Amplification of GC-rich Putative Mouse PeP Promoter using Betaine and DMSO in Ammonium Sulfate Polymerase Chain Reaction Buffer

Tahere Seifi 1, Kamran Ghaedi 1,2*, Ahmad Salamian 2, Sommayeh Tanhaei 2, Forouzan Safari 1, Zohreh Hojati 1, Manuchehr Tavassoli 1, Hossein Baharvand 3,4, and Mohammad-Hossein Nasr Esfahani 2

1. Department of Biology, School of Sciences, University of Isfahan, Isfahan, Iran
2. Department of Cell and Molecular Biology, Cell Science Research Center, Royan Institute for Animal Biotechnology, ACECR, Isfahan, Iran
3. Department of Stem Cells and Developmental Biology, Cell Science Research Center, Royan Institute for Stem Cell Biology and Technology, ACECR, Tehran, Iran
4. Department of Developmental Biology, University of Science and Culture, ACECR, Tehran, Iran

Abstract

Background: Recently, we have shown that peroxisomal protein expression was induced upon retinoic acid treatment in mouse embryonic stem cells during the process of neurogenesis. Thus, characterization of the respective promoter could elucidate the molecular aspects of transcriptional regulation of this gene.

Methods: Using the conventional software programs for promoter prediction, a putative promoter region was identified approximately 561 bp upstream of the peroxisomal protein coding sequence. In order to clone this region with a GC-content of 71.01%, a cocktail of ammonium sulfate buffer supplied with two additive components, betaine and dimethyl sulfoxide, and a high concentration of MgCl2 was used.

Results: The modulated polymerase chain reaction composition significantly improved the amplification of GC-rich DNA target sequences. Improved amplification of this region was due to reduction in the formation of secondary structures by the GC-rich region.

Conclusion: Therefore, this polymerase chain reaction composition could be generally used to facilitate the amplification of other GC-rich DNA sequences as verified by amplification of different GC-rich regions.

Keywords: Amplification, GC-rich region, Peroxisomal protein, Promoter

Introduction

Polymerase Chain Reaction (PCR) is a conventional method for amplification of DNA regions while templates with high percentages of GC bases including eukaryotic promoter regions are not easily amplified 1,4. In this state, PCR results in production of undesired truncated fragments instead of bona fide product 1,5.

Several approaches have been proposed to overcome this problem including additives usage in different PCR procedures 2,6. Different additives were utilized to amplify GC-rich regions including betaine (N,N,N-trimethylglycin), dimethyl sulfoxide (DMSO), formamide, glycerin, glycerol, dithiothreitol (DTT), Bovine Serum Albumin (BSA), non-ionic detergents such as Triton X-100, nucleotide analogs 7-deaza dGTP, ethylene glycol and 1, 2-propanediol, and reducing agents such as β-mercapto-ethanol 1,9. These additives not only prevent secondary structure formation of both templates and primers, but also increase the annealing chance of primers to plate strands at temperatures lower than the melting temperature of primers 1,3,5,7,9.

In the present study, we have utilized a combination of betaine and DMSO to amplify the putative promoter region of mouse peroxisomal protein (PeP). Mouse PeP which has been identified as a fibronectin type III domain containing protein (FNDC5; NCBI accession no: NC_000070) is supposed to undergo proteolytic cleavage and releasing to the extracellular space 10,11. Our data have elucidated that its expression increased upon Retinoic Acid (RA) treatment during the process of neurogenesis 12.

Very recently Bostrom et al have indicated that the cleaved form of PeP named irisin is a secreting myokine which its production could be induced after exercise, may improve the glucose homeostasis by regulating the obesity 11. To find out molecular mechanisms involved in the regulation of PeP expression, a putative promoter region at position -561/+101 relative to the Transcription Start Site (TSS), upstream of the PeP gene was chosen for amplification and further cloning.
Materials and Methods

To predict a potential promoter region of mouse PeP (Accession No: NC_000070), approximately 2x10^6 bp upstream of the PeP gene was submitted to the Genomatix (http://www.Genomatix.de/en/index.html) (13) and Proscan (http://www.proscan.co.za/) (Version 1.7) software programs. DNA extraction was performed on i) one Caucasian individual after signing a consent form and ii) a heart tissue sample of NMRI mouse, which was obtained from Royan Institute (Tehran, Iran) using DNeasy Blood and Tissue Kit (Qiagen, Germany). The ethics approval to use specimen was obtained from Royan Institute Bioethics Committee (Approval ID. No: EC-90-1077). Then 857 bp upsteam of the PeP gene including a part of the transcriptional region was chosen for amplification using specific primers in conventional PCR (Table 1). Then, a set of PCRs was performed to identify the optimal amplification conditions using betaine (Sigma), DMSO (Merck), glycerol (Merck), formamide (Scharlau), DTT (Sigma) (14). Moreover, different approaches of PCR including touch-down PCR (16), slow-down PCR (12) and hot-start PCR were applied. Finally, the successful amplification condition for amplicon (875 bp), included 10x PCR buffer AMS [composed of 750 mM of tris-HCl (pH=8.8), 200 mM of (NH4)2SO4, 0.1% Tween 20 (Cinna-Gen, Iran) and 4 mM Mg2+ (Cinnagen, Iran). To improve PCR efficiency, different concentrations of betaine (0.5 to 1 M) and DMSO (5-10% v/v) were applied. Simultaneously, similar amplification condition was repeated in Pfu buffer [200 mM of Tris-HCl (pH=8.8), 100 mM of KCl, 100 mM of (NH4)2SO4, 1% Triton X-100 and 1 mg/ml of nuclease-free BSA] (Fermentas, Lithuania) instead of 10 x PCR buffer AMS. Touch-down PCR was carried out in an Eppendorf thermocycler (Eppendorf, Germany) with one step of initial denaturation at 95°C for 5 min followed by twenty repetitive cycles, including 94°C for 10 s, a variable annealing step and an elongation step at 72°C for 4 min. The variable annealing step was started at a temperature of 66°C which decreased by 0.5°C/ cycle to reach a temperature of 56 °C for 30 s. Subsequently, additional amplification was continued for 20 cycles with a constant annealing temperature of 56°C for 30 s. PCR was terminated after final elongation step at 72°C for 10 min.

To demonstrate whether optimized PCR condition was applicable for other GC enriched regions, similar procedure was repeated for amplification of different loci on human genome (Table 1). Secondary structure predictions of the putative promoter of PeP and other amplified loci were assessed by Mfold web server (http://mfold.Bio-info.rpi.edu/) (14).

Results

Putative promoter prediction and amplification

Computational analyses predicted a putative promoter region in location of -561 to +101 relative to the TSS of PeP gene (Figure 1A) with high GC content (approximately 71.01%) (Figure 1B). Secondary structure prediction of this region indicated formation of nine independent structures owing to its high internal energy in the range of -199.73 to -209.77 kcal/mol (data not shown). Two extreme structures with the highest and the lowest internal energy are shown in figure 1C. Cloning of this region was carried out using different PCRs to obtain proper amplification. However, there were no product yields regardless of different additives (Figure 2A). Data indicated utilization of a mixture containing 10x PCR buffer AMS with different amounts of betaine (final: 0.5, 0.75 and 1 M), and 5-10% (v/v) of DMSO improved PCR yield (Figure 2B, lanes 5-7). Meanwhile, substitution of Pfu buffer with 5% (v/v) DMSO in the PCR reaction resulted in amplification of desired fragment (Figure 2C, lane 2).

Improvement of the PCR yield

Interestingly increased amounts of MgCl2 (3 and 4 mM) improved PCR yields. Particularly, PCR buffer AMS supplied with DMSO (10% v/v), betaine (final concentration: 1 M) and 4 mM of MgCl2 improved PCR efficiency (Figure 3). While, utilization of touch down PCR did not enhance the PCR product amounts compared to the conventional PCR approach (Figure 4).

To verify the efficacy of our approach, Androgen Receptor (Exon 1) and Eukaryotic Releasing Factor 3a (Exon 1) genes from a human source and a portion of human Elongation Factor 1a Promoter encompassing 71.8, 75.94 and 60.09% GC bases respectively were chosen for amplification (Table 1). The expected 270 bp PCR product band for exon 1 of human Androgen receptor was observed significantly when 5-10% DMSO, 0.5-0.75 M betaine was used in PCR buffer AMS or Pfu buffer (Figure 5A, left panel). Similarly, the amplified region of Eukaryotic Releasing Factor 3a

<table>
<thead>
<tr>
<th>Amplified DNA (species)</th>
<th>Primers sequence</th>
<th>Length of product (bp)</th>
<th>Product GC%</th>
<th>Accession No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>PeP Promoter (Mouse)</td>
<td>F: 5’ GCT ATT AAT GAA AGG ACT GCT CGT AGG TCT CAG 3’</td>
<td>875 bp</td>
<td>70.01%</td>
<td>NC_000070</td>
</tr>
<tr>
<td></td>
<td>R: 5’ GTT GCT AGC CTA GGG TCA GGT TAG GAG AGT AGG 3’</td>
<td>270 bp</td>
<td>71.80%</td>
<td>NC_000023</td>
</tr>
<tr>
<td>Androgen receptor (Exon I) (Human)</td>
<td>F: 5’ ACC CTC AGC CGC CTC TTC CTC ACC 3’</td>
<td>142 bp</td>
<td>75.94%</td>
<td>NC_000166</td>
</tr>
<tr>
<td></td>
<td>R: 5’ CGT GGA TAG GGC ACT CGT CTC ACC 3’</td>
<td>270 bp</td>
<td>71.80%</td>
<td>NC_000023</td>
</tr>
<tr>
<td>Eukaryotic releasing factor 3a (Exon I) (Human)</td>
<td>F: 5’ CAT TTC TCG TCT TCT GTC CAC 3’</td>
<td>142 bp</td>
<td>75.94%</td>
<td>NC_000166</td>
</tr>
<tr>
<td></td>
<td>R: 5’ GTC GTC CCA CCA GCA GTG AGG 3’</td>
<td>270 bp</td>
<td>71.80%</td>
<td>NC_000023</td>
</tr>
<tr>
<td>Elongation factor 1a promoter (Human)</td>
<td>F: 5’ GTT ATT AAT CGT GAG GCT CCG GTG 3’</td>
<td>1186 bp</td>
<td>60.09%</td>
<td>NC_000020</td>
</tr>
<tr>
<td></td>
<td>R: 5’ GCC GCT AGC TCA CGA CAC CTG A3’</td>
<td>270 bp</td>
<td>71.80%</td>
<td>NC_000023</td>
</tr>
</tbody>
</table>

F and R, are referred as forward and reverse primers respectively. Axel (ATTAAAT) and Nhel (GCTAGC) restriction sites are underlined.
PeP Promoter Amplification

(142 bp) was visualized when 5-7.5% DMSO, 0.5-0.75 M betaine in PCR buffer AMS reaction was used (Figure 5A, right panel). Moreover, amplification for promoter region of Elongation Factor 1a was achieved when 4 mM of MgCl2 was added to the Pfu buffer (Figure 5A, middle panel and Figure 5B). Secondary structure prediction of aforementioned amplicons indicated formation of secondary structures due to presence of high internal energy (Figure 5C).

Discussion

Amplification of GC-rich genomic regions requires time-consuming work to be optimized. Additives such as betaine (1,3,5), DMSO (1,3,5,6,8), form-amide (1,3), glycerol (1,6), glycerin (7), BSA (3), non-ionic detergents such as Triton X-100 (1), nucleotide analogs 7-deaza dGTP (5-7), DTT (3), ethylene glycol (9), 1,2-propanediol (9), reducing compounds such as β-mercapto-ethanol (3) and their combination have been utilized to optimize amplification. However, effects of these additives on amplification of different fragments are not similar (9). For instance, betaine not only acts to reduce the DNA thermal melting transition dependent on base pair composition, but also does not allow DNA polymerase to pause near hairpin-loop structures (7). Thus, a combination of betaine and DMSO make GC-rich fragments of DNA accessible for DNA polymerase (7). However, in this research no efficient amplification was seen using both betaine and DMSO. Previous reports have indicated that amplification of 71%> GC-rich DNA fragments was achieved by the addition of 16.6 mM (NH4)2SO4, 3.1 mM MgCl2 and 0.01% Tween 20 (3). In the present study, in order to amplify mouse PeP promoter with 71.01% GC, two different reaction buffers (10x PCR reaction buffer AMS and Pfu buffer) containing (NH4)2SO4 were utilized. The application of these reaction buffers resulted in efficient amplification.

Figure 1. DNA sequence, GC and CpG nucleotide composition graph of the putative promoter of PeP gene. A) Sequence of DNA fragment consisting of PeP gene (Accession no: NC_000070) putative promoter. The initiation codon is underlined. B) Genetyx software was used to plot the GC-graph and CpG plot programs (http://www.ebi.ac.uk/Tools/ecgi-bin/jobresults.ecgi/cpgplot/cpgplot-20100023-1343338151.html). Translation start site (TSS) is located at position +21 relative to the transcription start site. Primer sequences are shown below the graph. C) Two different secondary structures predicted graphs of target DNA based on the Mfold web server (http://mfold.bioinfo.rpi.edu/) for nucleic acid folding and hybridization prediction as noted in the Materials and Methods section. The internal energy required for formation of these constructs is indicated as kcal/mole.
due to reduction in the formation of secondary structure due to GC-rich regions. Computational analysis confirmed that secondary structure formation of a putative region of mouse PeP promoter was highly possible. Previous studies have shown that proper concentration of Mg\textsuperscript{2+} ion increased the productivity and fidelity of DNA polymerases due to the increased DNA/DNA interactions and formation of complexes with dNTPs. However, higher amounts of MgCl\textsubscript{2} may inhibit polymerase activity\textsuperscript{15-17}. In the present study, increased MgCl\textsubscript{2} concentration (4 mM) was efficient. In some instances, modifications of the PCR procedure including a touch-down PCR is needed to gain satisfactory PCR products. However, utilization of this procedure did not yield more product than the conventional PCR. Our data demonstrated that addition of (NH\textsubscript{4})\textsubscript{2}SO\textsubscript{4} in PCR reaction with suitable concentration of MgCl\textsubscript{2}, betaine and DMSO would yield the highest amount of several amplicons.

Acknowledgement

We thank Mr. Mohammad Hadi Sekhavati for his excellent technical assistance and helpful advice. This project was financed by a grant-in-aid of research from Royan Institute (Grant No. 148) to Kamran Ghaedi as the principal investigator and in support of Ms. Tahere Seifi for obtaining her MSc degree from the University of Isfahan.

References


17. http://wiki.answers.com/Q/why is a variable in PCR.


