



## Fabrication of Calcium Sulfate Coated Selenium Nanoparticles and Corresponding *In-Vitro* Cytotoxicity Effects Against 4T1 Breast Cancer Cell Line

Elnaz Faghfuri<sup>1†</sup>, Ramak Ajideh<sup>1†</sup>, Faranak Shahverdi<sup>2</sup>, Mina Hosseini<sup>3</sup>, Faranak Mavandadnejad<sup>3</sup>,  
Mohammad Hossein Yazdi<sup>2,3</sup>, and Ahmad Reza Shahverdi<sup>1,2,3\*</sup>

1. Biotechnology Research Center, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran

2. Recombinant Vaccine Research Center, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran

3. Department of Pharmaceutical Biotechnology, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran

†The first and the second authors have had equal contribution to this manuscript

### Abstract

**Background:** The inhibitory effect of selenium nanoparticles (SeNPs) on cancer cells has been reported in many studies. In this study, the purpose was to compare the *in vitro* effects of SeNPs and calcium sulfate coated selenium nanoparticles (CaSO<sub>4</sub>@SeNPs) on breast cancer cells.

**Methods:** CaSO<sub>4</sub>@SeNPs and SeNPs were chemically synthesized and characterized with Field Emission Scanning Electron Microscope (FESEM) and energy-dispersive X-ray spectroscopy (EDX). By applying MTT assay, the cytotoxicity effect of both nano-materials on the 4T1 cancer cells was investigated.

**Results:** While LD<sub>50</sub> of SeNPs on 4T1 cancer cells was 80  $\mu$ g, the LD<sub>50</sub> of CaSO<sub>4</sub>@SeNPs was reported to be only 15  $\mu$ g. The difference between the inhibition rates obtained for SeNPs and CaSO<sub>4</sub>@SeNPs was statistically significant (p=0.05). In addition, at higher concentrations (50  $\mu$ g) of CaSO<sub>4</sub>@SeNPs, the cytotoxicity was 100% more than SeNPs alone.

**Conclusion:** According to the result of the present work, it can be concluded that decoration of SeNPs with calcium sulfate leads to an increase in potency by decreasing the effective dose. This effect can be attributed to activation of intrinsic apoptosis signaling and/or pH regulatory properties of CaSO<sub>4</sub>@SeNPs. However, further studies are still needed to determine the exact corresponding mechanisms of this synergistic effect.

*Avicenna J Med Biotech 2021; 13(4): 201-206*

**Keywords:** Apoptosis, Breast neoplasms, Calcium sulfate, Nanoparticles, Selenium

### Introduction

Besides conventional treatments such as chemotherapy and radiotherapy and the latest breakthroughs in molecular biology and immunotherapy in cancer treatment, breast cancer is the most common type of female cancer worldwide representing nearly a quarter (23%) of all cancers in women and a significant challenge to public health<sup>1</sup>. Despite significant progresses in the management of preventive or therapeutic modalities to control this cancer type, the search for a curative treatment is still ongoing<sup>2-4</sup>.

Selenium (Se), a vital micronutrient with proven benefits for human biology, is obtained from dietary sources. This trace element has many important biological effects, particularly chemo preventative and therapeutic properties<sup>5,6</sup>. Small amounts of selenium are vital for specific biological functions in humans. As it

is an integral part of glutathione peroxidase and thio-reductase enzyme, it is capable of scavenging free radicals and regulating the function of the thyroid gland. Additionally, it has been found to be involved in fertility improvement and ensures proper functioning of the immune system<sup>7-9</sup>. The use of elemental selenium, an insoluble metalloid compound produced at nano-scale, chemically or biologically is gaining a great deal of attention due to low therapeutic index of organic and inorganic selenium<sup>10,11</sup>. Various studies have shown that Se supplementation reduces the incidence of prostate, lung, and colon cancers<sup>11-13</sup>. Selenium Nanoparticles (SeNPs) are known as a metalloid form of Se species which have been reported as potential cancer therapeutic agents and drug carriers<sup>14-17</sup>.

In our previous researches, the indirect effect of

SeNPs on augmentation of immune responses against breast cancer tumor cells and enhancing the lifespan of tumor bearing animals was demonstrated<sup>18</sup>. The mechanism of apoptosis induction has attracted researchers' attention for fighting against cancer<sup>19-21</sup>. On the other hand, applying a non-vehicular system for delivering apoptogenic agents to cancer cells to induce apoptosis has been investigated<sup>20</sup>. Moreover, the direct cytotoxicity of SeNPs with different surface modification forms has been reported against different cell lines in recent years<sup>22,23</sup>. For instance, in our recent study, FA@SeNPs, a nanocomposite consisting of SeNPs and folic acid, indicated a considerable potential to target cancer cells<sup>22</sup>. On the other hand, calcium has been remarked for anti-cancer therapy because of its availability, low cost, safety, outstanding biocompatibility, pH-sensitivity, and slow biodegradability<sup>20,24,25</sup>. Calcium could have toxic effect on cancer cells through triggering apoptosis by increasing the pH of tumor microenvironment that can sensitize the drug resistant cancer cells<sup>26</sup>.

There are always risks and adverse effects of administering chemicals, drugs, and medicine *via* nano carriers. In this study, the purpose was to reduce such risks regarding the properties of calcium and also its possible synergistic effect with SeNPs to eradicate cancer cells. Additionally, the cytotoxicity of CaSO<sub>4</sub>@SeNPs on breast cancer cells was highlighted.

## Materials and Methods

### Fabrication of SeNPs and CaSO<sub>4</sub>@SeNPs

One-step method with some modifications was used to synthesize calcium sulfate coated selenium nanoparticles (CaSO<sub>4</sub>@SeNPs) according to the previous report<sup>27</sup>. In the first step, by applying a well-known method, SeNPs were synthesized<sup>22</sup>. For this purpose, 4.8 ml of ascorbic acid in aqueous solution (50 mM) was gradually added to the 100 ml selenium dioxide solution (50 mM) by continuous stirring (300 rpm). At that point, the mixture was centrifuged and washed three times with double-distilled water. For preparing CaSO<sub>4</sub>@SeNPs, plain SeNPs (160 mg) were re-suspended in 50 ml sodium sulfate solution (25 mM). In the next step, 20 ml calcium chloride (350 mM) was added drop by drop into the mixture with continuous stirring (300 rpm). Finally, the reaction mixture was centrifuged and washed three times with double-distilled water. Stock suspension of SeNPs and CaSO<sub>4</sub>@SeNPs was prepared (10 mg/ml) and used for additional cytotoxicity assays.

### Characterization of prepared NPs

To observe the NPs' surface features and determine the elemental composition, a Field Emission Scanning Electron Microscope (FESEM) equipped with Energy-Dispersive X-ray Spectroscopy (EDS) was used. For FESEM observation, NPs were mounted on specimen stubs and coated with gold. Samples were analyzed with MIRA 3 FESEM (TESCAN MIRA3, USA) operated

at 15 kV, and EDS was recorded by focusing on a cluster of NPs.

### Cell culture

The 4T1 breast cancer cell line (Product code: ATCC CRL-2539) was purchased from the National Cell Bank of Iran (NCBI), Pasteur Institute of Iran, Tehran, Iran. The RPMI 1640 medium with 10% Fetal Bovine Serum (FBS), 100 u/ml penicillin, and 100 µg/ml of streptomycin was prepared. The 4T1 cancer cells were cultured in this medium in the condition of 5% CO<sub>2</sub> at 37°C for 1 week. For the cytotoxicity assay, the serum starvation condition was applied by using fetal bovine serum-free RPMI 1640 medium containing the antibiotics mentioned above and the cells were re-incubated at 37°C for 18 hr (5% CO<sub>2</sub>).

### Cell cytotoxicity assay

To evaluate cell cytotoxicity, thiazolyl blue tetrazolium bromide (MTT) assay was used and 2×10<sup>4</sup> starved cells were seeded in 96 well plates and incubated with 12.5, 25, 50, 100, and 200 µg of CaSO<sub>4</sub>@SeNPs and SeNPs for 24 hr. Thereupon, 15 µl MTT (0.5 mg/ml) was added to each well, and the plates were incubated for 2 hr. After that, the medium was replaced with 200 µl of Dimethyl Sulfoxide (DMSO), and the absorbance was recorded at 570 nm<sup>28</sup>. Each experiment was repeated three times, and the MTT assay was performed in three replicates for each experiment. The percentage of viable cells was measured using the following equation, Viable cells (%)=[OD<sub>exp</sub>/OD<sub>con</sub>]×100, where OD<sub>exp</sub> and OD<sub>con</sub> are the optical densities of the treated and untreated cells, respectively. Finally, the cytotoxicity bar histogram was obtained by plotting the percentage of viable cells against the concentration of cytotoxic compounds.

### Statistical analysis

The Holm-Sidak multiple comparison test was used for statistical analysis followed by one-way analysis of variance (ANOVA). Shapiro-Wilk test was used to examine normal distribution of data and Levene's test showed the homogeneity of the variances between the compared groups. The values are reported as mean±Standard Deviation (SD), and a p-value of less than 0.05 is considered statistically significant.

## Results

### Preparation and characterization of SeNPs and CaSO<sub>4</sub>@SeNPs

SeNPs were chemically synthesized and their shape and size were confirmed by FESEM, as shown in figure 1A. The FESEM image showed that the particles size range was below 100 nm, with an average size of about 60 nm. In addition, the FESEM image of the prepared CaSO<sub>4</sub>@SeNPs in figure 1B indicated that the generated nanocomposites were rod shaped (400×100 nm). The EDS spectra of the SeNPs and CaSO<sub>4</sub>@SeNPs are demonstrated in figures 2 and 3, respectively. It reveals the presence of Se element peaks.

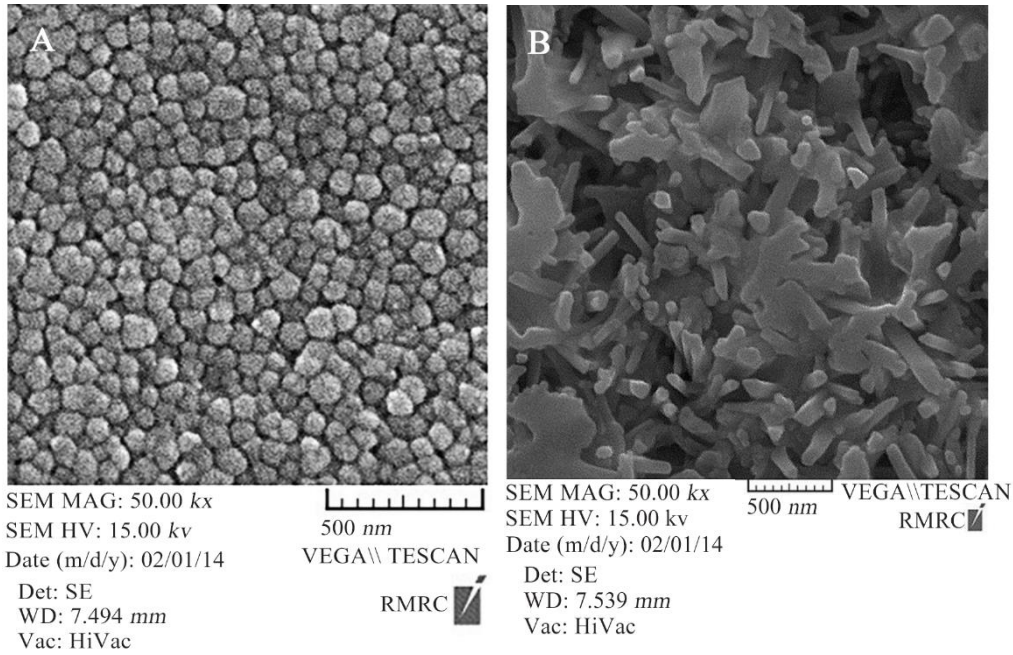


Figure 1. FSEM images of SeNPs, A) FESEM image of plain SeNPs showing the size of 60 nm. B) FESEM image of fabricated CaSO<sub>4</sub>@SeNPs demonstrating nanocomposites are rod shaped (400×100 nm).

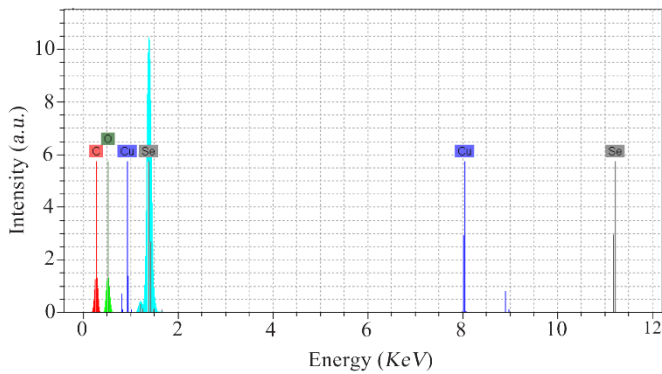


Figure 2. The EDS spectrum of SeNPs confirming the presence of Se atoms and the existence of SeNPs. Additional peaks of copper and carbon elements are attributed to the grid used for FESEM imaging.

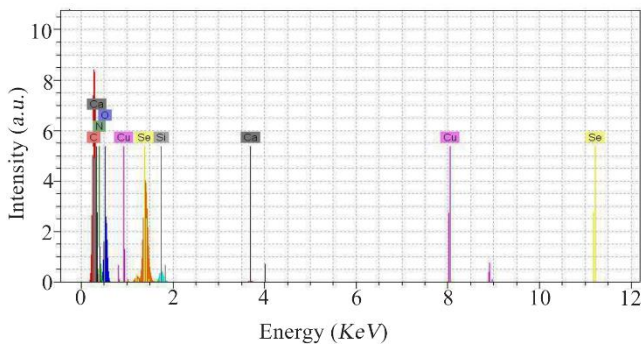


Figure 3. The illustration shows the presence of Se atoms and the existence of elemental NPs in the EDS spectrum of CaSO<sub>4</sub>@SeNPs. Additional peaks of copper, carbon, oxygen, and nitrogen elements are attributed to the grid used for FESEM imaging or calcium compound.

These EDS peaks confirm the existence of elemental SeNPs in both types of prepared NPs. Additional peaks related to calcium, copper, carbon, oxygen, and nitrogen elements are attributes of the grid used for ESEM imaging or calcium sulfate substance.

**Cytotoxicity effects**

Cytotoxicity effects of both CaSO<sub>4</sub>@SeNPs and SeNPs were evaluated *in vitro* at different concentrations (10, 25, 50, 100, and 200) on 4T1 breast cancer cells. The inhibition rate of CaSO<sub>4</sub>@SeNPs increased at higher concentrations which was shown by cytotoxicity analysis of different doses with the aid of statistical data analysis (p<0.05) (Figure 4). The considerable concentrations that led to 50% cell death (IC<sub>50</sub>) were reported to be approximately 80 μg in the cells treated with SeNPs. On the other hand, the CaSO<sub>4</sub>@SeNPs caused the same effect at the concentration of 15 μg. In this study, the possible combinatorial effect of CaSO<sub>4</sub> with SeNPs to kill breast cancer cells was significant at the doses of 25 and 50 μg. In other words, although the doses of SeNPs should be 25 or 50 μg to show 40% inhibitory effect, equal doses of CaSO<sub>4</sub>@SeNPs show two-fold higher impact on killing cancer cells (Nearly 80% of IR). In 100 μg and 200 μg doses of SeNPs and CaSO<sub>4</sub>@SeNPs, the combinatorial effect decreased, which may be due to particle agglomeration at higher concentrations of SeNPs.

**Discussion**

Previous researches demonstrated that different types of selenium have various benefits for human



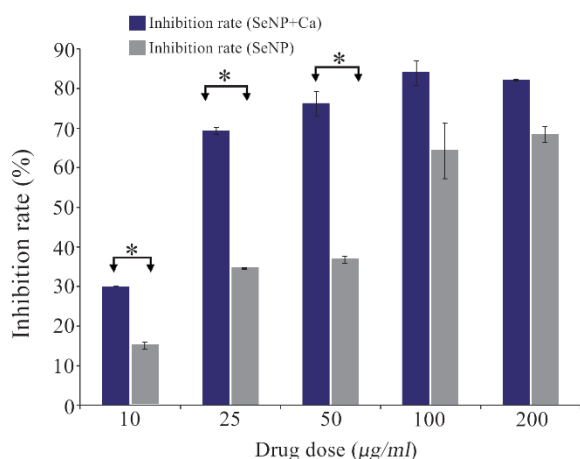


Figure 4. Comparison of cytotoxicity effect of  $\text{CaSO}_4\text{@SeNPs}$  and SeNPs on 4T1 cells. The mean value of three repeats is presented and the standard deviation was negligible (<5%), yet the difference between the inhibition rates obtained for SeNPs and  $\text{CaSO}_4\text{@SeNPs}$  at concentrations of 10, 25 and 50  $\mu\text{g}$  was statistically significant ( $p=0.05$ ).

body<sup>29</sup>. The biological impacts of Se mostly depend on the incorporation of this metalloid into seleno-proteins in the form of seleno-cysteine amino acid<sup>30</sup>. However, many past and recent clinical trials are trying to use selenium and its related compounds as a chemopreventive agent to control the incidence of cancers. The trials have had controversial results and some rejected the prophylactic role<sup>29</sup>. On the other hand, in spite of a bunch of clinical and laboratory data, the mechanism of selenium activity is not fully determined for cancer treatment. One possible beneficial effect of selenium is related to its indirect effect on immune responses, as the consumption of selenium in a therapeutic regimen in addition to conventional anticancer treatments seems more efficient due to the immune stimulatory effect of this element. However, several researches support the direct anti-cancer effect of selenium as a result of anti-oxidant capacity of this trace element. However, like other trace elements, a major concern which limits selenium application is the toxicity.

Beside the anti-oxidant effect, SeNPs are also able to kill the cancer cells through the apoptosis mechanism. In fact, it has been already demonstrated that SeNPs can inhibit the tumor growth *via* induction of p53-mediated apoptosis<sup>31</sup>. Moreover, in this study, SeNPs have been decorated with calcium sulfate which also may have some anti-cancer and apoptotic effects<sup>32</sup>. Cytosolic calcium at high levels induces apoptosis *via* the mitochondrial pathway<sup>33</sup>. Therefore, calcium sulfate is considered an apoptotic inducer that can trigger the intrinsic (Classical: mitochondrial) pathways. Regarding these facts,  $\text{CaSO}_4\text{@SeNPs}$  fabricated during this study can be considered as double-edged sword by which targeted cancer cells are faced to two powerful cancer killing agents.

In the past decade, with the growing attention to nanotechnology, SeNPs were highly research friendly elements especially because the biological properties of SeNPs are similar to Se ions even at lower doses with lower toxicity<sup>14</sup>. One of the most probable applications of nanoparticles in the field of medicine is their possible capacity to be used in drug delivery systems although in the context of SeNPs, the story is somehow different. Like other metalloid NPs, SeNPs have been used in different modalities for cancer diagnosis and therapy. In fact, SeNPs have both capabilities to be used for targeting and also for killing cancer cells. Transferrin conjugated SeNPs as therapeutic agents loaded with doxorubicin showed a synergistic effect for cancer therapy with higher efficacy and fewer side effects<sup>34</sup>. Likewise, hyaluronic acid decorated SeNPs as therapeutic agents demonstrated a higher tumor inhibition ratio and reduced tumor weight<sup>35</sup>. These recent researches showed the potential of this nanoparticle to be used in drug delivery systems. On the other hand, the activation of apoptosis pathways is the mechanism that probably induces the direct anti-cancer effect of SeNPs. It is well documented that SeNPs induce apoptosis by depletion of mitochondrial membrane potential, Reactive Oxygen Species (ROS) overproduction, and cytochrome C release<sup>34</sup>.

In addition to overproduction and accumulation of free radicals, selenium causes oxidative stress and/or endoplasmic reticulum stress and mediates cell survival by modulating  $\text{Ca}^{2+}$  release and consecutive apoptosis<sup>36</sup>. In the present research, regarding the role of SeNPs in induction of apoptosis and also the similar role of calcium, the synergistic effect of these two pro-apoptotic agents has been demonstrated and results showed that the anti-cancer effect of  $\text{CaSO}_4\text{@SeNPs}$  at the dose of 50  $\mu\text{g}$  doubled in comparison to SeNPs alone. The same effect can also be observed at the doses of 25  $\mu\text{g}$  and 10  $\mu\text{g}$ . In fact, by these preliminary data, it can be proposed that  $\text{CaSO}_4\text{@SeNPs}$ , in comparison to SeNPs, can act as a double-edged sword. However, the capacity of NPs for delivery of anti-cancer drug to their target cells should not be ignored<sup>37</sup>. In other words, NPs not only have this ability to induce an anticancer response through the activation of apoptosis, but also are an appropriate candidate for targeting the cancer cells and are considered as a powerful delivery system which may accumulate more SeNPs in cancer cells in comparison with SeNPs that exclusively target the cancer cells.

However, interaction of selenium with intracellular proteins such as Glutathione peroxidase (Gpx), Superoxide Dismutase (SOD) or catalase, and any other enzymes that have cysteine in their active site and inhibition of their activity results in the accumulation of ROS in the cell cytoplasm<sup>38</sup>. Therefore, considering the higher level of these enzymes in cancer cells due to higher level of metabolism and mitochondrial respiration, an optimum dose of selenium may cause more

toxicity for cancer cells compared to normal cells.

### Conclusion

To sum up, results of the present work demonstrate that the decoration of SeNPs as known cytotoxic NPs for breast cancer cells with calcium sulfate may increase their anti-cancer potential, which can be observed by lowering the effective dose. However, for discovering the exact corresponding mechanisms of this synergistic effect which are proposed to be related to activation of intrinsic apoptosis signaling and/or pH regulatory properties of CaSO<sub>4</sub>@SeNPs, further *in vitro* and *in vivo* studies are still needed.

### Acknowledgement

Research reported in this publication was supported by Elite Researcher Grant Committee under award number [963338] from the National Institute for Medical Research Development (NIMAD), Tehran, Iran.

### Conflict of Interest

The authors declare no conflicts of interest.

### References

- Gupta A, Shridhar K, Dhillon P. A review of breast cancer awareness among women in India: Cancer literate or awareness deficit? *Eur J Cancer* 2015;51(14):2058-66.
- Uysal E. Top 100 cited classic articles in breast cancer research. *Eur J Breast Health*. 2017;13(3):129-37.
- Montagna E, Cancelli G, Dellapasqua S, Munzone E, Colleoni M. Metronomic therapy and breast cancer: a systematic review. *Cancer Treat Rev* 2014;40(8):942-50.
- Sharma R. Global, regional, national burden of breast cancer in 185 countries: evidence from GLOBOCAN 2018. *Breast Cancer Res Treat* 2021.
- Sonkusre P, Nanduri R, Gupta P, Cameotra SS. Improved extraction of intracellular biogenic selenium nanoparticles and their specificity for cancer chemoprevention. *J Nanomed Nanotechnol* 2014;5(2):1.
- Maiyo F, Singh M. Selenium nanoparticles: Potential in cancer gene and drug delivery. *Nanomedicine (Lond)* 2017;12(9):1075-89.
- Mao J, Pop VJ, Bath SC, Vader HL, Redman CW, Rayman MP. Effect of low-dose selenium on thyroid autoimmunity and thyroid function in UK pregnant women with mild-to-moderate iodine deficiency. *Eur J Nutr* 2016;55(1):55-61.
- Wang H, Zhang J, Yu H. Elemental selenium at nano size possesses lower toxicity without compromising the fundamental effect on selenoenzymes: comparison with selenomethionine in mice. *Free Radic Biol Med* 2007;42(10):1524-33.
- Tabinda S. Selenium role in reproduction, pregnant/postpartum women and neonates: A current study. *Current Nutrition & Food Science* 2021;17(1):28-37.
- Rock C, Moos PJ. Selenoprotein P regulation by the glucocorticoid receptor. *Biometals* 2009;22(6):995.
- Fang W, Han A, Bi X, Xiong B, Yang W. Tumor inhibition by sodium selenite is associated with activation of c-Jun NH<sub>2</sub>-terminal kinase 1 and suppression of  $\beta$ -catenin signaling. *Int J Cancer* 2010;127(1):32-42.
- Reid ME, Duffield-Lillico AJ, Garland L, Turnbull BW, Clark LC, Marshall JR. Selenium supplementation and lung cancer incidence: an update of the nutritional prevention of cancer trial. *Cancer Epidemiol Biomarkers Prev* 2002;11(11):1285-91.
- Sygit K, Cipora E, Smorawiński J. The role of selenium in inherited breast cancer in women. *Journal of Health Inequalities* 2020;6(2):160-5.
- Zhang JS, Gao XY, Zhang LD, Bao YP. Biological effects of a nano red elemental selenium. *Biofactors* 2001;15(1):27-38.
- Zhang J, Wang X, Xu T. Elemental selenium at nano size (Nano-Se) as a potential chemopreventive agent with reduced risk of selenium toxicity: comparison with selenomethionine in mice. *Toxicol Sci* 2007;101(1):22-31.
- Lu J, Berndt C, Holmgren A. Metabolism of selenium compounds catalyzed by the mammalian selenoprotein thioredoxin reductase. *Biochim Biophys Acta* 2009;1790(11):1513-9.
- Nayak V, Singh K, Singh A, Singh R. Potentialities of selenium nanoparticles in biomedical sciences. *New Journal of Chemistry* 2021;45(6):2849-78.
- Yazdi MH, Masoudifar M, Varastehmoradi B, Mohammadi E, Kheradmand E, Homayouni S, et al. Effect of oral supplementation of biogenic selenium nanoparticles on white blood cell profile of BALB/c mice and mice exposed to X-ray radiation. *Avicenna J Med Biotechnol* 2013;5(3):158-67.
- Wu JH, Deng YL, Liu Q, Yu JC, Liu YL, He ZQ, et al. Induction of apoptosis and autophagy by calcifying nanoparticles in human bladder cancer cells. *Tumor Biol* 2017;39(6):1010428317707688.
- Pourbaghi-Masouleh M, Hosseini V. Amorphous calcium phosphate nanoparticles could function as a novel cancer therapeutic agent by employing a suitable targeted drug delivery platform. *Nanoscale Res Lett* 2013;8(1):449.
- Khakinezhad Tehrani F, Ranji N, Kouhkan F, Hosseinzadeh S. Apoptosis induction and proliferation inhibition by silibinin encapsulated in nanoparticles in MIA PaCa-2 cancer cells and deregulation of some miRNAs. *Iran J Basic Med Sci* 2020;23(4):469-82.
- Shahverdi AR, Shahverdi F, Faghfuri E, Reza khosha-yand M, Mavandadnejad F, Yazdi MH, et al. Characterization of folic acid surface-coated selenium nanoparticles and corresponding *in vitro* and *in vivo* effects against breast cancer. *Arch Med Res* 2018;49(1):10-7.
- Xuan G, Zhang M, Chen Y, Huang S, Lee I. Design and characterization of a cancer-targeted drug co-delivery system composed of liposomes and selenium nanoparticles. *J Nanosci Nanotechnol* 2020;20(9):5295-304.
- Maleki Dizaj S, Barzegar-Jalali M, Zarrintan MH, Adibkia K, Lotfipour F. Calcium carbonate nanoparticles as

- cancer drug delivery system. *Expert Opin Drug Deliv* 2015;12(10):1649-60.
25. Khalifehzadeh R, Arami H. Biodegradable calcium phosphate nanoparticles for cancer therapy. *Adv Colloid Interface Sci* 2020;279:102157.
  26. Abdel-Gawad EI, Hassan AI, Awwad SA. Efficiency of calcium phosphate composite nanoparticles in targeting Ehrlich carcinoma cells transplanted in mice. *J Adv Res* 2016;7(1):143-54.
  27. Pi J, Jin H, Liu R, Song B, Wu Q, Liu L, et al. Pathway of cytotoxicity induced by folic acid modified selenium nanoparticles in MCF-7 cells. *Appl Microbiol Biotechnol* 2013;97(3):1051-62.
  28. Soumya RS, Vineetha VP, Reshma PL, Raghu KG. Preparation and characterization of selenium incorporated guar gum nanoparticle and its interaction with H9c2 cells. *PloS One* 2013;8(9):e74411.
  29. Dennert G, Zwahlen M, Brinkman M, Vinceti M, Zeegeers MP, Horneber M. Selenium for preventing cancer. *Cochrane Database Syst Rev* 2011;(5):CD005195.
  30. Tapiero H, Townsend D, Tew K. The antioxidant role of selenium and seleno-compounds. *Biomed Pharmacother* 2003;57(3-4):134-44.
  31. Huang Y, He L, Liu W, Fan C, Zheng W, Wong YS, et al. Selective cellular uptake and induction of apoptosis of cancer-targeted selenium nanoparticles. *Biomaterials* 2013;34(29):7106-16.
  32. Criddle DN, Gerasimenko JV, Baumgartner HK, Jaffar M, Voronina S, Sutton R, et al. Calcium signalling and pancreatic cell death: apoptosis or necrosis? *Cell Death Differ* 2007;14(7):1285-94.
  33. Wyllie AH, Morris RG, Smith AL, Dunlop D. Chromatin cleavage in apoptosis: association with condensed chromatin morphology and dependence on macromolecular synthesis. *J Pathol* 1984;142(1):67-77.
  34. Zhang Y, Li X, Huang Z, Zheng W, Fan C, Chen T. Enhancement of cell permeabilization apoptosis-inducing activity of selenium nanoparticles by ATP surface decoration. *Nanomedicine* 2013;9(1):74-84.
  35. Ren Y, Zhao T, Mao G, Zhang M, Li F, Zou Y, et al. Antitumor activity of hyaluronic acid-selenium nanoparticles in Heps tumor mice models. *Int J Biol Macromol* 2013;57:57-62.
  36. Uğuz AC, Nazıroğlu M, Espino J, Bejarano I, González D, Rodríguez AB, et al. Selenium modulates oxidative stress-induced cell apoptosis in human myeloid HL-60 cells through regulation of calcium release and caspase-3 and-9 activities. *J Membr Biol* 2009;232(1-3):15-23.
  37. Masouleh MP, Hosseini V, Pourhaghgouy M, Bakht MK. Calcium phosphate nanoparticles cytocompatibility versus cytotoxicity: A serendipitous paradox. *Curr Pharm Des* 2017;23(20):2930-51.
  38. Liang CC, Park AY, Guan JL. In vitro scratch assay: a convenient and inexpensive method for analysis of cell migration in vitro. *Nat Protoc* 2007;2(2):329-33.