Use of Raman Spectroscopy to Decrease Time for Identifying the Species of Candida Growth in Cultures

Nitin S. Chouthai, Anuj A. Shah, Hossein Salimnia, Olena Palyvoda, Suneetha Devpura, Michael Klein, and Basim Asmar

1. Division of Neonatal-Perinatal Medicine, Wayne State University, Detroit, MI, United States of America
2. Division of Pathology, Wayne State University, Detroit, MI, United States of America
3. Lumigen Instrument Center, Wayne State University, Detroit, MI, United States of America
4. Department of Radiation Oncology, Henry Ford Health System, Detroit MI, United States of America
5. Pediatric Surgery, Wayne State University, Detroit, MI, United States of America
6. Division of Pediatric Infectious Diseases, Wayne State University, Detroit, MI, United States of America

* Corresponding author: Nitin S. Chouthai, MBBS, DCH, MRCP, FAAP, Division of Neonatal-Perinatal Medicine, Carman and Ann Adams Department of Pediatrics, Wayne State University Children’s Hospital of Michigan and Hutzel Women’s Hospital, Detroit, MI, USA
Tel: +313 745 5638
Fax: +313 745 5807
E-mail: nchouthai@med.wayne.edu
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Abstract

**Background:** The objective of this study is to establish Raman signatures from pure cultures of different Candida species using Raman Spectroscopy (RS) and use these signatures for rapid identification of unknown Candida species.

**Methods:** Pure cultures of five Candida species were evaluated using RS to build a limited signature library. ‘Raman Processing’ (RP) software was used for Principal Component Analysis (PCA) and Differential Functional Analysis (DFA).

**Results:** Eleven principal components described at least 95% variance in the spectra. Raman signatures from these known Candida species were able to identify the species of unknown Candida cultures with 100% accuracy.

**Conclusion:** Raman spectroscopy can improve early identification of Candida species and may facilitate early optimal antifungal therapy.

Keywords: Candida species, Raman, Spectrum analysis

**Introduction**

Late-onset sepsis occurs in 2-20% of extremely low birth weight neonates and up to 12% of these infections are due to candida species. Furthermore, candida Blood Stream Infections (BSIs) are associated with a 25-40% mortality rate. Invasive candida infections have also been associated with a 73% rate of neurodevelopmental impairment or death. Candida growth in cultures were directly placed under the Raman identification system. All culture plates were incubated for 24 hr. The plates with growth of fungi were directly placed under the Raman

**Materials and Methods**

Colonies from five commonly occurring candida species grown on SDA media were evaluated. Candida species were obtained from American Type Culture Collection (C. albicans: ATCC 28815, C. parapsilosis: ATCC 34136, C. glabrata: ATCC 2001, C. krusei: ATCC 6258 and C. tropicalis: ATCC 13803). The identities of the samples were confirmed using API 20C identification system. All culture plates were inoculated using standard techniques of equal amount of inoculum and incubated for 24 hr. The plates with growth of fungi were directly placed under the Raman...
spectroscope without additional processing. All spectral measurements (C. albicans: 115 spectra, C. parapsilosis: 112 spectra, C. glabrata: 108 spectra, C. krusei: 117 spectra, C. tropicalis: 128 spectra from five SDA plates each and Unknown: 90 spectra from nine SDA plates) were obtained from 20-25 different points on each sample’s area of interest using Renishaw Wire 2.0 software.

Raman measurements were acquired using a Renishaw InVia Raman microscope (Renishaw, Gloucestershire, United Kingdom) by a 785 nm (infrared, IR) excitation laser with 50× objective. A single-grating spectrograph with a 1200 line/mm grating combined with a holographic notch filter for Rayleigh scattering rejection were used. A 50× objective lens was used to focus the excitation laser beam to a spot size of ~4 µm×30 µm on a single colony on the SDA medium. Once the laser was initiated, collection of the back-scattered light took place with the laser power set at 100% (3 mW). Each spectrum consisted of the average of 2 collections with a 10 s collection time and an extended range of 600 to 1800 cm⁻¹. Measurements were obtained using Renishaw Wire 2.0 software. At least 20 spectra were measured from different randomly selected points on each sample. The Raman Processing Software (RP Software) was used to import and analyze the data. The acquired raw spectra were corrected by subtracting background fluorescence, reducing noise, and normalizing the intensities. Data were analyzed using Principal Component Analysis (PCA) and Discriminant Function Analysis (DFA). The peaks in Raman spectra (wavenumbers) that were most significant to the classification process were identified. The biochemical correlates of these Raman peaks were also described using previously published work.

A data bank or library of different Raman signatures for each of the five candida species was created. Unknown colonies of candida species were also derived using ATCC reference species (C. albicans, C. parapsilosis, C. glabrata, C. krusei, and C. tropicalis) after creating the initial library databank. The investigating team was blinded to the identity of the unknown candida species. Colonies from cultures of unknown candida species, derived from sources other than those that were used to create the library, were compared with the known Raman signatures to identify their species.

**Results**

When processed with RP Software, five candida species examined showed eleven principal components generated by PCA, which account for 95.2 percent of the variance. Then the principal components were fed into DFA classifier, which enabled categorizing of unknown species. Figure 1 represents the mean normalized curves for each species and demonstrates the key differentiators among them. The chemical structure of the fungal elements represents different peaks within the Raman spectra. Based on the known spectra, the trained DFA classifier was able to identify unknown sample signatures with 100% accuracy. These differentiating peaks for different biological molecules have already been described. The Raman shift regions (wavenumbers/cm⁻¹) associated with significant peaks within the Raman spectra and corresponding biochemical elements are described in table 1. The Raman signatures for all the unknown species evaluated were compiled and placed together in figure 2. This figure again demonstrates key differences among the unknown species.
Raman Spectroscopy for Candida Cultures

Table 1. Wavenumbers of significant peaks in Raman spectra and corresponding biochemical elements

<table>
<thead>
<tr>
<th>Wave numbers (cm⁻¹)</th>
<th>Biochemical elements</th>
</tr>
</thead>
<tbody>
<tr>
<td>649</td>
<td>Amino acid</td>
</tr>
<tr>
<td>716</td>
<td>Nucleotide</td>
</tr>
<tr>
<td>910</td>
<td>Glucose</td>
</tr>
<tr>
<td>1003</td>
<td>Phenylalanine ring breathing mode</td>
</tr>
<tr>
<td>1083</td>
<td>Nucleic acid</td>
</tr>
<tr>
<td>1152</td>
<td>Protein/carotenoid</td>
</tr>
<tr>
<td>1263</td>
<td>Amide III band component protein</td>
</tr>
<tr>
<td>1337</td>
<td>Amide III/tryptophan/nucleic acid</td>
</tr>
<tr>
<td>1452</td>
<td>Delta CH2/Delta CH3</td>
</tr>
<tr>
<td>1603</td>
<td>Phenylalanine</td>
</tr>
<tr>
<td>1656</td>
<td>Amide bonds</td>
</tr>
</tbody>
</table>

Discussion

In very low birth weight neonates with fungal BSIs, it is difficult at times to obtain blood samples for cultures and the amount of blood collected is sometimes suboptimal. This limits the sensitivity of blood cultures and some neonates with candida infections might go undiagnosed until an autopsy is performed. There are some non-culture methods for candida identification including testing for fungal antigens and DNA available either commercially or in the evaluation phase. Use of DNA technology for detection of candida has shown promise. In very low birth weight neonates, however, the detection of fungal BSIs using DNA technology is not superior to fungal lysis-centrifugation isolator system. The DNA technology would need use of costly reagents and technical expertise. The use of RS did not include use of any consumables and evaluation could be completed within 30 min per unknown sample.

Conclusion

This study is the first to demonstrate the use of RS for identification of candida species recovered in cultures. After colonies grow on solid culture media, it takes approximately 48-72 hr to identify the candida species. The API 20C identification system used as a standard of practice involves carbohydrate assimilation by fungal colonies. RS involves identification of peaks of chemical elements followed by comparing those using PCA and DFA techniques.

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