



Bioinformatic Investigation of Micro RNA-802 Target Genes, Protein Networks, and Its Potential Prognostic Value in Breast Cancer

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Abstract

Background: An increasing number of studies have suggested that unveiling the molecular network of miRNAs may provide novel therapeutic targets or biomarkers. In this study, we investigated the probable molecular functions that are related to miRNA-802 (miR-802) and evaluated its prognostic value in breast cancer utilizing bioinformatics tools.

Methods: PPI network, pathway enrichment and transcription factor analysis were applied to obtain hub genes among overlapping genes of four miRNA target prediction databases. Prognosis value assessments and expression analysis of hub genes using bioinformatics tools, as well as their literature validation were performed.

Results: Our results showed a significant correlation of the miR-802 overexpression with poor patient survival rate (BC, $p=2.7e-5$). We determined 247 target genes significant for GO and KEGG terms. Analysis of TFs by TRUST showed that RUNX3, FOXO3, and E2F1 are possible TFs that regulate the miR-802 expression and target genes network. According to our analysis; 21 genes might have an important function in miR-802 molecular processes and regulatory networks. The result shows that among these 21 genes, 8 genes (*CASC3*, *ITGA4*, *AGO3*, *TARDBP*, *MED13L*, *SF1*, *SNRPE* and *CRNKL1*) are positively correlated with patient survival. Therefore these genes could be considered and experimentally evaluated as a prognostic biomarker for breast cancer.

Conclusion: The comprehensive bioinformatics study on miR-802 target genes provided insight into miR-802 mediated pathways and processes. Furthermore, representing candidate target genes by prognostic values indicates the potential clinical application of miR-802 in breast cancer.

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Introduction

By estimating 276,480 new cases in 2020 in the United States, breast cancer will be the most expected diagnosed malignancy in women that accounts for 30% of all new cancer occurrences in women. In addition, breast cancer will be the second cause of cancer deaths in women by estimating 42,690 deaths¹. Despite all progression in diagnostic and therapeutic approaches for breast cancer, it is estimated by WHO that the total

worldwide cases will increase from 2,069,792 to 2,778,850 cases between 2018 and 2040, that shows the necessity of more investigation on the molecular mechanisms of the disease as well as detecting more efficient biomarkers².

Recently molecular approaches have been mentioned for breast cancer classifications and understanding its underlying tumorigenesis mechanisms³. Since

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now, alteration in signaling pathways, dysregulation of cells proliferation or apoptosis, mutation of oncogenes, different tumor metabolism, epithelial-to-mesenchymal transition, and Breast Cancer Stem Cells (BCSCs) development are among the most outlined molecular mechanisms⁴⁻⁷.

Also many treatments, survival or follow-up investigations are fulfilled based on the molecular pattern of the patients^{8,9}. In this regard, prognostic markers are so important due to their use in the estimation of disease recurrence and treatment response. Predictions based on prognostic markers are used to determine the proper treatment strategy that has a direct effect on patient's life prolongation as well as recurrence-free or overall survival¹⁰. A systematic review has demonstrated that the expression of *PI3K*, *TIMP-1*, *CEACAM6*, and aromatase are some reliable prognostic biomarkers for breast cancer¹¹. However, the poor outcome of the patients, especially in the advanced stage, reflects the need for more investigation for prognostic markers which are more clinically applicable¹².

Since the discovery of 22 nucleotides noncoding microRNAs about two decades ago, the role of these molecules has been highlighted in different biological processes such as development, differentiation, apoptosis, cell proliferation, and so on¹³⁻¹⁵. The microRNAs biological effect is exerted by post-translation regulation of about 30% of human genes, half of which are considered tumor-associated¹⁶. Due to their fundamental regulatory roles, dysregulation of microRNAs profile may result in malignancies including oncogenic transformation¹⁷. Regarding breast cancer, many studies have been conducted to identify diagnostic, prognostic, stage discriminating, or metastasis predictive biomarkers using aberrant expression profiles of microRNAs as well¹⁸⁻²¹.

Recently it has been reported that an emerging microRNA, miR-802, has shown important regulatory roles in many malignancies including breast cancer. Previous investigations have revealed dis-regulation of miR-802 in the liver, gastric and cervical cancer²²⁻²⁴. It has been reported that miR-802 inhibits epithelial-mesenchymal transition in human prostate cancer by targeting flotillin-2²⁵. Moreover, miR-802 targeted *ZNF521* gene and suppressed the malignant progression of hepatocellular carcinoma²⁶. A study in breast cancer has demonstrated that miR-802 inhibits proliferation through the suppression of FoxM1²⁷. Regarding the prognostic value of miR-802, it is reported that patients with hepatocellular carcinoma and prostate cancer that have lower expression of miR-802 showed better survival²⁸. Despite the researches for determining miR-802 roles, there are not enough reports about the roles of miR-802 in the molecular basis of diseases and its clinical potential in malignancies as well as breast cancer. Therefore more investigations are required to shed more light on the miR-802 regulatory network and its potential prognostic value²⁷.

In the past several years, bioinformatic analysis of large-scale gene expression and clinical data have been an effective and applicable tool for investigating signaling pathways, hub genes, or tumorigenesis mechanisms in different cancers²⁹⁻³¹. Analyzing biological data by using computational and bioinformatic tools and websites is a useful approach for discovering new biomarkers that could have potential importance in clinical research and routine³². Determining prognostic biomarkers based on bioinformatic approaches is among the recent interesting investigation areas and is performed for different types of cancers³³⁻³⁵.

In this study, we explored the miR-802 target genes, signaling pathways protein-protein interaction network, and key cluster and hub genes via bioinformatics tools. Then the resulted hub genes were assessed to predict their potential biomarker value for prognosis in breast cancer. Figure 1 shows the workflow of the present study.

Materials and Methods

Evaluation of the prognostic value of miR-802

Pan-cancer survival analysis of miR-802 in TCGA dataset: Analysis of pan-cancer overall survival, utilizing pan-cancer TCGA miRNA database was performed by Kaplan-Meier Plotter ([HTTP:// kmplot.com/analysis/](http://kmplot.com/analysis/))^{36,37}. Kaplan Meier plotter performs its assessments based on mRNA, miRNA, protein content of cancer samples. One of the sources of the Kaplan Meier plotter is the TCGA (The Cancer Genome Atlas) Dataset³⁸. TCGA contains a molecular characterization of over 20,000 primary cancerous and matched normal samples of 33 cancer types and has generated over 2.5 petabytes of genomic, epigenomic, transcriptomic, and proteomic (<https://www.cancer.gov/tcga>). We used the survival data of breast cancer (BRCA) tissues of 1078 patients from TCGA dataset (n=1078, BRCA). All molecular subtypes of breast cancers have been includ-

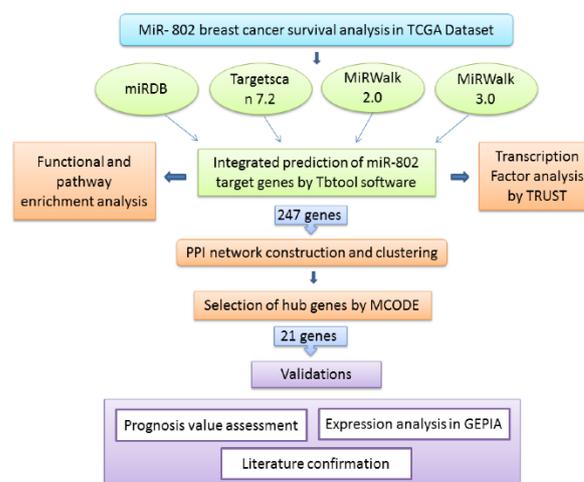


Figure 1. The workflow of the current study. This figure is a graphical overview of datasets, software and bioinformatic analysis that was exploited to investigate miR-802 target genes.

ed and the overall survival of the patients was assessed. Split of the patients for low expression or high expression of miRNA was performed based on auto select best cutoff mode. In this way, all possible cut-off values between the lower and upper quartiles are computed and the best performing threshold is used. The false discovery rate of auto-selected cut-off was 1%. Log-rank p-value <0.05 is considered a statistically significant finding, and the hazard ratio is calculated with 95% confidence interval.

Integrated prediction of miR-802 target genes

We first used four highly used and well-known tools for the prediction of miR-802 target genes (miRDB: <http://mirdb.org/>, Targetscan7.2: <http://www.targetscan.org/>, MiRWalk 3.0: <http://mirwalk.umm.uni/>, and MiRWalk 2.0: <http://zmf.umm.uni-heidelberg.de/mirwalk2>). In the second step, we adopted an integrated approach to select target genes from mentioned databases. In order to get an insight of all possible molecular functions of a miRNA, the range of considered target genes should be comprehensive and wide enough. However, adopting a suitable approach would be challenging. Integrating the data of different research tools is becoming an interesting approach for the prediction of miRNA target genes^{39,40}. Due to large number of predicted genes in every database, while a target gene is annotated in more than one reliable database, it is usually a robust and valid one. In this regard we used TBtool software, a user-friendly toolkit, to analyze and visualize overlapping targets of miR-802 from four prediction data bases⁴¹. Venn diagram and Upset Plot from TBtool were utilized to display the intersection of target genes between four microRNA predictive databases.

Gene ontology (GO) and kyoto encyclopedia of genes and genomes (KEGG) pathway enrichment analysis

The overlapping target genes selected by TBtool software were then analyzed by Metascape (metascape.org/gp/index.html) for GO and KEGG Pathway Enrichment Analysis. Metascape is a web-based portal that retrieves information from the latest version of databases and provides functional enrichment, gene annotation, interaction analysis, and so on to interpret OMICS-based studies⁴². To GO and KEGG pathway enrichment analysis in this study, the names of the overlapped gene were inputted in Metascape online tool and the result was exported. GO analysis included Biological Processes (BP); Molecular Functions (MF) and Cellular Components (CC). p-value <0.05 was considered statistically significant for GO and KEGG pathway enrichment analysis. Transcription Factors (TFs) related to target genes were retrieved from TRUST (version 2.0) database which contains the data of regulatory relationship of Transcription factors⁴³. TRUST analysis was performed on terms with a p-value <0.01 which were collected and grouped into clusters. The trust data is accessible from Metascape as well.

Protein-protein interaction (PPI) network

STRING database (STRING, <https://string-db.org/>) version 11 was used to analyze the Protein-Protein Interaction (PPI) of overlapping genes. A confidence score higher than 0.4 was set to build the interaction network. The PPI network of genes was then extracted for more analysis by Cytoscape software (version 3.7.1). Potential clusters and modules of the PPI network were screened by Molecular Complex Detection (MCODE) plugin Cytoscape. MCODE finds highly interconnected regions in PPI network. Detected modules with MCODE score >3 and nodes number >2 were presented.

Analyzing the overall survival rate and expression of target genes

To analyze the potential prognostic value of selected target genes obtained from GO and network analysis, overall survival analysis was carried out by submitting the selected gene names in the Kaplan-Meier Plotter mRNA data set (p-value adjustment, 0.05 significance). Expression analysis of the selected genes was performed by the GEPIA database as well. p-value <0.05 was considered significant for mean expression differences between normal and tumor samples of breast cancers. GEPIA database used the expression data of 1085 tumor samples and 291 normal samples of breast cancer patients retrieved from TCGA.

Results

Prognostic values of miR-802 in breast cancer and *in silico* exploration of its target genes: To evaluate the prognostic values of miR-802 in breast cancers, a breast-cancer survival analysis was performed based on the breast-cancer TCGA miRNA database. The results showed that the high expression of miR-802 was significantly correlated with poor patient survival rate (BC, p=2.7e-5) which demonstrates its biological importance (Figure 2). Based on this analysis, miR-802 and its target genes might have the potential to be considered as a biomarker for breast cancer prognosis.

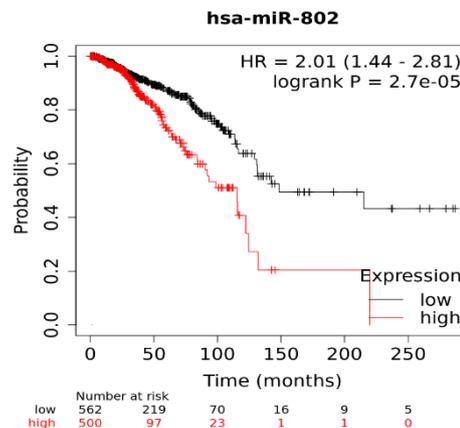


Figure 2. Overall Survival (OS) of miR-802 based on TCGA miRNA in breast cancer.

To figure out more about the roles of miR-802 in the biological processes of cells and its regulatory network, the predicted targets of miR-802 were retrieved from miRDB, Targetscan, miRwalk 2, and miRwalk 3. A set of genes was predicted by every database that intersection genes of these target genes were visualized by TBtool software in figure 3A (Figure 3A). This software perfectly provides clusters between databases and each cluster gene can be easily extracted from software. As is displayed in figure 3A, many genes have been annotated in more than one database. Prediction of a specific gene in different databases illustrated the higher probability of that gene to be the accurate target of miR-802. Therefore we selected overlapping genes, and in order not to miss any possible information, we chose every overlapping gene between two databases and more. In this way, 247 genes were selected for further analyses which are shown in figure 3B (Figure 3B).

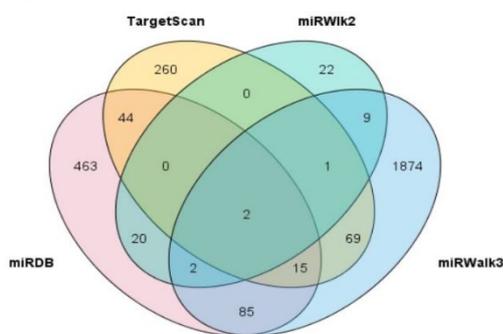


Figure 3A. Venn diagram shows the intersection of target genes that were retrieved from four miRNA target prediction tools (miRDB, TargetScan 7.2, miRWalk2, and miRWalk3).

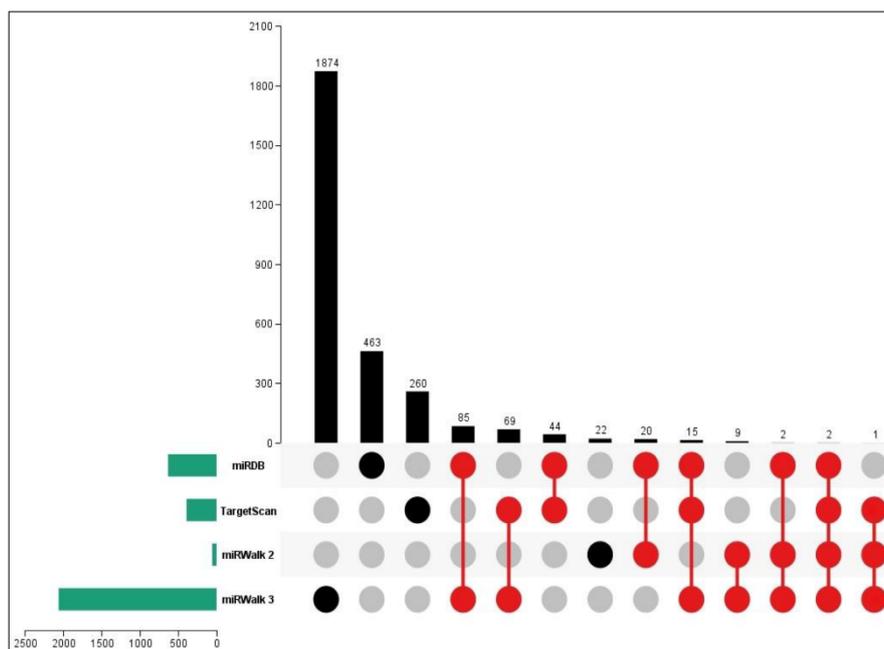


Figure 3B. Upset Plot displayed Overlapped target genes between four databases which were 247 genes in total and are marked in red.

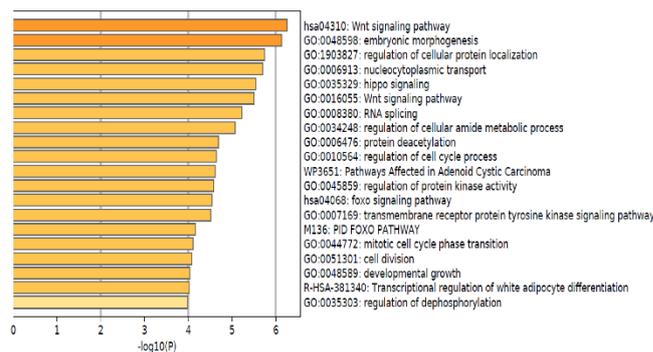


Figure 4. Bar chart of enriched terms resulted from gene ontology analysis.

Functional analyses of the predicted target genes

For GO annotation and KEGG Enrichment analyses of 247 selected target genes of miR-802, the Metascape which is an online functional enrichment tool, was exploited. p-values <0.05 were considered significant for enriched terms. Figure 4 shows the result of the molecular function gene ontology of 247 target genes. For GO enrichment analysis on the MF level, the selected target genes were mainly enriched in transcription coregulator activity, kinase regulator activity, chromatin binding and mRNA binding. On the BP level, the genes were mainly enriched in embryonic morphogenesis, regulation of cell cycle process, regulation of protein kinase activity, transmembrane receptor protein tyrosine kinase signaling pathway, regulation of cellular protein localization and Wnt signaling pathway. On the CC level, the genes were mainly enriched in axon,

dendrite, glutamatergic synapse, spliceosomal complex and cytoplasmic ribonucleoprotein granule. Other significantly enriched GO terms are represented in table 1. The most significant result of GO and KEGG pathway enrichment along with two other databases has been Wnt signaling pathway, foxo signaling pathway, Pathways Affected in Adenoid Cystic Carcinoma, and regu-

lation of dephosphorylation (Table 1). Analysis of TFs by TRUST showed that RUNX3, FOXO3, and E2F1 are possible TFs that regulate miR-802 expression and its target genes network (Figure 5).

PPI network construction and survival analysis of clustered genes

247 overlapping target genes included in PPI Net-

Table 1. Summary of pathway and GO enrichment analysis

| ID | Category | Description | Count | % | Log10(P) |
|--|-------------------------|--|-------|------|----------|
| Enriched Pathways | | | | | |
| hsa04310 | KEGG Pathway | Wnt signaling pathway | 10 | 4.07 | -6.25 |
| hsa04068 | KEGG Pathway | foxo signaling pathway | 8 | 3.25 | -4.54 |
| WP3651 | WikiPathways | Pathways Affected in Adenoid Cystic Carcinoma | 6 | 2.44 | -4.61 |
| M136 | Canonical Pathways | PID FOXO pathway | 5 | 2.03 | -4.17 |
| Enriched GO Biological Processes (BP) | | | | | |
| GO:0048598 | GO Biological Processes | Embryonic morphogenesis | 19 | 7.72 | -6.14 |
| GO:1903827 | GO Biological Processes | Regulation of cellular protein localization | 18 | 7.32 | -5.74 |
| GO:0006913 | GO Biological Processes | Nucleocytoplasmic transport | 14 | 5.69 | -5.69 |
| GO:0035329 | GO Biological Processes | Hippo signaling | 6 | 2.44 | -5.53 |
| GO:0016055 | GO Biological Processes | Wnt signaling pathway | 17 | 6.91 | -5.50 |
| GO:0008380 | GO Biological Processes | RNA splicing | 16 | 6.50 | -5.23 |
| GO:0034248 | GO Biological Processes | Regulation of cellular amide metabolic process | 16 | 6.50 | -5.07 |
| GO:0006476 | GO Biological Processes | Protein deacetylation | 7 | 2.85 | -4.69 |
| GO:0010564 | GO Biological Processes | Regulation of cell cycle process | 20 | 8.13 | -4.64 |
| GO:0045859 | GO Biological Processes | Regulation of protein kinase activity | 20 | 8.13 | -4.57 |
| GO:0007169 | GO Biological Processes | Transmembrane receptor protein tyrosine kinase signaling pathway | 19 | 7.72 | -4.51 |
| GO:0044772 | GO Biological Processes | Mitotic cell cycle phase transition | 16 | 6.50 | -4.11 |
| GO:0051301 | GO Biological Processes | Cell division | 16 | 6.50 | -4.07 |
| GO:0048589 | GO Biological Processes | Developmental growth | 16 | 6.50 | -4.03 |
| GO:0035303 | GO Biological Processes | Regulation of dephosphorylation | 9 | 3.66 | -3.99 |
| Enriched GO Molecular Functions (MF) | | | | | |
| GO:0003712 | GO Molecular Functions | Transcription coregulator activity | 17 | 6.91 | -5.89 |
| GO:0019207 | GO Molecular Functions | Kinase regulator activity | 10 | 4.07 | -4.59 |
| GO:0003682 | GO Molecular Functions | Chromatin binding | 16 | 6.50 | -4.44 |
| GO:0003729 | GO Molecular Functions | mRNA binding | 15 | 6.10 | -4.04 |
| GO:0008234 | GO Molecular Functions | Cysteine-type peptidase activity | 7 | 2.85 | -3.01 |
| GO:0120227 | GO Molecular Functions | Acyl-CoA binding | 3 | 1.22 | -2.88 |
| GO:0047485 | GO Molecular Functions | Protein N-terminus binding | 5 | 2.03 | -2.62 |
| GO:0016301 | GO Molecular Functions | Kinase activity | 15 | 6.10 | -2.61 |
| Enriched GO Cellular Components (CC) | | | | | |
| GO:1902911 | GO Cellular Components | Protein kinase complex | 8 | 3.25 | -5.42 |
| GO:0030424 | GO Cellular Components | Axon | 17 | 6.91 | -4.44 |
| GO:0005681 | GO Cellular Components | Spliceosomal complex | 9 | 3.66 | -4.30 |
| GO:0036464 | GO Cellular Components | Cytoplasmic ribonucleoprotein granule | 9 | 3.66 | -3.64 |
| GO:0030425 | GO Cellular Components | Dendrite | 15 | 6.10 | -3.44 |
| GO:0098978 | GO Cellular Components | Glutamatergic synapse | 10 | 4.07 | -3.39 |
| GO:0005923 | GO Cellular Components | Bicellular tight junction | 6 | 2.44 | -3.23 |

The count stands for the number of miR-802 target genes which are involved in the enriched term.

% stands for percent of gene per enriched term.

Log10(P): stands for the p-value regarding each term enrichment.

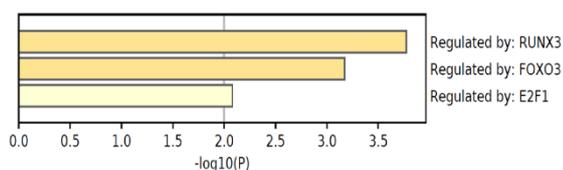


Figure 5. predicted transcription factors that regulate miR-802 target gene network.

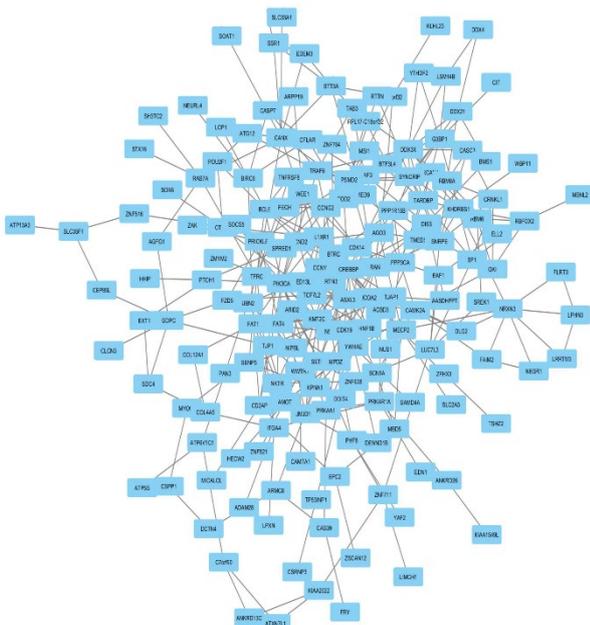


Figure 6. The PPI network of 247 selected target genes of miR-802. Nodes and edges represent genes and interactions between them respectively.

work exploration *via* the STRING database with a medium confidence score (interaction score >0.400). The resulted network is displayed in figure 6 which has 247 nodes and 366 edges. PPI enrichment p-value is 6.46e-10. Three clusters with highly interconnected regions were extracted from PPI network by the MCODE clustering plug-in with MCODE score >3 and nodes number >2. Highly connected genes in a network mostly played an important role in the biological processes and would be taken as hub genes. 21 genes were involved in the currents study resulted clusters and considered as hub genes. Figure 7 shows the three selected clusters and their involved genes. It is supposed that these 21 genes might have an important function in miR-802 molecular processes and regulatory networks. Therefore the prognostic value of them in breast cancer was evaluated by Kaplan-Meier plotter (<http://kmplot.com/analysis/>) on gene expression data of 1880 breast cancer patients. The result show that among these 21 genes, high expression of *CASC3* (207842_s_at), *ITGA4* (213416_at), *AGO3* (EIF2C3/219426_at), *TARDBP* (221264_s_at), *MED13L* (THRAP2/ 212209_at) and *SFI* (ZNF162/

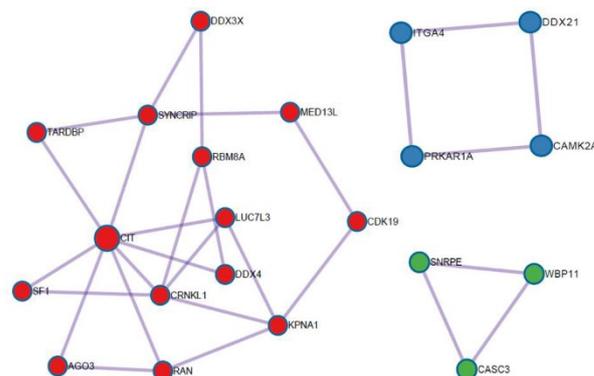


Figure 7. Clusters extracted from PPI network by MCODE. Three clusters and their involved genes are shown in different colors.

208313_s_at) genes are positively correlated with patient survival, while high expression of *SNRPE* (203316_s_at) and *CRNKL1* (219913_s_at) indicate poor patients survival (Figure 8). It seems these 8 genes could be considered and experimentally evaluated as a prognostic biomarker for breast cancer. Moreover, expression analysis in the GEPIA database showed that the mean expression of 12 genes (*CASC3*, *MED13L*, *SFI*, *CRNKL1*, *SNRPE*, *SYNCYRIP*, *RBM8A*, *RAN*, *CIT*, *LUC7L3*, *WBP11*, *KPN1A1*) out of 21 selected genes, is significantly (p-value <0.05) different between normal and tumor breast samples, and 5 genes with prognostic value are among differentially expressed genes as well (Figure 9). The differential expression pattern of most of miR-802 hub genes in breast cancer samples, made the probable role of this microRNA in breast cancer development stronger.

Discussion

MicroRNAs play several key regulatory roles in biological processes⁴⁴. Similarly, miR-802 has shown multi functions in different conditions. It had been reported that miR-802 is involved in several malignancies such as liver cancer²², prostate cancer⁴⁵, gastric cancer²³, cervical cancer²⁴, ovarian cancer⁴⁶, pancreatic cancer⁴⁷, and so on. Other studies had shown the association of miR-802 with impaired glucose metabolism and obesity^{48,49}. There were also some reports of physiological functions of miR-802 in kidney or intestine development^{50,51}. In consistent with previous studies, the *in silico* investigation of the current study showed similar results that would be argued in the following. Deciphering miR-802 target genes in the large picture, we applied an integrated approach for target prediction from four miRNA-Target gene prediction tools that resulted in 247 selected genes. The most significant GO terms that was enriched from these selected genes, were Wnt signaling pathway (hsa04310, p-value: -6.25), regulation of cell cycle process morphogenesis (GO:0010564, p-value: -4.64), and FOXO path-

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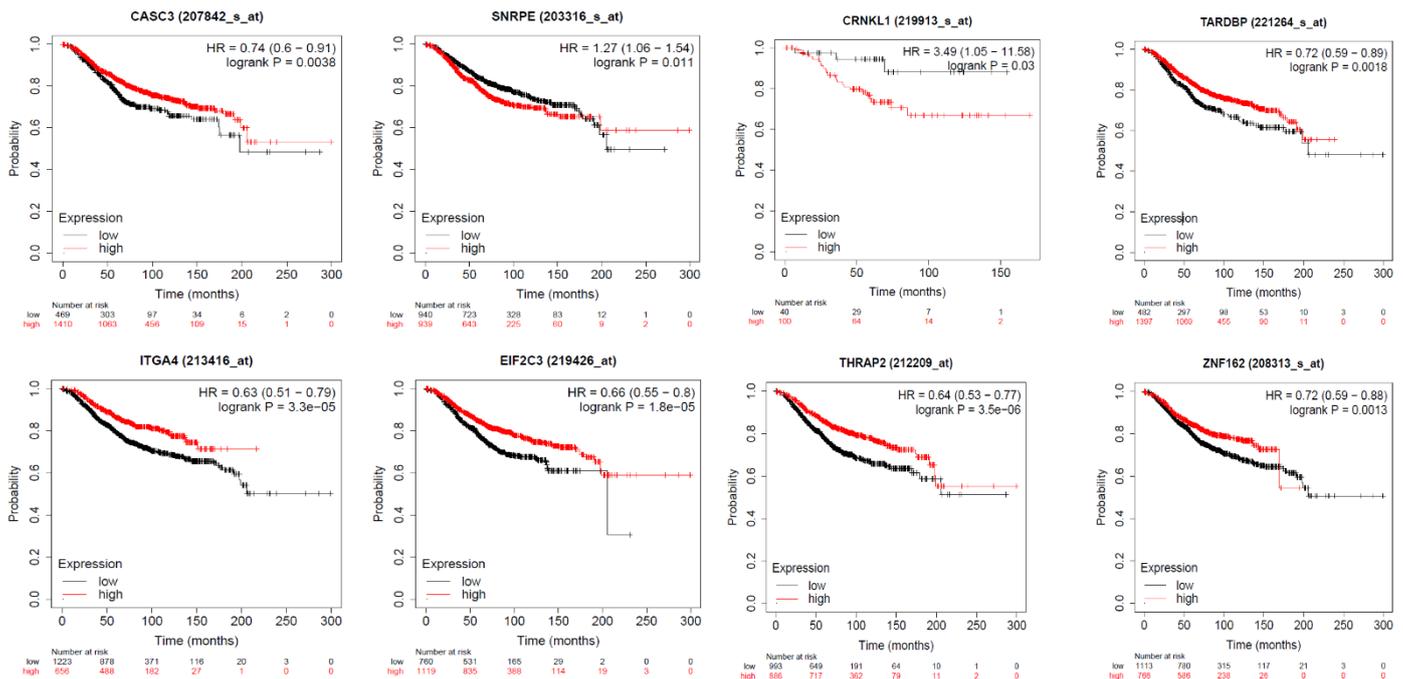


Figure 8. Prognostic value of miR-802 target genes. These 8 genes have better prognosis in breast cancer patients.

way (hsa04068, p-value: -4.54) which all are among the molecular mechanisms of cancer development that suggest the potential role of miR802 in cancer progression and probable pathways that this micro RNA plays role in them⁵²⁻⁵⁴. In addition to our results, there are other evidences that support the role of miR-802 in Wnt signaling pathway which verify the reliability of our results. It is reported that miR-802 and miR-1 regulate mesenchymal-epithelial transition during kidney development by regulation of Wnt-4/ β -catenin signaling⁵⁵. In addition, it has been shown that Tmed9, a modulator of Wnt and lysozyme/defensin secretion in the mouse small intestine, as well as Fzd5 and Tcf4, the downstream components of Wnt signaling, are targeted and suppressed by miR-802⁵⁶.

As illustrated in the results, E2F1 was identified as one of the transcription factors that regulate miR-802 expression. The important issue that should be mentioned here is the known regulatory effect of E2F1 on several genes involved in cell cycle which is verified in different literature⁵⁷⁻⁵⁹. On the other hand *in silico* findings of the current study demonstrated that miR-802 played role in the cell cycle process (GO:0010564), mitotic cell cycle phase transition (GO:0044772) and cell division (GO:0051301). Consistent with our results, there was also a report on effect of miR-802 in inhibiting cancer cell proliferation by targeting FOXM1 and suppressing Cyclin A and Cyclin B1 which are key regulators of cell-cycle progression²⁷. Taken together, it seems that miR-802 is an effective microRNA in the

cell cycle that its regulatory network could be further explored.

FOXO (Forkhead box O) is a transcription factor that plays important role in fundamental cellular processes⁶⁰. The result of this study indicated that FOXO is another transcription factor of miR-802 and demonstrated its role in FOXO signaling pathways (hsa-04068). Consistent with our result, it is reported in another study that FOXO regulates transcription of miR-802 which supports our finding⁴⁹.

Among other terms that are shown in figure 4, were transcriptional regulation of white adipocyte differentiation (R-HAS-381340) and regulation of dephosphorylation (GO:0035303); both of which are involved in cellular metabolisms⁶¹⁻⁶³. In line with our results, it has been demonstrated that dysregulated miR-802 is critically associated with glucose metabolisms, obesity and some of their most important related conditions^{48,64}. Hence, by considering the results of the bioinformatics ontology study of miR-802 target genes and the consistency of the results with the literature, it could be validated that the platform of the study and integrated target prediction was promising and applicable approach.

Regarding the prognosis value of candidate genes, our result using bioinformatics tools demonstrated that *CASC3*, *ITGA4*, *AGO3*, *TARDBP*, *MED13L*, *SFI*, *SNRPE* and *CRNK1* could have prognostic value in breast cancer. Supporting our findings, the prognostic value of most of these genes has been reported in breast can-

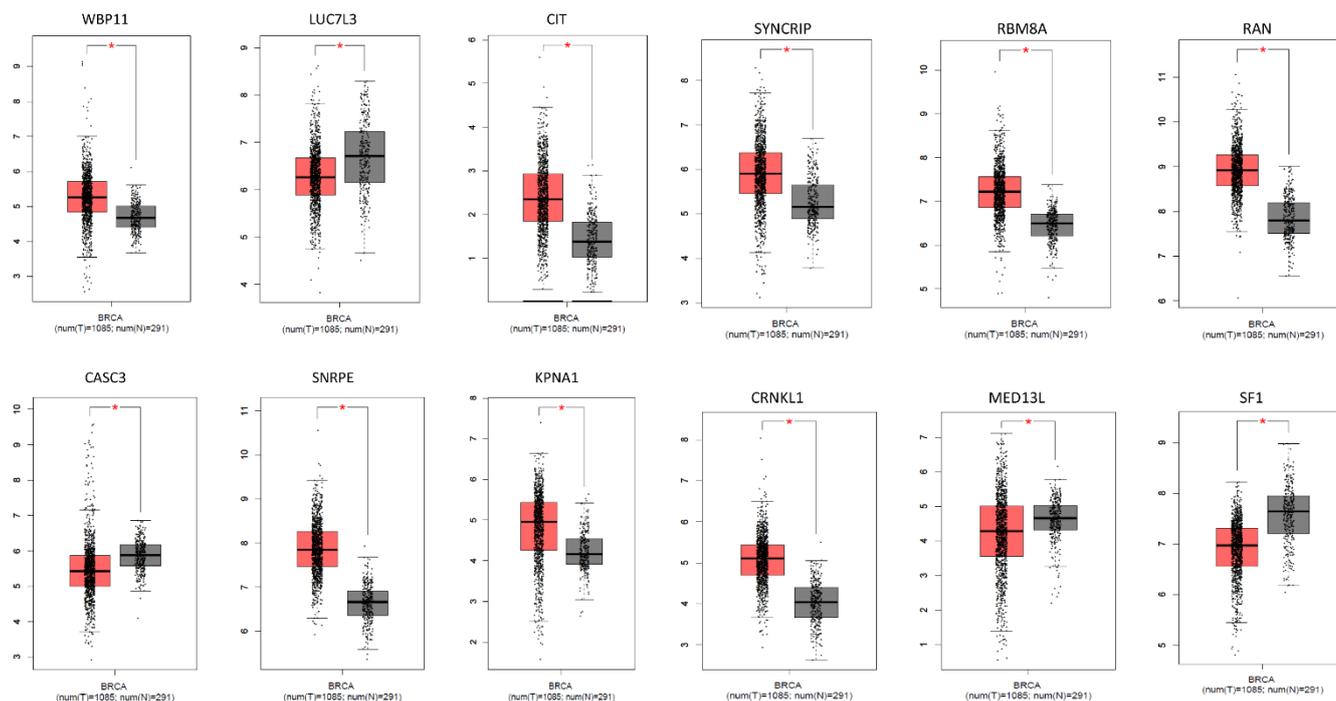


Figure 9. Gene expression analysis of selected hub genes. The red color bar represents gene expression in tumor samples (n=1085) and the gray color bar represents gene expression in normal samples (n=291). BRCA: breast cancer, (p-value < 0.05).

cer or other malignancies, either *in silico* or experimental. In agreement with our result, another study on the RNA network of triple negative breast cancer has verified that *TARDBP* had prognostic value⁶⁵. Furthermore, another *in silico* study suggested the prognostic value of *ITGA4* in basal-like and HER2+ breast cancer⁶⁶. In an integrated study utilizing public bioinformatics datasets, *ITGA4* has been identified as a prognostic marker of early ovarian cancer⁶⁷. Other studies also demonstrated a significant association of *SFI* with the overall survival of pancreatic cancer through aberrant alternative splicing events⁶⁸. Moreover, *SFI* regulated interferon λ which had prognostic value in many cancers⁶⁹. In lung cancer, *SNRPE* and *MED13L* had been reported as potential prognostic markers^{70,71}. Furthermore, it has been indicated that *CASC3*, *AGO3* and *TARDBP* could have a significant role in the prognosis of gastric cancer, hepatocellular carcinoma and cervical cancer respectively⁷²⁻⁷⁴.

Regarding 21 selected target genes of miR-802 in PPI clusters, there are several articles indicating that almost all of these genes are involved in different malignancies including breast cancer. Some of the genes that have evidence in breast cancer are *DDX21* which regulates epithelial-mesenchymal transition in breast cancer⁷⁵, *LUC7L3* inhibits breast cancer progression⁷⁶, *CDK19* plays regulatory function in triple-negative breast cancer⁷⁷, downregulation of Ran GTPase limits proliferation and migration of breast cancer cells⁷⁸,

KPNA1 involves in tamoxifen resistance⁷⁹, *DDX3X* affects breast cancer cell cycle⁸⁰ and so on.

About molecular function of selected genes in other cancers we can mention, *WBP11* that inhibits gastric cancer migration⁸¹, *AGO3* that regulates the Wnt/ β -catenin signaling pathway in cervical cancer⁸², *MED13L* which is functional in lung cancer radiosensitivity⁷¹, *SYNCRIP* that is involved in poor prognosis of pancreatic cancer⁸³ and Down-regulation of *CIT* which limits human bladder cancer cells proliferation⁸⁴. All of these make the possible role of miR-802 in breast cancer more strong, therefore the regulatory network and prognostic value of its target genes can be further studied in breast cancer.

Conclusion

This study provides comprehensive insight into miR-802 functional roles and regulatory networks. Besides the survival analysis provides promising candidate targets for the prognosis of breast cancer. Additionally, it is demonstrated that the exploitation of bioinformatics tools could be a reliable and effective approach in medical investigations.

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Conflict of Interest

The authors declare that they have no competing interests.

References

- Ruhl J, Callaghan C, Hurlbut A. Cancer facts & figures 2020. American Cancer Society. 2020.
- WHO. WHO-CancerReport-2020-Global Profile. WHO. 2020.
- do Nascimento RG, Otoni KM. Histological and molecular classification of breast cancer: what do we know? *Mastology* 2020;30:e20200024.
- Feng Y, Spezia M, Huang S, Yuan C, Zeng Z, Zhang L, et al. Breast cancer development and progression: Risk factors, cancer stem cells, signaling pathways, genomics, and molecular pathogenesis. *Genes Dis* 2018;5(2):77-106.
- Kuo MH, Chang WW, Yeh BW, Chu YS, Lee YC, Lee HT. Glucose transporter 3 is essential for the survival of breast cancer cells in the brain. *Cells* 2019;8(12):1568.
- Tungskruthai S, Petpiroon N, Chanvorachote P. Molecular mechanisms of breast cancer metastasis and potential anti-metastatic compounds. *Anticancer Res* 2018;38(5):2607-18.
- Scioli MG, Storti G, D'Amico F, Gentile P, Fabbri G, Cervelli V, et al. The role of breast cancer stem cells as a prognostic marker and a target to improve the efficacy of breast cancer therapy. *Cancers (Basels)* 2019;11(7):1021.
- Munoz DF, Plevritis SK. Estimating breast cancer survival by molecular subtype in the absence of screening and adjuvant treatment. *Med Decis Making* 2018;38(1_suppl):32S-43S.
- Howlander N, Cronin KA, Kurian AW, Andridge R. Differences in breast cancer survival by molecular subtypes in the United States. *Cancer Epidemiol Biomarkers Prev* 2018;27(6):619-26.
- Mohebian MR, Marateb HR, Mansourian M, Mañanas MA, Mokarian F. A hybrid computer-aided-diagnosis system for prediction of breast cancer recurrence (HPBCR) using optimized ensemble learning. *Comput Struct Biotechnol J* 2017;15:75-85.
- Kutomi G, Mizuguchi T, Satomi F, Maeda H, Shima H, Kimura Y, et al. Current status of the prognostic molecular biomarkers in breast cancer: A systematic review. *Oncol Lett* 2017;13(3):1491-8.
- Walaszczyk A, Gabryś D. Molecular markers used in breast cancer diagnosis—current practice and future perspectives. *Nowotwory J Oncol* 2018;68(5-6):259-67.
- Lee R, Feinbaum R, Ambros V. The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell* 1993;75(5):843-54.
- He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev Genet* 2004;5(7):522-31.
- Giovannetti E, Erozeñci A, Smit J, Danesi R, Peters GJ. Molecular mechanisms underlying the role of microRNAs (miRNAs) in anticancer drug resistance and implications for clinical practice. *Crit Rev Oncol Hematol* 2012;81(2):103-22.
- Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noch E, Yendamuri S, et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Natl Acad Sci* 2004;101(9):2999-3004.
- Jansson MD, Lund AH. MicroRNA and cancer. *Mol Oncol* 2012;6(6):590-610.
- Mattie MD, Benz CC, Bowers J, Sensinger K, Wong L, Scott GK, et al. Optimized high-throughput microRNA expression profiling provides novel biomarker assessment of clinical prostate and breast cancer biopsies. *Mol Cancer* 2006;5:24.
- Lowery AJ, Miller N, Devaney A, McNeill RE, Davoren PA, Lemetre C, et al. MicroRNA signatures predict oestrogen receptor, progesterone receptor and HER2/neu receptor status in breast cancer. *Breast Cancer Res* 2009;11(3):R27.
- Bertoli G, Cava C, Castiglioni I. MicroRNAs: new biomarkers for diagnosis, prognosis, therapy prediction and therapeutic tools for breast cancer. *Theranostics* 2015;5(10):1122-43.
- Fridrichova I, Zmetakova I. MicroRNAs contribute to breast cancer invasiveness. *Cells* 2019;8(11):1361.
- Tao J, Ji J, Li X, Ding N, Wu H, Liu Y, et al. Distinct anti-oncogenic effect of various microRNAs in different mouse models of liver cancer. *Oncotarget* 2015;6(9):6977-88.
- Zhang X, Mu J, Liu L, Zhang H. Upregulation of miR-802 suppresses gastric cancer oncogenicity via targeting RAB23 expression. *Eur Rev Med Pharmacol Sci* 2017;21(18):4071-8.
- Zhang Q, Lv R, Guo W, Li X. microRNA-802 inhibits cell proliferation and induces apoptosis in human cervical cancer by targeting serine/arginine-rich splicing factor 9. *J Cell Biochem* 2019;120(6):10370-9.
- Wang D, Lu G, Shao Y, Xu D. microRNA-802 inhibits epithelial-mesenchymal transition through targeting flotillin-2 in human prostate cancer. *Biosci Rep* 2017;37(2):BSR20160521.
- Yang N, Wang L, Chen T, Liu R, Liu Z, Zhang L. ZNF521 which is downregulated by miR-802 suppresses malignant progression of Hepatocellular Carcinoma through regulating Runx2 expression. *J Cancer* 2020;11(19):5831-9.

27. Yuan F, Wang W. MicroRNA-802 suppresses breast cancer proliferation through downregulation of FoxM1. *Mol Med Rep* 2015;12(3):4647-51.
28. Gao T, Zou M, Shen T, Duan S. Dysfunction of miR-802 in tumors. *J Clin Lab Anal* 2021;35(11):e23989.
29. Pappalardo F, Russo G, Candido S, Pennisi M, Cavalieri S, Motta S, et al. Computational modeling of PI3K/AKT and MAPK signaling pathways in melanoma cancer. *PLoS One* 2016;11(3):e0152104.
30. Underwood T. Pan-cancer analysis of whole genomes. *Nature* 2020;578(7793):82-93.
31. Deng JL, Xu YH, Wang G. Identification of potential crucial genes and key pathways in breast cancer using bioinformatic analysis. *Front Genet* 2019;10:695.
32. Fattahi F, Zanjani LS, Shams ZH, Kiani J, Mehrzama M, Najafi M, et al. High expression of DNA damage-inducible transcript 4 (DDIT4) is associated with advanced pathological features in the patients with colorectal cancer. *Sci Rep* 2021;11(1):13626.
33. Wang C, Yang Y, Yin L, Wei N, Hong T, Sun Z, et al. Novel potential biomarkers associated with epithelial to mesenchymal transition and bladder cancer prognosis identified by integrated bioinformatic analysis. *Front Oncol* 2020;10:931.
34. Zeng ML, Zhu XJ, Liu J, Shi PC, Kang YL, Lin Z, et al. An integrated bioinformatic analysis of the S100 gene family for the prognosis of colorectal cancer. *BioMed Res Int* 2020;2020:4746929.
35. Kudryavtseva AV, Lukyanova EN, Kharitonov SL, Nyushko KM, Krashennnikov AA, Pudova EA, et al. Bioinformatic identification of differentially expressed genes associated with prognosis of locally advanced lymph node-positive prostate cancer. *J Bioinform Comput Biol* 2019;17(01):1950003.
36. Györfy B, Lanczky A, Eklund AC, Denkert C, Budczies J, Li Q, et al. An online survival analysis tool to rapidly assess the effect of 22,277 genes on breast cancer prognosis using microarray data of 1,809 patients. *Breast Cancer Res Treat* 2010;123(3):725-31.
37. Lanczky A, Nagy Á, Bottai G, Munkácsy G, Szabó A, Santarpia L, et al. miRpower: a web-tool to validate survival-associated miRNAs utilizing expression data from 2178 breast cancer patients. *Breast Cancer Res Treat* 2016;160(3):439-46.
38. Chang K, Creighton C, Davis C, Donehower L. The cancer genome atlas pan-cancer analysis project. *Nat Genet* 2013;45(10):1113-20.
39. Quillet A, Saad C, Ferry G, Anouar Y, Vergne N, Lecroq T, et al. Improving bioinformatics prediction of microRNA targets by ranks aggregation. *Frontiers in genetics*. 2020;10:1330.
40. Liu C, Min L, Kuang J, Zhu C, Qiu XY, Zhu L. Bioinformatic identification of miR-622 key target genes and experimental validation of the miR-622-RNF8 axis in breast cancer. *Front Oncol* 2019;9:1114.
41. Chen C, Chen H, Zhang Y, Thomas HR, Frank MH, He Y, et al. TBtools: an integrative toolkit developed for interactive analyses of big biological data. *Mol Plant* 2020;13(8):1194-202.
42. Zhou Y, Zhou B, Pache L, Chang M, Khodabakhshi AH, Tanaseichuk O, et al. Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nat Commun* 2019;10(1):1523.
43. Han H, Cho JW, Lee S, Yun A, Kim H, Bae D, et al. TRRUST v2: an expanded reference database of human and mouse transcriptional regulatory interactions. *Nucleic Acids Res* 2018;46(D1):D380-D6.
44. Gebert LF, MacRae IJ. Regulation of microRNA function in animals. *Nat Rev Mol Cell Biol* 2019;20(1):21-37.
45. Wang D, Lu G, Shao Y, Xu D. microRNA-802 inhibits epithelial-mesenchymal transition through targeting flotillin-2 in human prostate cancer. *Biosci Rep* 2017;37(2).
46. Yang B, Sun L, Liang L. MiRNA-802 suppresses proliferation and migration of epithelial ovarian cancer cells by targeting YWHAZ. *J Ovarian Res* 2019;12(1):100.
47. Qin Y, Liu X, Pan L, Zhou R, Zhang X. Long noncoding RNA MIR155HG facilitates pancreatic cancer progression through negative regulation of miR-802. *J Cell Biochem* 2019;120(10):17926-34.
48. Kornfeld JW, Baitzel C, Könnner AC, Nicholls HT, Vogt MC, Herrmanns K, et al. Obesity-induced overexpression of miR-802 impairs glucose metabolism through silencing of Hnf1b. *Nature* 2013;494(7435):111-5.
49. Zhang F, Ma D, Zhao W, Wang D, Liu T, Liu Y, et al. Obesity-induced overexpression of miR-802 impairs insulin transcription and secretion. *Nat Commun* 2020;11(1):1-16.
50. Lin DH, Yue P, Pan C, Sun P, Wang WH. MicroRNA 802 stimulates ROMK channels by suppressing caveolin-1. *J Am Soc Nephrol* 2011;22(6):1087-98.
51. Sansom SE, Nuovo GJ, Martin MM, Kotha SR, Parinandi NL, Elton TS. miR-802 regulates human angiotensin II type 1 receptor expression in intestinal epithelial C2BBel1 cells. *Am J Physiol Gastrointest Liver Physiol* 2010;299(3):G632-G42.
52. Liu H, Song Y, Qiu H, Liu Y, Luo K, Yi Y, et al. Downregulation of FOXO3a by DNMT1 promotes breast cancer stem cell properties and tumorigenesis. *Cell Death Differ* 2020;27(3):966-83.
53. Yin P, Wang W, Zhang Z, Bai Y, Gao J, Zhao C. Wnt signaling in human and mouse breast cancer: focusing on Wnt ligands, receptors and antagonists. *Cancer Sci* 2018;109(11):3368-75.
54. Thu K, Soria-Bretones I, Mak T, Cescon D. Targeting the cell cycle in breast cancer: towards the next phase. *Cell Cycle* 2018;17(15):1871-85.
55. Zhang L, Wu T, Qiao S. miR-1 and miR-802 regulate mesenchymal-epithelial transition during kidney development by regulating Wnt-4/ β -catenin signaling. *Am J Transl Res* 2019;11(11):7000-8.
56. Goga A, Yagabasan B, Herrmanns K, Godbersen S, Silva PN, Denzler R, et al. miR-802 regulates Paneth cell function and enterocyte differentiation in the mouse small intestine. *Nat Commun* 2021;12(1):3339.
57. Iwanaga R, Komori H, Ishida S, Okamura N, Nakayama K, Nakayama K, et al. Identification of novel E2F1 target

- genes regulated in cell cycle-dependent and independent manners. *Oncogene* 2006;25(12):1786-98.
58. Johnson DG, Ohtani K, Nevins JR. Autoregulatory control of E2F1 expression in response to positive and negative regulators of cell cycle progression. *Genes Dev* 1994;8(13):1514-25.
 59. Lin S-Y, Black AR, Kostic D, Pajovic S, Hoover CN, Azizkhan JC. Cell cycle-regulated association of E2F1 and Sp1 is related to their functional interaction. *Mol Cell Biol* 1996;16(4):1668-75.
 60. Farhan M, Wang H, Gaur U, Little PJ, Xu J, Zheng W. FOXO signaling pathways as therapeutic targets in cancer. *Int J Biol Sci* 2017;13(7):815-27.
 61. Farmer SR. Transcriptional control of adipocyte formation. *Cell Metab* 2006;4(4):263-73.
 62. Sim CK, Kim SY, Brunmeir R, Zhang Q, Li H, Dharmasegaran D, et al. Regulation of white and brown adipocyte differentiation by RhoGAP DLC1. *PLoS One* 2017;12(3):e0174761.
 63. Humphrey SJ, James DE, Mann M. Protein phosphorylation: a major switch mechanism for metabolic regulation. *Trends Endocrinol Metab* 2015;26(12):676-87.
 64. Yang X, Xing H, Liu J, Yang L, Ma H, Ma H. MicroRNA-802 increases hepatic oxidative stress and induces insulin resistance in high-fat fed mice. *Mol Med Rep* 2019;20(2):1230-40.
 65. Qin W, Qi F, Li J, Li P, Zang YS. Prognostic biomarkers on a competitive endogenous RNA network reveals overall survival in triple-negative breast cancer. *Front Oncol* 2021;11:681946.
 66. Rojas K, Baliu-Piqué M, Manzano A, Saiz-Ladera C, García-Barberán V, Cimas FJ, et al. In silico transcriptomic mapping of integrins and immune activation in Basal-like and HER2+ breast cancer. *Cell Oncol* 2021;44(3):569-80.
 67. Wu A, Zhang S, Liu J, Huang Y, Deng W, Shu G, et al. Integrated analysis of prognostic and immune associated integrin family in ovarian Cancer. *Front Genet* 2020;11:705.
 68. Yu M, Hong W, Ruan S, Guan R, Tu L, Huang B, et al. Genome-wide profiling of prognostic alternative splicing pattern in pancreatic cancer. *Front Oncol* 2019;9:773.
 69. Yang L, Wei J, He S. Integrative genomic analyses on interferon- λ s and their roles in cancer prediction. *Int J Mol Med* 2010;25(2):299-304.
 70. Yang L, Zhang R, Guo G, Wang G, Wen Y, Lin Y, et al. Development and validation of a prediction model for lung adenocarcinoma based on RNA-binding protein. *Annals Transl Med* 2021;9(6):474.
 71. Zhang N, Song Y, Xu Y, Liu J, Shen Y, Zhou L, et al. MED13L integrates mediator-regulated epigenetic control into lung cancer radiosensitivity. *Theranostics* 2020;10(20):9378-94.
 72. Tomioka N, Morita K, Kobayashi N, Tada M, Itoh T, Saitoh S, et al. Array comparative genomic hybridization analysis revealed four genomic prognostic biomarkers for primary gastric cancers. *Cancer Genet Cytogenet* 2010;201(1):6-14.
 73. Kitagawa N, Ojima H, Shirakihara T, Shimizu H, Kokubu A, Urushidate T, et al. Downregulation of the micro RNA biogenesis components and its association with poor prognosis in hepatocellular carcinoma. *Cancer Sci* 2013;104(5):543-51.
 74. Wu B, Xi S. Bioinformatics analysis of the transcriptional expression of minichromosome maintenance proteins as potential indicators of survival in patients with cervical cancer. *BMC Cancer* 2021;21(1):928.
 75. Zhang H, Zhang Y, Chen C, Zhu X, Zhang C, Xia Y, et al. A double-negative feedback loop between DEAD-box protein DDX21 and Snail regulates epithelial-mesenchymal transition and metastasis in breast cancer. *Cancer Lett* 2018;437:67-78.
 76. Sang K, Yi T, Huang X, Pan C, Zhou J, Yu L. MiR-370-5p inhibits the progression of breast cancer via targeting LUC7L3. *J Recept Signal Transduct Res* 2021;41(5):442-450.
 77. Hsieh RW, Kuo AH, Scheeren FA, Zarnegar MA, Sikanadar SS, Antony J, et al. CDK19 is a Regulator of Triple-Negative Breast Cancer Growth. *BioRxiv* 2018:317776.
 78. Sheng C, Qiu J, Wang Y, He Z, Wang H, Wang Q, et al. Knockdown of Ran GTPase expression inhibits the proliferation and migration of breast cancer cells. *Mol Med Rep* 2018;18(1):157-68.
 79. Mok K, Tsoi H, Chou K, Khoo U, Man P, editors. KPN1 mediates the nuclear import of BQ323636. 1 and confer tamoxifen resistance in breast cancer cells. *Proceedings of the Annual Meeting of the American Association for Cancer Research*; 2018: American Association for Cancer Research.
 80. Cannizzaro E, Bannister AJ, Han N, Alendar A, Kouzarides T. DDX3X RNA helicase affects breast cancer cell cycle progression by regulating expression of KLF4. *FEBS Lett* 2018;592(13):2308-22.
 81. Wang L, Yu T, Li W, Li M, Zuo Q, Zou Q, et al. The miR-29c-KIAA1199 axis regulates gastric cancer migration by binding with WBP11 and PTP4A3. *Oncogene* 2019;38(17):3134-50.
 82. Pan L, Xu C, Mei J, Chen Y, Wang D. Argonaute 3 (AGO3) promotes malignancy potential of cervical cancer via regulation of Wnt/ β -catenin signaling pathway. *Reproductive Biology*. 2021;21(1):100479.
 83. Zhang P, Cao M, Zhang Y, Xu L, Meng F, Wu X, et al. A novel antisense lncRNA NT5E promotes progression by modulating the expression of SYNCRIP and predicts a poor prognosis in pancreatic cancer. *J Cell Mol Med* 2020;24(18):10898-912.
 84. Liu Z, Yan H, Yang Y, Wei L, Xia S, Xiu Y. Downregulation of CIT can inhibit the growth of human bladder cancer cells. *Biomed Pharmacother* 2020;124:109830.