

## Conjugation of Monoclonal Antibodies to Super Paramagnetic Iron Oxide Nanoparticles for Detection of her2/neu Antigen on Breast Cancer Cell Lines

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### Abstract

Conjugation of monoclonal antibodies to super paramagnetic nanoparticles is an effective method for cancer diagnosis and treatment. In this study the humanized anti her2/neu monoclonal antibody- Herceptin- was conjugated to super paramagnetic iron oxide (SPIO) nanoparticles using EDC method. The concentration of the conjugated antibodies was measured by Bradford assay. The antibody-nanoparticle conjugates were incubated with SKBR-3 and T47D human breast carcinoma cell lines and the presence of the conjugates on cell surface was confirmed by Prussian blue iron staining method. Conjugation of Herceptin to SPIO resulted in a precipitate-free conjugate containing 20µg antibody/mg SPIO. Prussian blue iron-staining of cells showed successful binding of the conjugates to the cell surfaces. Conjugation of monoclonal antibodies to SPIO may be a useful method for detection of tumor cells, especially by MRI techniques.

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### Introduction

Molecular probes for biomolecular recognition are of great importance in the fields of chemistry, biology, medical sciences and in biotechnology as well. These probes have been used in studies of biological functions and in ultrasensitive detection of biological factors responsible for many diseases<sup>(1)</sup>. On the other front, the developments of nontoxic and biocompatible magnetic particles have been disclosed for biological applications since mid-1980s<sup>(2)</sup>.

Recently, magnetic particles have attracted growing interest as high performance biomaterial which is used for transport and separation of cells or cell parts<sup>(2, 3)</sup>, MRI<sup>(4)</sup>, hyperthermia<sup>(5)</sup> and drug delivery<sup>(6)</sup>. Biological samples such as blood, serum, cell suspensions and cell lysates are allowed to be exposed to specific ligand-coupled particles, and the captured molecules or cells are then rapidly separated using magnetic fields<sup>(3,7,8)</sup>.

Magnetic particles conjugated with anti tumor monoclonal antibodies provide a new approach to identify tumor cells.

Antibodies labeled with magnetic nanoparticles give magnetic signals on exposure to a magnetic field. Iron oxide particles are usually coated with different organic shells including dextran, albumin or polyethylene glycol. Coated nanoparticles can be manufactured with a variety of functional groups (such as amino, aldehyde, hydroxyl, sulfate and carboxyl groups) on their surfaces. Considering these properties, we used super paramagnetic iron oxide (nanomag-D-SPIO 20nm) with COOH group on the surface, for conjugation to a humanized anti her2/neu monoclonal antibody (Herceptin) as a cancer targeting antibody.

### Materials and Methods

Nanomag-D-SPIO 20 nm nanoparticles (surface COOH) and MACS separator with MS columns were purchased from Micromod (Miltenyi Biotech GmbH, Germany). The breast carcinoma cell lines SKBR-3 and T47D were obtained from Pasteur Institute of Iran. Other reagents and chemicals were obtained from Merck and Sigma.

#### Conjugation of anti her2 antibody (Herceptin) with nanoparticles by EDC method

N-ethyl-N-(3-dimethyl aminopropyl) carbodiimide hydrochloride (EDC, 26mM) and 10 mM N-hydroxy succinimide (NHS) were dissolved in 0.1 M 2-(N-morpholino) ethane-

sulfonic acid (MES) buffer (pH=8.3). The mixture was added to 1 ml of 5 mg/ml nanomag-D-SPIO 20 nm nanoparticles, and shaken at room temperature for 2 hours.

The particles were washed twice with phosphate buffered saline (PBS) pH=7.4 and then 0.5-1 mg/ml of Herceptin was added to the activated particles. The mixture was shaken for 3 hours and the reaction was quenched by the addition of glycine for 30 minutes<sup>(9, 10)</sup>.

The unconjugated antibodies were separated from conjugated antibodies by MACS column. The amount of immobilized antibody was estimated based on the Bradford method.

#### Spectrophotometric measurement of Iron

Iron concentration of conjugated samples was obtained by potassium thiocyanate method<sup>(12,13)</sup>. In brief, samples were diluted with 300  $\mu$ l 6N HCl containing %1 H<sub>2</sub>O<sub>2</sub>; under this condition, the iron in the samples is dissolved and oxidized to ferric state. The samples were then added to a 5% solution of potassium thiocyanate where the Fe III formed a red complex with the thiocyanate which could be measured by absorbance at 480 nm.

#### Cell culture

The her2/neu expressing cell lines SKBR3 and T47D were grown in RPMI 1640 medium with 10% (v/v) fetal calf serum and %1 penicillin/streptomycin. Cells were incubated at 37°C containing 5% Co<sub>2</sub>.

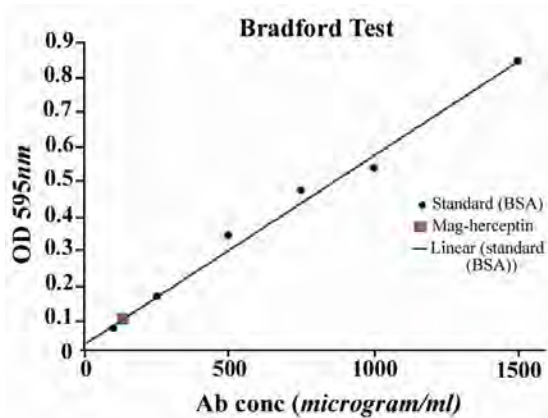


Figure 1. Antibody concentration measurement by Bradford assay (Ab conc. 100  $\mu$ g/ml)

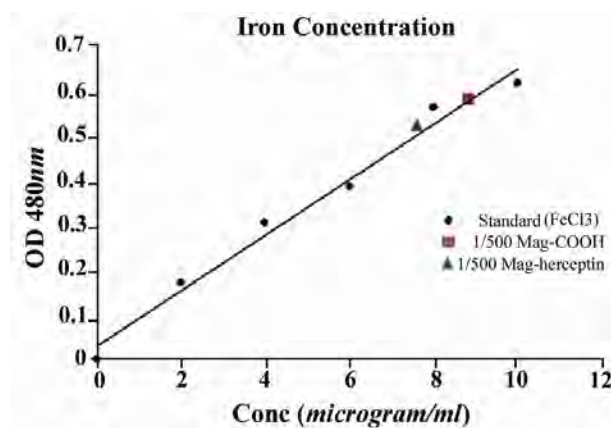


Figure 2. Magnetic nanoparticles concentration measurement by Potassium Thiocyanate method (particles conc. 4-4.5mg/ml)

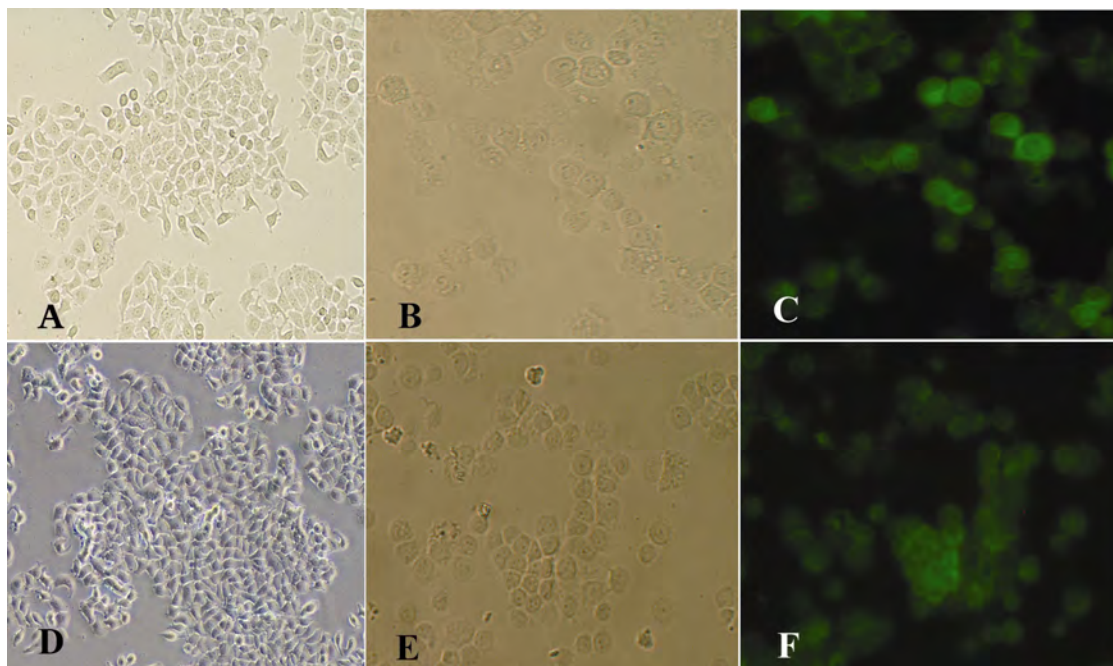


Figure 3. Light microscopy and Immunofluorescence of SKBR3 and T47D cell lines  
 A: SKBR3 cells in culture, B: suspension of SKBR3 cells, C: SKBR3 cells after incubation with Herceptin-FITC, D: T47D cells in culture, E: suspension of T47D cells, F: T47D cells after incubation with Herceptin- FITC.

### Immunofluorescence staining

To verify the expression of her2 proteins on the cells, the SKBR3 and T47D cells were incubated with anti her2/neu (Herceptin) at  $10 \mu\text{g/ml}$  concentration for 1 hour at  $37^\circ\text{C}$ . After being washed in PBS, FITC-labeled anti human IgG (diluted 1/20, Avicenna Research Institute, Tehran, Iran) was added and incubated for 1 hour at room temperature. Cells were then observed directly on a fluorescence microscope (Olympus, Japan).

### In vitro cell labeling

SKBR3 and T47D cells were counted and adjusted to a suspension of  $4 \times 10^5 \text{ cells/ml}$ ;  $100 \mu\text{l}$  of each cell suspension were cytopinned on microscope slides (Shandon cyto-spin 4, Thermo, Germany). The cells were incubated with  $100 \mu\text{l}$  magnetic nanoparticles (with or without antibody;  $5 \mu\text{g Ab}$  and  $0.2 \text{ mg iron}$ ) for 1 hour at  $37^\circ\text{C}$ . Then cells were washed extensively with PBS to remove unbound particles.

The nanoparticles that bounded on the cell surface were detected by iron staining with Prussian blue staining method<sup>(14, 15)</sup>.

### Results

We used  $20 \text{ nm}$  nanoparticles (a magnetic core covered with dextran) with carboxyl group for conjugation to Herceptin as a cancer targeting antibody. The final products of conjugation were suspensions without precipitate and the amount of immobilized antibody was  $20\text{-}36 \mu\text{g Ab/mg}$  nanoparticles (Figure 1).

The amount of iron conjugated was determined by potassium thiocyanate method using  $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$  as a reference (Figure 2).

The SKBR3 and T47D cell lines showed high levels of her2/neu expression, after incubation with Herceptin-FITC (Figure 3), but not with the isotype control antibody (data not shown).

These results confirmed the presence of antigen on the cell surface. Specific binding of the conjugates to the cells can be determined by using electron microscopy, MRI imaging and iron staining<sup>(13,18,19)</sup>.

In this study after incubation of the cells with conjugated nanoparticles, the blue stain induced by iron staining showed the accumulation of conjugated nanoparticles on the cell surface (Figure 4).



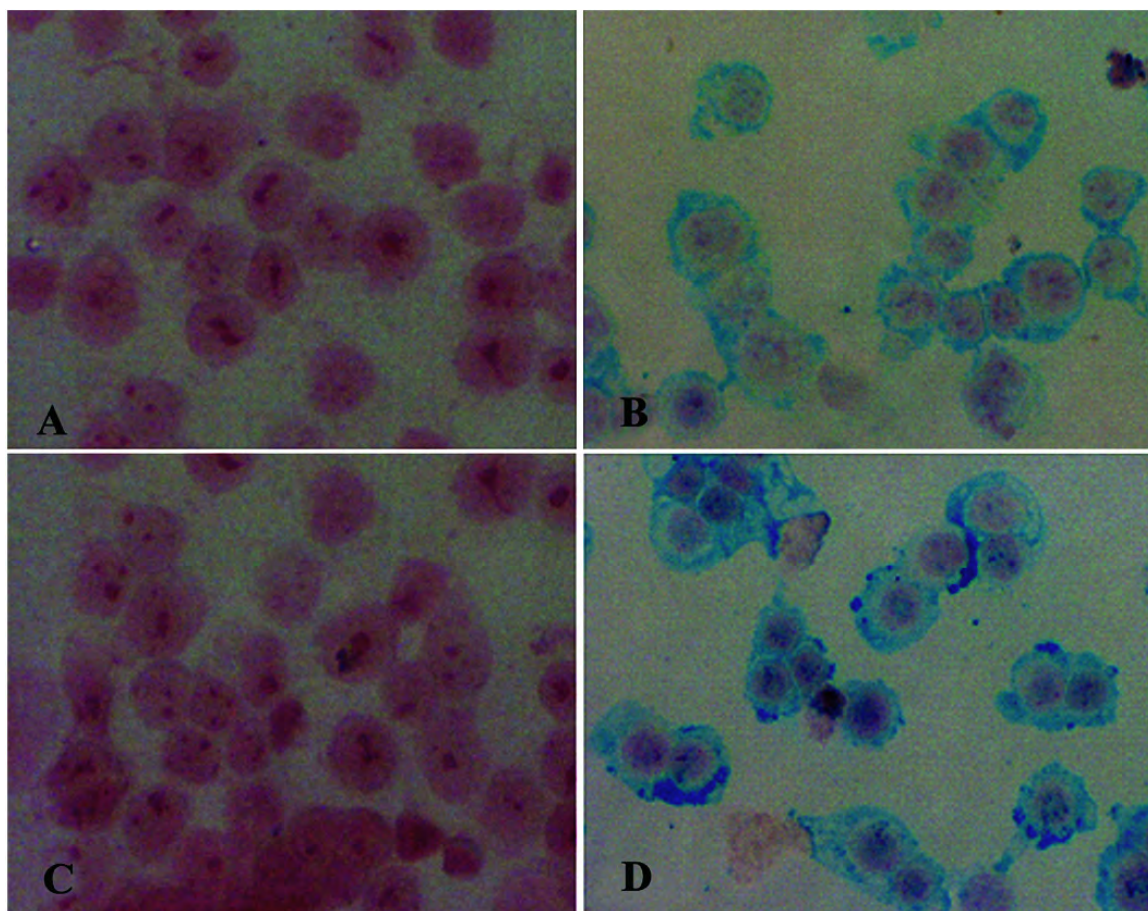


Figure 4. Iron staining of cells after incubation with conjugated nanoparticles  
A: SKBR3 cells, B: KBR3 with iron staining, C: T47D cells, D: with iron staining.

### Discussion

Polymer-coated magnetic particles with particle sizes 5-500 nm have been employed in medicine or biotechnology for many years<sup>(16)</sup>. In this study the 20 nm nanoparticles were coupled via their surface carboxyl group to the amino groups on the Herceptin antibody using the EDC method<sup>(10, 17)</sup>.

After conjugation, the amount of immobilized antibody was approximately 20 µg/mg magnetite. However by increasing concentration of antibody during the process, the efficiency of the conjugation did not improve. In other studies the efficiency of conjugation has been reported as 5-20 µg Ab/mg particles<sup>(9,18)</sup>. Conjugated nanoparticles bound specifically to the her2/neu antigen.

Iron staining, confirmed the presence of nanoparticles on the cell surface. In the present study we showed specific binding of Herceptin-nanomagnetic particle conjugates

to her2/neu over expressing cells, suggesting a future application of Herceptin-magnetite for MR imaging of breast cancer.

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