



Comparison of Antifungal Properties of Gold, Silver, and Selenium Nanoparticles Against Amphotericin B-Resistant *Candida glabrata* Clinical Isolates

Ensieh Lotfali¹, Hossein Toreyhi², Kamyab Makhdoomi Sharabiani², Azam Fattahi^{3*}, Amirali Soheili², Reza Ghasemi², Mahyar Keymaram⁴, Yasaman Rezaee⁵, and Sayna Iranpanah⁵

1. Department of Medical Parasitology and Mycology, Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

2. Student Research Committee, Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

3. Center for Research and Training in Skin Diseases and Leprosy, Tehran University of Medical Sciences, Tehran, Iran

4. Department of Medical Parasitology and Mycology, Faculty of Medicine, Iran University of Medical Sciences, Tehran, Iran

5. Student Research Committee, Faculty of Pharmacy, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Abstract

Background: The present study aimed to investigate the antifungal activity of Nanoparticles (NPs) against amphotericin B-resistant *Candida glabrata* (*C. glabrata*) strains.

Methods: Twelve resistant (*C. glabrata*) strains were isolated from archived clinical isolates. Antifungal activity was conducted according to Clinical and Laboratory Standards Institute's (CLSI) guidelines, document M27-A3/S4. The Scanning Electron Microscope (SEM) was used to observe the morphological changes of strains exposed to each nanoparticle.

Results: Minimum Inhibitory Concentration (MIC) of nanoparticles of all strains was in the concentration range of 0.125 to 0.5 $\mu\text{g}/\text{Ml}$. The synthesized Ag-NPs showed superior antifungal activity against (*C. glabrata*) compared to Se-NPs and Au-NPs. The scanning electron microscope images revealed the difference in the fungal morphology between the untreated and treated fungi with nanoparticles.

Conclusion: The Ag-NPs, followed by Se-NPs synthesized, revealed significant antifungal activity against resistance regardless of their antifungal-resistant mechanisms.

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Introduction

Candida glabrata (*C. glabrata*) is one of the most common opportunistic human fungal pathogens, which is responsible for 90-100% of mucosal infections and invasive candidiasis with the high mortality rate in immunocompromised and critically ill individuals¹. Over the past decade, the improper and misuse of existing antifungal drugs have led to the development of drug-resistant clinical isolates of *Candida* species (spp.). The classes of antifungal drugs active against *C. glabrata* include pyrimidine analogs (Flucytosine), azoles, echinocandins, and polyenes². Resistance to amphotericin B is rare, however, a change in the amount of ergosterol or production of different types of sterol that construct fungal cell membranes is the probable mechanism for this resistance without causing a major challenge in normal cell functionality².

The synthesis of Nanoparticles (NPs) has been studied for some time, and various attempts have been made to apply them to advance the field of nanomedicine. Nanoparticles have diverse features depending on the metallic element, size, and shape of the particles. Specifically, silver nanoparticles are well-known for their biocidal properties, including antibacterial, antifungal, antiviral, and anticancer activities³. The resistance of *C. glabrata* to routine antifungal therapy poses a major challenge to the management of infection, hence the use of other agents with antifungal property helps to improve the management of infection. Here, it is hypothesized that silver (Ag), selenium (Se) and gold (Au) nanoparticles could change cell wall structure in the resistant strains through forming pores, possibly improving the sensitivity of resistant isolates. The

present study aimed to investigate the antifungal activity of Ag-NPs, Se-NPs, and Au-NPs against amphotericin B-resistant *C. glabrata* strains.

Materials and Methods

Fungal strain

Twelve resistant *C. glabrata* strains (Table 1) were isolated from the vaginal tract of patients in an epidemiological study of antifungal resistance. This study was performed at Department of Medical Parasitology and Mycology of Shahid Beheshti University of Medical Sciences. *C. glabrata* (ATCC 90030) was used as a control strain. Antifungal activity of three NPs, including Ag-NPs, Se-NPs, and Au-NPs were investigated according to Clinical and Laboratory Standards Institute's (CLSI) guidelines mentioned in the document M27-A3/S4⁴.

Synthesis and characterization of NPs

The nanoparticles (Sigma, USA) were prepared according to the previous methods⁵. For the characterization of size and charge of NPs, the Z-average size and zeta potential were estimated using the Malvern Zetasizer Nano ZS-90 instrument (Malvern, England). A Scanning Electron Microscope (SEM) (AIS2100, Seron Technology, South Korea) was used to observe the morphological changes of *C. glabrata* strains exposed to the NPs (Ag, Se, Au)⁶.

Statistical analysis

All the data of the antifungal activity were expressed as mean±SD. The significant differences were defined at $p < 0.05$.

Results

Characteristics of NPs

The zeta-average sizes of Au, Se, and Ag-NPs were ranged from 10-15, 105-209, and 30 nm, respectively. The zeta potentials of Au, Se, and Ag NPs were -22.4, -49.93, and -23.5 Mv, respectively.

Minimum inhibitory concentration of the NPs

After 24 hr of incubation, the Minimum Inhibitory Concentration (MIC) of nanoparticles of all *C. glabrata* strains was in the concentration range of 0.125 to 0.5 µg/ml (Table 1). No inhibition was observed in the low concentration range of ≤ 0.125 µg/ml; however, the growth was significantly inhibited at 0.125 µg/ml or more compared to the control. The synthesized Ag-NPs had superior antifungal activity against *C. glabrata* compared to Se-NPs and Au-NPs.

Scanning electron microscope image

In *C. glabrata*, the untreated *C. glabrata* (Figure 1A) was only spherical or oval-shaped blastoconidia, which were considerably smaller than another *Candida* spp. Once *C. glabrata* was treated with Ag-NPs (Figure 1B), abnormal morphology, some pores, and distorted membranes appeared in the yeasts (white arrows on the image). As shown in figure 1C, Se-NPs were attached and agglomerated to the surface of nearby yeasts (white arrows on the image). After treatment with NPs at the MIC dose, the cell membrane lost its smoothness and resulted in unusual surface bulges, inducing impairment of cell membrane integrity. At 2× MIC dose concentration of Ag-NPs and SeNPs (Figures 1B and 1C), the fungal morphology underwent a dramatic change from cylindrical to distorted cell struc-

Table 1. Comparison MIC of 12 AMB-resistant *C. glabrata* strains exposure to Ag-NPs, Se-NPs, Au-NPs

Isolates	Antifungals			Nanoparticles		
	AMB	FCZ	ITZ	AgNPs	SeNPs	AuNPs
	MIC range (µg/ml)					
	R: ≥1 I: ND S: ≤1	R: ≥8 I: 4 S: ≤2	R: ≥1 I: ND S: ≤1	-	-	-
<i>C. glabrata</i> R1	2	8	1	0.5	1	2
<i>C. glabrata</i> R2	2	4	1	0.25	0.5	2
<i>C. glabrata</i> R3	2	4	0.5	0.25	0.25	2
<i>C. glabrata</i> R4	4	8	0.5	0.5	0.5	4
<i>C. glabrata</i> R5	2	8	1	0.25	0.5	2
<i>C. glabrata</i> R6	2	8	1	0.25	0.5	2
<i>C. glabrata</i> R7	2	4	0.25	0.5	0.5	2
<i>C. glabrata</i> R8	2	2	0.5	0.5	0.5	2
<i>C. glabrata</i> R9	4	4	0.5	0.125	0.5	4
<i>C. glabrata</i> R10	2	8	1	0.125	0.5	2
<i>C. glabrata</i> R11	2	4	0.25	0.25	0.5	2
<i>C. glabrata</i> R12	2	2	0.5	0.25	0.5	2
GM±SD	2.33±0.77	5.33±2.46	0.66±0.3	0.31±0.14	0.52±0.16	2.33±0.77
ATCC (90030)	0.5	2	0.25	0.5	0.5	0.5

Fluconazole (FCZ); Amphotericin B (AMB); Itraconazole (ITZ). S: Susceptible, I: Intermediate, R: Resistant, MIC: Minimum inhibitory concentration. No. of resistant *C. glabrata* strains (R1-R12). Geometric range; GM, Standard Deviation; SD.

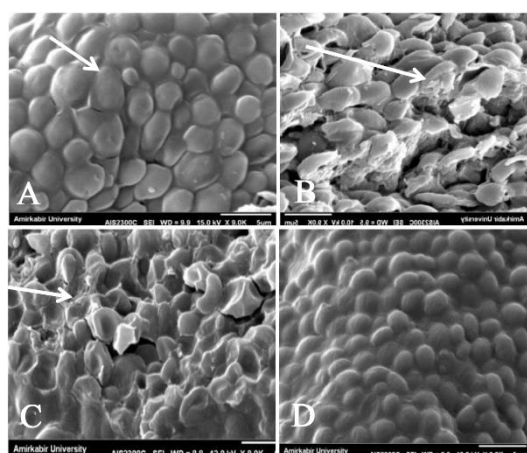


Figure 1. SEM images of *C. glabrata* treated with silver nanoparticles; 9000×magnification

- A) Untreated *C. glabrata*; the white arrow indicates the smooth cell membrane of normal yeast.
 B) *C. glabrata* treated with 0.5 µg/ml Ag-NPs.
 C) *C. glabrata* treated with 0.5 µg/ml Se-NPs.
 D) *C. glabrata* treated with 0.5 µg/ml Au-NPs.

ture with the breakdown of the cell membranes. However, at 2×MIC level concentration of Au-NPs (Figure 1D), the fungal morphology did not change much.

Discussion

Regarding the present observation, NPs have excellent antifungal effects, because of their low MIC ranges and, consequently, have more potency against amphotericin B-resistant *C. glabrata* strains.

The results of antifungal activity against resistant *C. glabrata* strains after exposure to Ag-NPs with inhibitory action at a 0.125-0.5 µg/ml concentration were better as compared to Se-NPs and Au-NPs. The superior activity of Ag-NPs to inhibit the growth of various pathogenic fungi, such as *Trichophyton* and *Microsporum* spp, was reported⁷. Ag-NPs followed by Se-NPs have not contributed to fungal resistance complicating antifungal therapy of candidiasis. Present results show that synthesized Ag-NPs have highly antifungal activity against resistant *C. glabrata* strains, which might be through the destruction of membrane integrity, resulting in the formation of pores and cell death (Figures 1B and 2B). It has been suggested that upon the treatment, the replication ability of DNA, expression of ribosomal subunit protein and the enzymes essential for ATP production are lost, leading to the inactivation of microorganisms⁸.

The mechanism of inhibitory effect of NPs on resistant isolates of *C. glabrata* creates a scenario in which Ag-NPs/Se-NPs like different routine antifungals have a different fungistatic or fungicidal mechanism, including interference with cell wall synthesis and inhibition of ergosterol synthesis. It means that these agents do exert their fungistatic/fungicidal effects in a broad range of targets in the organisms. Based on

the MIC results, it is safe to say that nanoparticles have an intensive antifungal property, making them a suitable alternative with regard to drug-resistant isolates.

Some studies have reported that the positive charge on the Ag⁺ ion is crucial for its antimicrobial activity through the electrostatic attractions between the negatively charged cell membrane of microorganisms and the positively charged nanoparticles⁹.

In the present study, no antifungal activity of Au-NPs was observed in all tested isolates. Au-NPs exhibited antifungal activity against *C. glabrata*, *Candida albicans* (*C. albicans*), *Candida krusei* (*C. krusei*), *Candida parapsilosis* (*C. parapsilosis*), and *Cryptococcus neoformans*, while *Candida tropicalis* *C. tropicalis* and dermatophyte spp. revealed resistance to Au-NPs (MIC for dermatophyte spp.: 10-30 µg/ml)¹⁰.

To sum up, the Ag-NPs, followed by Se-NPs synthesized, revealed significant antifungal activity against amphotericin B-resistant *C. glabrata* strains, disregarding their antifungal-resistant mechanisms. The activity analysis of nanoparticles indicated that the nanoparticles were capable of changing membrane and cell wall structure in the resistant strains, possibly through pore formation on the membrane and cell wall.

Conclusion

This research revealed the fungicidal capacities of Ag-NPs followed by Se-NPs against amphotericin B-resistant *C. glabrata* strains. This information can be useful in encapsulating antifungal drugs in combination with the mentioned NPs to decrease the side effects and enhance the effectiveness of the drugs.

Conflict of Interest

None declared.

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