000 (Invitrogen, USA). Forty-eight hours after transfection, the cells were treated with 10  $\mu$ l of MTT [3-(4,5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide] solution followed by a four-hour incubation. Afterwards, the media were removed, formazan crystals were dissolved by Dimethyl Sulfoxide (DMSO), and the absorbance was measured at 545 nm.

### Cell apoptosis assay

Annexin V/7AAD flow cytometry was used to evaluate the effect of *NDRG2* overexpression on cell apoptosis. In doing so, A549 cells were transfected with

NDRG2 or mock plasmids, as described previously. Forty-eight *hr* after transfection, the cells were harvested using trypsin digestion (0.25%) and centrifugation. The cells were then washed with ice-cold PBS, suspended in 1× annexin V binding buffer, and stained with PE annexin V and 7-AAD at room temperature in the dark for 15 *min*. The percentages of live and apoptotic cells were determined using the FACSCalibur instrument (BD Bioscience, USA) and flowjo software analysis.

**Migration and invasion assay**

Transwell migration assay was applied using 24-well transwell insert and 8 μm pore filters (BD Bioscience, Bedford, MA) to evaluate the effects of *NDRG2* overexpression on the migration of the transfected A549 cells. Briefly, 48 *hr* after transfection, A549 cells (5×10<sup>4</sup>) were suspended with a serum-free medium, kept serum starved for 24 *hr*, transferred to the upper part of the transwell, and incubated at 37°C. The cells were then allowed to migrate to the lower part of the transwell, which was filled with RPMI supplemented with 10% FBS. After 24 *hr* incubation, the cells that migrated to the lower surface of the transwell were fixed with methanol for five minutes and were stained with 0.1% crystal violet. For invasion assays, the transwell was coated with 100 μl (1 mg/ml) matrigel (BD Bioscience, Bedford, MA). Forty-eight *hr* after transfection, A549 cells (5×10<sup>4</sup>) were plated onto the upper part of the matrigel-coated transwell chamber and were incubated for 48 *hr*. The invaded cells were then fixed with methanol, stained, and counted. The number of invading or migrating cells was determined by counting five fields (400×) of a light microscope on each membrane and was calculated as the mean number of cells per field<sup>15,16</sup>.

**Real time polymerase chain reaction**

A549 and LX-2 cell lines were cultured at 1×10<sup>5</sup> and 1.5×10<sup>5</sup> counts in six-well plates, respectively. Both cell lines were transfected with pAdenoVator-Sur-P-NDRG2-IRES-GFP. Forty-eight *hr* after transfection, the cells were exposed to 1 ml TRIzol (Thermo Fisher Scientific, USA) per 1×10<sup>6</sup> cells and the RNAs were extracted according to the manufacturer’s protocol. After that, cDNAs were synthesized from the extracted RNAs using Takara kit according to the manufacturer’s instructions. Quantitative real-time PCR was performed using specific primers (Table 1), SYBR-

Green master mix, and the Rotor-Gene Q (Qiagen, Germany) in a total reaction volume of 20 μl.

**Statistical analysis**

All statistical analyses were performed using the SPSS software, version 15.0. Statistical differences between the study groups were analyzed using Mann-Whitney or Kruskal-Wallis test. The data represented mean ± Standard Deviation (SD) of at least three independent experiments. The p<0.05 was considered statistically significant.

**Results**

**Tumor specificity of survivin promoter**

Tumor specific activity of Sur-P was evaluated using fluorescence microscopy (Figure 1 A-D). As illustrated in figure 1A, transfection of pAdenoVator-CMV-NDRG2-IRES-GFP plasmid into normal LX-2 cells led to a high *GFP* expression (Figure 1A), while transfection with pAdenoVator-Sur-P-NDRG2-IRES-GFP did not lead to *GFP* expression (Figure 1B). In A549 cells, however, transfection with both pAdenoVator-CMV-NDRG2-IRES-GFP (Figure 1C) and pAd-

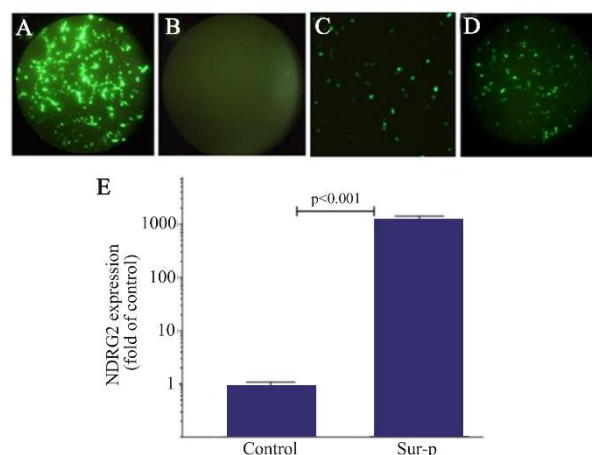


Figure 1. Tumor specificity of Green Fluorescent Protein (*GFP*) expression under Sur-P control. *GFP* was expressed in the LX-2 cells transfected with pAdenoVator-CMV-NDRG2-IRES-GFP (A), but not in those transfected with pAdenoVator-Sur-P-NDRG2-IRES-GFP(B). *GFP* was also expressed in the A549 cells transfected with both pAdenoVator-CMV-NDRG2-IRES-GFP (C) and pAdenoVator-Sur-P-NDRG2-IRES-GFP (D). Real-time PCR data (E) of *NDRG2* expression in the A549 cells transfected with the pAdenoVator-Sur-P-NDRG2-IRES-GFP plasmid and control non-transfected cells. The data represent the mean±SD of at least three independent experiments.

Table 1. The primer sequences for the real-time PCR analysis of *NDRG2* and *MMP-2* gene expressions

Primer	Sequence	Product length (bp)
NDRG2 (forward)	5'-CCTCACCTCTCCATTCC-3'	188
NDRG2 (reverse)	5'-TATCACCTCCACGCTCAA-3'	
MMP2 (forward)	5'-TGGAGATACAATGAGGTGAAGAAG-3'	261
MMP2 (reverse)	5'-GAAGGCAGTGGAGAGGAAG-3'	
B-actin (forward)	5'-GCACAGAGCCTCGCCTTT-3'	230
B-actin (reverse)	5'-GCCTCGTCGCCACATAG-3'	

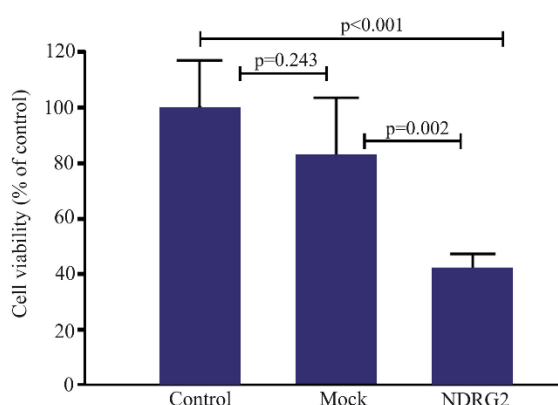


Figure 2. MTT assay results showing the effects of *NDRG2* overexpression on A549 cell viability. A549 cells were transfected with pAdenoVator-Sur-p-*NDRG2*-IRES-GFP or mock plasmids and cell viability was evaluated using the MTT assay. The data were analyzed using Kruskal-Wallis test. The data represent the mean±SD of at least three independent experiments.

enoVator-Sur-P-*NDRG2*-IRES-GFP (Figure 1D) led to *GFP* expression, suggesting tumor specificity of Sur-P. The results of real-time PCR also revealed an abundant *NDRG2* expression in the A549 cells (1200-fold) transfected with the pAdenoVator-Sur-P-*NDRG2*-IRES-GFP plasmid compared to the control non-transfected cells, suggesting the high strength of Sur-P in inducing *NDRG2* expression in A549 tumor cells.

**Cell viability assay**

Figure 2 shows the results of MTT assay. The data revealed that *NDRG2* overexpression decreases the viability of A549 cells compared to the control non-transfected cells ( $p<0.001$ ) and the mock plasmid-transfected group ( $p=0.002$ ). No significant difference was observed between the viability of control and mock groups ( $p=0.243$ ).

**Flow cytometric analysis of apoptosis**

As shown in figure 3, *NDRG2* overexpression increased the percentage of total apoptotic cells compared to the mock-transfected ( $p=0.047$ ) and control non-transfected cells ( $p=0.039$ ). However, a decrease was detected in cell viability following *NDRG2* overexpression compared to the mock and control non-transfected cells.

**Migration and invasion assay**

Figure 4 illustrates the effects of *NDRG2* overexpression on the migration and invasion of the A549 cells. Based on the data presented in figure 4, *NDRG2* overexpression reduced the number of migrated and invaded cells compared to the mock and control cells, suggesting the anti-metastatic effects of *NDRG2* on A549 cells.

**The effect of *NDRG2* overexpression on the level of MMP-2 mRNA**

The effect of *NDRG2* overexpression on *MMP-2* expression was evaluated using real-time PCR. The

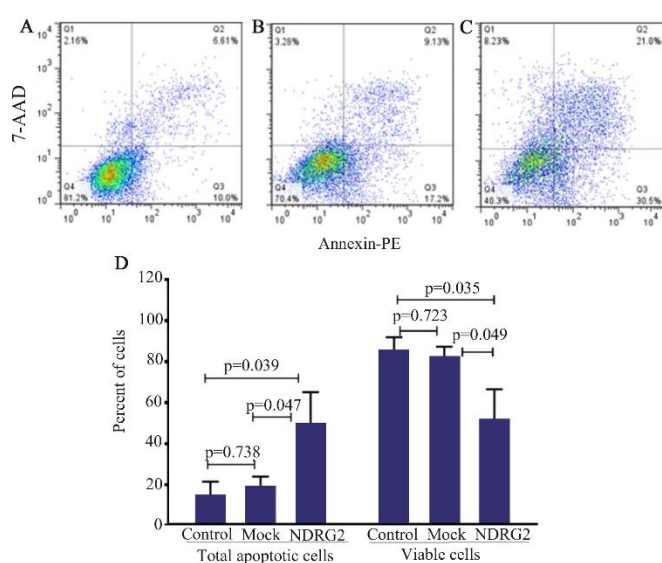


Figure 3. The effect of *NDRG2* overexpression on the percentage of live and total apoptotic A549 cells. The charts show the percentages of live cells (Q4), early apoptotic cells (Q3), late apoptotic cells (Q2), and necrotic cells (Q1) in the control (A), mock (B), and *NDRG2* (C) groups. The data (D) represent the mean±SD of three independent experiments. The data were analyzed using Kruskal-Wallis's test.

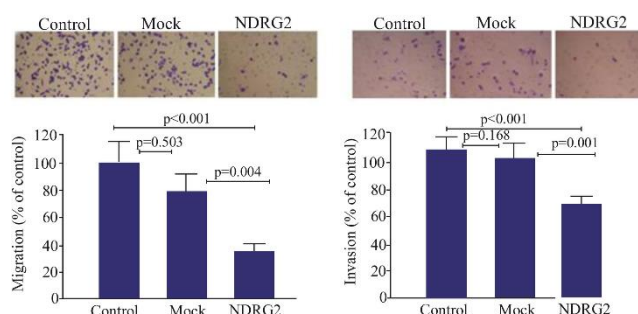


Figure 4. The effects of *NDRG2* overexpression on the migration and invasion of A549 cells using transwell assay. The percentage of migrated and invaded A549 cells in the mock plasmid transfected (mock), pAdenoVator-Sur-P-*NDRG2*-IRES-GFP-transfected (*NDRG2*), and control non-transfected cells. The data represent the mean±SD of three independent experiments. The data were analyzed using Kruskal-Wallis test.

results revealed a significant decrease in the *MMP-2* mRNA level ( $p=0.045$ ) compared to the control non-transfected cells (Figure 5).

**Discussion**

Several studies have revealed the beneficial effects of various gene therapy methods including inhibition of oncogenes and replacement of tumor-suppressor genes in the treatment of lung cancer <sup>5</sup>. However, lack of tumor specificity is the most important limitation of these approaches. Two main strategies have suggested for increasing tumor specificity of gene therapy including targeted transfer to tumor cells *via* tumor specific

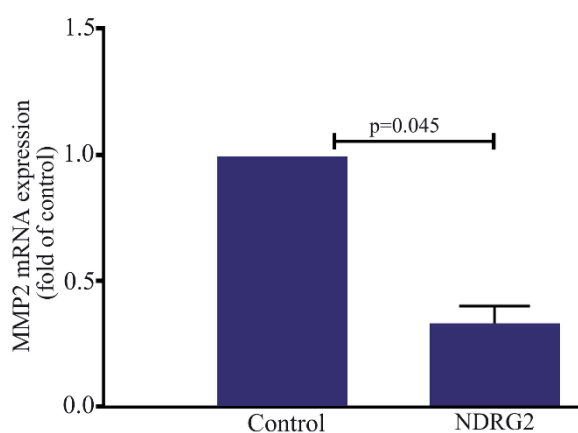


Figure 5. *NDRG2* overexpression reduced *MMP-2* expression in the A549 cell line. The data represent the mean $\pm$ SD of at least three independent experiments. The data were analyzed using Mann-Whitney test.

\* $p < 0.05$  was considered statistically significant

receptors and targeted transcription of anti-tumor genes using TSPs. Despite the tumor specificity of various TSPs, low transcriptional efficiency usually hinders their utilization for gene therapy strategies. The present study assessed the specificity and efficiency of Sur-P in the overexpression of *NDRG2* in A549 cells. The findings revealed an abundant expression of *NDRG2* under the transcriptional control of Sur-P in the lung cancer cell line, but not in the normal cell line, compared to the CMV promoter that enhanced *NDRG2* overexpression in both normal and tumor cells. Furthermore, *NDRG2* overexpression decreased cell viability and invasiveness of A549 cells, suggesting specificity and efficiency of this strategy and its possible effectiveness in the treatment of NSCLC.

Survivin is anti-apoptotic protein that is specifically expressed in the most of human cancers including lung cancer. Numerous studies have revealed that efficient downregulation of survivin using transcriptional and post-translational approaches resulted in death and apoptosis of tumor cells and sensitized tumor cells to chemotherapeutic regimens<sup>17</sup>. In addition, several studies have described the beneficial use of Sur-P for the targeted expression of anti-tumor genes<sup>18</sup>. Bao *et al* used a 1092-bp fragment of the human Sur-P to control the expression of alkaline phosphatase in several tumor cell lines and reported its promoter activity in all evaluated tumor cells<sup>19</sup>. However, the large size of the fragment hampered its use for the construction of recombinant plasmids<sup>8</sup>. Therefore, Yang *et al* evaluated the efficiency of a number of surviving promoter fragments and indicated that a 269-bp fragment of Sur-P was sufficient for the specific and efficient expression of genes in human pancreatic and breast cancer cell lines<sup>14</sup>. In the current study, we constructed a plasmid harboring Sur-P and human *NDRG2* cDNA. Our data revealed that Sur-P can mediate *NDRG2* overexpres-

sion in A549 cells, but not in LX-2 cells, suggesting the tumor specificity and strength of this promoter in lung cancer cells. LX-2 has been used in some studies to determine tumor specificity of some promoters<sup>20,21</sup>. Nonetheless, LX-2 is a hepatic stellate cell line and the lack of evaluation of Sur-P activity in the lung epithelial cells is among the limitations of this study. Further studies are needed to determine transcriptional activity of Sur-P in the normal lung epithelial cells.

Previous studies have shown that suppressing expression of *NDRG2* by genetic and epigenetic methods led to development of a variety of tumors including lung cancer. Li *et al* showed a decreased level of *NDRG2* in human lung cancer tissues that was associated with the invasion and tumor size. In addition, 5-years survival time of patients with low *NDRG2* expression was shorter than those with high *NDRG2* expression<sup>11</sup>. Thus, *NDRG2* has become a target for cancer therapy, and numerous studies have revealed the potent effect of *NDRG2* in reducing cell viability and invasiveness of tumor cells<sup>22-26</sup>. Nevertheless, the role *NDRG2* overexpression in lung cancer therapy has been investigated in a few studies<sup>13</sup>. In the present study, human *NDRG2* gene was specifically overexpressed using Sur-P in A549 cells. Consistent with the results of a previous study<sup>13,27</sup>, our findings revealed that *NDRG2* overexpression decreased viability and enhanced apoptosis of A549 cells. Several possible mechanisms can describe anti-tumor effects of *NDRG2*. In human colon carcinoma cells, it has been demonstrated that *NDRG2* overexpression arrested cell cycle in G1 phase by downregulating cyclin D expression<sup>28</sup>. Similarly, Kang *et al* evaluated the effects of *NDRG2* overexpression on cell proliferation of ovarian cancer cells and showed that *NDRG2* can suppress cell viability by increasing expression of apoptosis-related proteins. In addition, the authors revealed that *NDRG2* overexpression significantly arrested cell cycle of ovarian cancer cell lines by reducing expression of cyclins<sup>29</sup>. To our knowledge, the mechanisms of *NDRG2* effects on the cell viability of lung tumor cells have not been investigated so far and further studies are needed to clarify the possible role cell cycle and apoptosis proteins in this issue.

The results of the current research also showed that *NDRG2* overexpression remarkably reduced the invasion and migration of A549 cells, which was in agreement with the effects of *NDRG2* on other tumor cells<sup>23-25</sup>. MMPs are essential enzymes for the growth, invasion, and angiogenesis of lung tumor cells<sup>30</sup>. *MMP-2* (gelatinase A) is highly expressed in lung cancer and is correlated with the invasiveness of this malignancy<sup>31</sup>. Findings of the current study revealed a decreased level of *MMP-2* mRNA upon *NDRG2* overexpression, indicating the possible role of this enzyme in the anti-invasive effects of *NDRG2*. This finding is line with the results of our previous study which showed the inhibitory effects of *NDRG2* on *MMP-2* enzymatic activ-

ity<sup>13</sup>. Similarly, *NDRG2* overexpression was shown to be associated with MMPs downregulation in renal cell carcinoma<sup>32</sup>, prostate cancer<sup>24</sup>, and melanoma cell lines<sup>33</sup>. Several mechanisms may involve in suppressing effects of *NDRG2* on MMPs expression, notably inhibition of nuclear factor kappa B activity<sup>33</sup> and phosphoinositide 3-kinase (PI3K)/AKT signaling pathway<sup>34</sup>. Further investigations are required to determine the role these signaling pathways in the anti-metastatic effects of *NDRG2* in lung cancer.

### Conclusion

The study findings suggested that Sur-P resulted in the specific overexpression of *NDRG2* in lung tumor A549 cells. The results also exhibited increased cell death and apoptosis and decreased migration and invasion of A549 cells through inhibition of *MMP-2* and *MMP-9* expression. This strategy may be considered for the treatment of NSCLC. Yet, further *in vivo* studies are required to confirm the potential efficacy of this method in lung cancer therapy.

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