



Strong Association of Polymorphism in *SPRED2* Gene with Disease Susceptibility and Clinical Characteristics of Rheumatoid Arthritis in the Iranian Population

Bahram Pakzad ¹, Hamed Moghadammanesh ¹, Mansour Salesi ¹, and Rasoul Salehi ^{2*}

1. Division of Rheumatology, Department of Internal Medicine, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran

2. Department of Genetics and Molecular Biology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran

Abstract

Background: The high heritability of Rheumatoid Arthritis (RA) has been estimated from different studies. Recently, Genome-Wide Association Studies (GWAS) show a large number of Single Nucleotide Polymorphisms (SNPs) loci affecting susceptibility to RA. The rs934734 polymorphism in the *SPRED2* gene is one of these loci. Studies have shown that the *SPRED2* gene is involved in the regulation of inflammatory response, leukocyte infiltration, and local chemokine production. In the current study, the possible association between SNP rs934734 (intronic variant) in the *SPRED2* gene with RA risk in the Iranian population was evaluated.

Methods: One hundred fourteen RA patients and 120 healthy counterparts were recruited in this case-control study to evaluate rs934734 genotypes using the real-time PCR High Resolution Melting method (HRM).

Results: Logistic regression analysis demonstrated that GG and AG genotypes compared with AA genotype increase the risk of RA (GG vs. AA; OR=4.61; 95%CI [2.21-9.35]; p<0.001 and AG vs. AA; OR=2.54; 95%CI [1.36-4.76]; p=0.004). Furthermore, subjects with allele G were more frequently affected with RA than subjects with A allele (OR=2.33; 95%CI [1.61-3.38]; p<0.001). Besides, in the patient group, there was a significant correlation between Erythrocyte Sedimentation Rate (ESR) and C-reactive protein (CRP) concentration with rs934734 polymorphism (p<0.05).

Conclusion: Our findings suggest that rs934734 in *SPRED2* strongly underlies RA development and is associated with clinicopathological characteristics of this disease.

Avicenna J Med Biotech 2022; 14(2): 170-174

Keywords: Autoimmune disease, Genotype, Iran, Rheumatoid arthritis, Single nucleotide polymorphisms

Introduction

Rheumatoid Arthritis (RA), classified as an autoimmune disease, is a long-term progressive disorder that causes inflammation and painful swelling in and around the joints and other body organs ^{1,2}. The prevalence of RA varies between 0.5 and 1% and begins at any age, but its incidence increases with age ^{3,4}. RA, similar to most of the other autoimmune diseases, displays a striking imbalance between the two sexes, with two or three times higher prevalence in women than men ^{5,6}.

RA has a multifactorial etiology as the interaction of several genetic and environmental factors is essential for the development of this disorder ⁷. Multiple studies have shown that genetic factors are critically involved in the incidence of RA ⁸. With regard to twin studies, the heritability of RA has been estimated to be

50-60% ⁹. Furthermore, some works demonstrated that a positive family history of RA increases the risk of this disease 3-9 folds ¹⁰.

The most frequent type of variation in the human genome is Single Nucleotide Polymorphism (SNP) with prevalence of one in every 300 nucleotides which could be associated with diseases such as RA ¹¹⁻¹³. With recent advances in genotyping and sequencing techniques, studies have shown a large number of SNPs loci that impact the susceptibility to RA. For instance, more than 100 risk-bearing loci for RA have been identified in people of European and Asian descent by Genome-Wide Association Studies (GWAS) ^{14,15}. Several GWAS that have been conducted to recognize new predisposing genes of the RA disease have revealed that the rs934734 in sprouty-related protein

with EVH1 domain 2 (*SPRED2*) gene is one of the candidate loci¹⁶.

SPRED2 gene (located in chromosome 2) is a member of the SPRED family of proteins and negatively regulates the Ras/Raf/ERK/MAPK pathway^{17,18}. Studies on *Spred2* knockout (*Spred2*^{-/-}) mice demonstrated that this gene negatively regulates inflammatory response in the colonic mucosa, leukocyte infiltration, and local chemokine production in polymicrobial septic peritonitis¹⁹⁻²¹. The GWAS also demonstrated that *Spred2* gene is a candidate gene for the development of IBD²².

The rs934734 (g.65595586G>A) is an intronic variant of the *SPRED2* gene. GWAS demonstrated that the G allele of rs934734, as a risk factor, increases the risk of RA development^{23,24}. In the present study, for the first time, the possible association between SNP rs934734 in the *SPRED2* gene with RA risk in the Iranian population was assessed. Also, the possible influence of association between some demographic and laboratory characteristics and rs934734 genotypes on susceptibility to RA was assessed.

Materials and Methods

Study population and sample preparation

In this case-control study, a total of 114 unrelated subjects with RA as a case group (mean age: 47.39±10.50 years) and 120 unrelated healthy subjects as a control group (mean age: 45.39±12.73 years) were included. Patients were recruited from Alzahra Hospital in Isfahan city of Iran. All the RA patients met the diagnostic criteria introduced by the American College of Rheumatology (ACR). All healthy controls and their families had no symptoms or history of RA or other immunological and autoimmune disorders. The participants of the study were interviewed and data on sex, age (at sampling time and onset), Body Mass Index (BMI, calculated as weight [kg] divided by height [m²] squared), blood pressure, and family history of RA and other autoimmune conditions were obtained using a structured questionnaire. Also, laboratory characteristics such as Erythrocyte Sedimentation Rate (ESR), C-reactive protein (CRP), White Blood Cell (WBC) count, hemoglobin, Platelet count test (PLT), creatinine, Blood Urea Nitrogen (BUN), Fasting Blood Sugar (FBS), High-Density Lipoprotein (HDL), Low-Density Lipoprotein (LDL), and Triglyceride (TG) were recorded. This study was approved by the university ethics board (Approved number: IR.MUI.REC.1397.102) and all participants gave written informed consent.

DNA extraction and genotyping of polymorphism

Approximately, 5 ml of venous blood was collected into EDTA anticoagulant tubes from each participant and stored at -20°C for DNA isolation. Genomic DNA was extracted from 200 µl of peripheral blood samples using a kit (GenetBio Co., Korea) consistent with the instruction manual. The purity and concentration of all

genomic DNA samples were assessed by agarose gel electrophoresis and spectroscopy at wavelengths of 260 and 280 nm, respectively, and then DNA was stored at -20°C.

The real-time polymerase chain reaction high-resolution melting (HRM) method was used to determine rs934734 polymorphism genotypes. HRM was performed using Type-it HRM PCR kit that contained HotStarTaq plus DNA polymerase and EvaGreen dye (Qiagen GmbH, Germany), and the analysis was carried out with Rotor-Gene 6000™ (Corbett Research Pty Limited, Australia). The forward and reverse primer sequences for the 180-bp fragment that spanned the rs934734 in the *SPRED2* gene were CCTCCTCA CGAGGACTGC and TGAAGTGTGAACCTATTCT CCAAA, respectively. The thermal profile of the reaction was 5 min of denaturation at 95°C, 40 cycles of 95°C for 10 s, 58°C for 30 s and 72°C for 20 s. The melting curve was generated by increasing the temperature between 65°C and 95°C at the rate of 0.1°C/s. Finally, melting curves were normalized among the two temperatures to determine the specimens with known genotypes as standard. For utilizing sample genotypes in HRM analysis as a standard, some samples were subjected to direct Sanger sequencing and their correct genotypes were determined.

Statistical analyses

The SPSS software v22 (IBM, USA) was used for statistical analyses. The allele and genotype frequencies were tested for Hardy Weinberg equilibrium by the χ^2 test. Logistic regression analysis was accomplished to investigate the association between genotypes and RA and to calculate specific Odds Ratios (ORs), 95% Confidential Intervals (CIs), and p-values. For demographic, clinical, and laboratory characteristics, p-values were calculated using independent sample t-test, Chi-square or Mann-Whitney U test. The significance level was set at p<0.05.

Results

To evaluate the correlation between rs934734 polymorphism with RA incidence, totally 234 subjects in case and control groups were analyzed among whom 114 patients (82 females and 32 males, mean age at sampling time: 47.39±10.50) were in the case group and 120 (81 females and 39 males, mean age at sampling time: 45.39±12.73) were healthy subjects in the control group. Demographic and clinical characteristics of subjects who were included in the study are shown in table 1. There was no substantial correlation between case and control groups in terms of age (p=0.189) and gender (p=0.480), demonstrating that for these variables matching was adequate. From all patients, twenty (17.5%) patients had a positive family history whereas controls did not have a history of RA or other autoimmune diseases. Based on laboratory tests, ESR, CRP, WBC count, and creatinine were significantly higher in patients than in healthy controls

rs934734 as the Strong Determinant of Rheumatoid Arthritis

Table 1. Baseline characteristics of RA patients and control subjects

Characteristics	Patients	Controls	p-value
Total number	114	120	
Age at sampling time	47.39±10.50	45.39±12.73	0.189
Gender n (%)			
Male	32 (28.1%)	39(32.5%)	0.480
Female	82 (71.9%)	81 (67.5%)	
Age of onset	41.13±10.44	--	--
BMI	26.22±2.47	24.14±3.32	0.468
SBP	122.46±12.50	120.92±9.74	0.293
DBP	78.42±7.80	78.75±8.28	0.755
Positive family history n (%)	20 (17.5%)	0 (0%)	--

Data are mean±SD, or n (%). * p-value<0.05; SD: Standard deviation; RA: Rheumatoid arthritis; BMI: Body mass index; SBP: Systolic blood pressure; DBP: Diastolic blood pressure.

(p<0.05) while hemoglobin level in control group was significantly higher in comparison to patients group (p<0.001). Positive RF was observed in all patients. Other laboratory factors including PLT, BUN, FBS, HDL, LDL, and TG were not significantly different between patients and healthy controls (p>0.05). The details of laboratory characteristics of patients with RA and healthy controls are presented in table 2.

Genotype and allele distribution

In this study, it was demonstrated that the genotype distribution of rs934734 polymorphism in two groups was in agreement with Hardy-Weinberg equilibrium. Significant association was found between GG (compared with AA; p<0.001) and AG (compared with AA; p=0.004) genotypes with the risk of RA. The frequency of AA, AG, and GG genotypes was 20.2%, 46.5%, and 33.3% in patients and 44.2%, 40%, and 15.8% in controls, respectively (Table 3). Analysis of the combined genotype demonstrated that the GG+AG increases the risk of RA compared with the AA genotype (p<0.001). In allele distribution analysis, an increased level of allele G in patients (56.6%) was observed when com-

Table 2. Laboratory characteristics of RA patients and control subjects

	Patients (114)	Controls (120)	p-value
ESR (mm/h)	36.53±23.22	15.58±6.92	<0.001 *
CRP (mg/l)	16.38±18.57	4.45±2.58	<0.001 *
White blood cell (10 ⁹ /l)	7.33±2.18	6.57±1.37	0.002 *
Hemoglobin (HB)	12.67±0.58	14.32±1.60	<0.001 *
PLT (10 ⁹ /l)	261.74±72.17	251.03±66.77	0.239
Creatinine (mg/dL)	0.70±1.40	0.86±0.17	<0.001 *
BUN	17.15±4.68	16.17±4.1	0.073
FBS	96.46±15.84	92.92±21.95	0.160
HDL	49.44±7.65	50.41±11.10	0.439
LDL	109.84±29.01	106.91±31.15	0.450
TG	169.32±48.31	155.58±59.86	0.055

Data are mean±SD, or n (%). * p<0.05; SD: Standard deviation; RA: Rheumatoid arthritis; ESR: Erythrocyte sedimentation rate; CRP: C-reactive protein; BUN: Blood urea nitrogen; PLT: Platelet; HDL: High-density lipoprotein; LDL: Low-density lipoprotein; TG: Triglyceride; FBS: Fasting blood sugar.

pared with controls (35.8%), which was statistically significant (p<0.001). Furthermore, stratification based on sex and age of onset displayed a non-significant correlation between these subgroups and SNP genotypes in patients (p=0.183 and p=0.870, respectively). The mean concentration of ESR and CRP in the patient group was significantly different in genotype stratification (p<0.05) while there was no significant association between BMI, creatinine, and hemoglobin with this polymorphism (p>0.05) (Table 4).

Discussion

Convergent lines of evidence indicated that a large number of SNPs loci affect susceptibility to autoimmune disorders^{25,26}. Several GWAS demonstrated that numerous SNPs loci in different genes are associated with RA^{15,27}. The association of rs934734 polymorphism in the *SPRED2* gene (located at chromosome 2p14) with RA was revealed by two GWAS^{23,24}. Researches have shown that the *SPRED2* gene is involved

Table 3. Association between genotypes and allele frequency with RA risk

Genotype group	Patients (n=114) n (%)	Controls (n=120) n (%)	OR (95% CI)	p-value
AA	23 (20.2%)	53 (44.2%)	Reference	---
AG	53 (46.5%)	48 (40.0%)	2.54 (1.36, 4.76)	<0.004 *
GG	38 (33.3%)	19 (15.8%)	4.61 (2.21, 9.35)	<0.001 *
Combined genotype				
AA	23 (20.2%)	53 (44.2%)	Reference	---
AG+GG	91 (79.8%)	67 (55.8%)	3.13 (1.32, 5.62)	<0.001 *
Allele				
A	99 (43.4%)	154 (64.2%)	Reference	---
G	129 (56.6%)	86 (35.8%)	2.33 (1.61, 3.38)	<0.001 *

* p-value<0.05.

Table 4. Stratification analysis of the *SPRED2* polymorphism (rs934734) in patients

Genotype group	AA (n=23)	AG (n=53)	GG (n=38)	p-value
Age of onset	40.74±10.25	40.77±10.86	41.87±10.19	0.870
Sex				
Males	3 (13.0%)	16 (30.2%)	13 (34.2%)	0.183
Females	20 (87.0%)	37 (69.8%)	25 (65.8%)	
ESR (mm/h)	24.13±10.27	35.04±20.51	46.11±28.29	<0.001 *
CRP (mg/l)	3.70±2.36	11.10±6.32	31.48±24.87	<0.001 *
Creatinine (mg/dL)	0.99±0.20	1.04±0.18	1.03±0.19	0.642
BMI	25.92±3.26	26.37±1.80	26.20±2.77	0.773
Hemoglobin (HB)	12.63±1.20	12.20±6.50	12.65±1.10	0.073

Data are mean±SD, or n (%). * p-value<0.05. ESR: Erythrocyte sedimentation rate; CRP: C-reactive protein; BMI: Body mass index; SD: Standard deviation.

in the regulation of inflammatory response, leukocyte infiltration, and local chemokine production^{19,20}. Therefore, dysregulation of this gene could be involved in autoimmune pathogenesis. To the best of our knowledge, the present study was the first single locus association study that showed the association between rs934734 polymorphism in *SPRED2* gene with the risk of RA. In our study, logistic regression analysis demonstrated that homozygous GG and heterozygous AG genotypes compared with the AA genotype increase the risk of RA (GG vs. AA; OR=4.61; 95%CI [2.21-9.35] and AG vs. AA; OR=2.54; 95%CI [1.36, 4.76]). Likewise, combined genotype analyses indicated that GG+AG compared with the AA genotype increases the risk of disease (OR=3.13; 95%CI [1.32-5.62]). Moreover, individuals with allele G were more frequently affected with RA than subjects with A allele (OR=2.33; 95%CI [1.61, 3.38]) (Table 3). Our analysis was consistent with a GWAS that carried out on people of European descent and revealed that the G allele is a risk factor for RA¹⁹. However, these results were not confirmed in the other GWAS on African-American populations²⁸. Furthermore, Mizuno *et al* reported that this polymorphism was not correlated with the progression of functional disability in RA patients²⁹.

Besides, in the patient group, a significant correlation between the sex group, the mean age of onset, and also ESR and CRP concentration and different genotypes of rs934734 polymorphism (p<0.05) was found (Table 4). One of the main limitations of the current study was the small sample size.

Conclusion

Our results showed that rs934734 in *SPRED2* is a strong determinant for the development of RA and clinicopathological characteristics of this disease. In our previous studies, similar results were also found about association of specific polymorphisms with clinicopathological characteristics of RA disease including

variants in *NLRP3*, *NOD2*, and *microRNA-124* genes³⁰⁻³².

However, performing replication studies in different populations is a necessity to validate these results. Finally, further association studies with larger sample size would help to confirm the suggested correlations. Also, other polymorphisms that were not included in our study might be involved in determining the risk of RA, thus future studies are necessary.

Acknowledgement

We would like to appreciate any support provided by Isfahan University of Medical Sciences.

References

- Silman AJ, Pearson JE. Epidemiology and genetics of rheumatoid arthritis. *Arthritis Res* 2002; 4 Suppl 3(Suppl 3):S265-72.
- Guo Q, Wang Y, Xu D, Nossent J, Pavlos NJ, Xu J. Rheumatoid arthritis: pathological mechanisms and modern pharmacologic therapies. *Bone Res* 2018;6(1): 15.
- Hunter TM, Boytsov NN, Zhang X, Schroeder K, Michaud K, Araujo AB. Prevalence of rheumatoid arthritis in the United States adult population in healthcare claims databases, 2004–2014. *Rheumatol Int* 2017;37(9):1551-7.
- Crowson CS, Matteson EL, Myasoedova E, Michet CJ, Ernste FC, Warrington KJ, et al. The lifetime risk of adult-onset rheumatoid arthritis and other inflammatory autoimmune rheumatic diseases. *Arthritis Rheum* 2011; 63(3):633-9.
- Ortona E, Pierdominici M, Maselli A, Veroni C, Aloisi F, Shoenfeld Y. Sex-based differences in autoimmune diseases. *Ann Ist Super Sanita* 2016;52(2):205-12.
- van Vollenhoven RF. Sex differences in rheumatoid arthritis: more than meets the eye. *BMC Med* 2009;7:12.
- Deane KD, Demoruelle MK, Kelmenson LB, Kuhn KA, Norris JM, Holers VM. Genetic and environmental risk

- factors for rheumatoid arthritis. *Best Pract Res Clin Rheumatol* 2017;31(1):3-18.
8. Raychaudhuri S. Recent advances in the genetics of rheumatoid arthritis. *Curr Opin Rheumatol* 2010;22(2): 109-18.
 9. Svendsen AJ, Kyvik KO, Houen G, Junker P, Christensen K, Christiansen L, et al. On the origin of rheumatoid arthritis: the impact of environment and genes--a population based twin study. *PLoS One* 2013;8(2): e57304.
 10. Karlson EW, van Schaardenburg D, van der Helm-van Mil AH. Strategies to predict rheumatoid arthritis development in at-risk populations. *Rheumatology (Oxford)* 2016;55(1):6-15.
 11. Saad MN, Mabrouk MS, Eldeib AM, Shaker OG. Identification of rheumatoid arthritis biomarkers based on single nucleotide polymorphisms and haplotype blocks: a systematic review and meta-analysis. *J Adv Res* 2016;7(1):1-16.
 12. Karimzadeh MR, Zarin M, Ehtesham N, Khosravi S, Soosanabadi M, Mosallaei M, et al. MicroRNA binding site polymorphism in inflammatory genes associated with colorectal cancer: literature review and bioinformatics analysis. *Cancer Gene Ther* 2020;27(10):739-53.
 13. Simonian M, Mosallaei M, Khosravi S, Salehi R. rs12904 polymorphism in the 3'-untranslated region of ephrin A1 ligand and the risk of sporadic colorectal cancer in the Iranian population. *J Cancer Res Ther* 2019;15 (1):15.
 14. Okada Y, Eyre S, Suzuki A, Kochi Y, Yamamoto K. Genetics of rheumatoid arthritis: 2018 status. *Ann Rheum Dis* 2019;78(4):446-53.
 15. Danila MI, Laufer VA, Reynolds RJ, Yan Q, Liu N, Gregersen PK, et al. Dense genotyping of immune-related regions identifies loci for rheumatoid arthritis risk and damage in African Americans. *Mol Med* 2017;23: 177-87.
 16. Julià A, López-Longo FJ, Venegas JJP, Bonàs-Guarch S, Olivé A, Andreu JL, et al. Genome-wide association study meta-analysis identifies five new loci for systemic lupus erythematosus. *Arthritis Res Ther* 2018;20(1):100.
 17. Siljamäki E, Abankwa D. SPRED1 interferes with K-ras but not H-ras membrane anchorage and signaling. *Mol Cell Biol* 2016;36(20):2612-25.
 18. Wakioka T, Sasaki A, Kato R, Yoshida T, Matsumoto A, Miyoshi K, et al. Spred is a Sprouty-related suppressor of RAS signalling. *Nature*. 2001;412:647-51.
 19. Takahashi S, Yoshimura T, Ohkura T, Fujisawa M, Fushimi S, Ito T, et al. A novel role of spread2 in the colonic epithelial cell homeostasis and inflammation. *Sci Rep* 2016;6:37531.
 20. Itakura J, Sato M, Ito T, Mino M, Fushimi S, Takahashi S, et al. Spred2-deficiency protects mice from polymicrobial septic peritonitis by enhancing inflammation and bacterial clearance. *Sci Rep* 2017;7(1):12833.
 21. Ohkura T, Yoshimura T, Fujisawa M, Ohara T, Marutani R, Usami K, et al. Spred2 regulates high fat diet-induced adipose tissue inflammation, and metabolic abnormalities in mice. *Front Immunol* 2019;10:17.
 22. Van Limbergen J, Radford-Smith G, Satsangi J. Advances in IBD genetics. *Nat Rev Gastroenterol Hepatol* 2014;11(6):372-85.
 23. Stahl EA, Raychaudhuri S, Remmers EF, Xie G, Eyre S, Thomson BP, et al. Genome-wide association study meta-analysis identifies seven new rheumatoid arthritis risk loci. *Nat Genet* 2010;42(6):508-14.
 24. Scott IC, Seegobin SD, Steer S, Tan R, Forabosco P, Hinks A, et al. Predicting the risk of rheumatoid arthritis and its age of onset through modelling genetic risk variants with smoking. *PLoS Genet* 2013;9(9):e1003808.
 25. Ye J, Gillespie KM, Rodriguez S. Unravelling the roles of susceptibility loci for autoimmune diseases in the post-GWAS era. *Genes (Basel)* 2018;9(8):377.
 26. Gregersen PK, Olsson LM. Recent advances in the genetics of autoimmune disease. *Annu Rev Immunol* 2009;27:363-91.
 27. McAllister K, Eyre S, Orozco G. Genetics of rheumatoid arthritis: GWAS and beyond. *Open Access Rheumatol* 2011;3:31-46.
 28. Hughes LB, Reynolds RJ, Brown EE, Kelley JM, Thomson B, Conn DL, et al. Most common single-nucleotide polymorphisms associated with rheumatoid arthritis in persons of European ancestry confer risk of rheumatoid arthritis in African Americans. *Arthritis Rheum* 2010;62(12):3547-53.
 29. Mizuno A, Yoshida S, Ikari K, Toyama Y, Taniguchi A, Yamanaka H, et al. THU0458 analysis of genetic factors associated with progression of functional disability in Japanese patients with rheumatoid arthritis. *Annals of the Rheumatic Diseases* 2014;73(Suppl 2):341-2.
 30. Ehtesham N, Alani B, Mortazavi D, Azhdari S, Kenarangi T, Esmaeilzadeh E, et al. Association of rs3135500 and rs3135499 polymorphisms in the microRNA-binding site of nucleotide-binding oligomerization domain 2 (NOD2) gene with susceptibility to rheumatoid arthritis. *Iran J Allergy Asthma Immunol* 2021;20(2):178-87.
 31. Nasrollahzadeh Sabet M, Nasrabadi N, Jalili Z, Pakzad B, Davar S, Ehtesham N, et al. Association of three functional polymorphisms in the NLRP3 gene with susceptibility to rheumatoid arthritis in Iranian population. *Iran J Immunol* 2021;18(3):249-58.
 32. Hassani M, Dehani M, Rafie MZ, Esmaeilzadeh E, Davar S, Pakzad B, et al. Investigation of rs531564 polymorphism in the primary microRNA-124 gene in patients with systemic lupus erythematosus and rheumatoid arthritis: association with disease susceptibility and clinical characteristics. *Iran J Allergy Asthma Immunol* 2021;20 (3):303-13.