Inclusion Body Expression and Refolding of Recombinant Bone Morphogenetic Protein-2

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Abstract

Background: Bone Morphogenetic Protein-2 (BMP-2) is a cysteine rich growth factor expressed in homodimeric form and has a pivotal role in osteochondral development and fracture healing. Recent studies have benefited more from recombinant BMP-2 in osteochondral tissue engineering. Cost-effective and easy production at large scale makes Escherichia coli (E. coli) the first choice for recombinant protein expression programs. However, inclusion body aggregation and refolding process limits production and purification of recombinant BMP-2 in bacterial systems.

Methods: BMP-2 encoded gene was optimized for expression in bacterial expression system and synthesized with proper restriction sites. The optimized sequence was then cloned in a pET28a expression vector and expressed in Origami™ E. coli strain. The aggregated and monomeric BMP-2 was refolded and purified comparing two oxidoreductase systems and refolding methods as well as different purification techniques. The biological activity of recombinant protein was investigated by increasing alkaline phosphatase activity (ALK) of ATDC-5 cell line.

Results: No difference was observed between oxidoreductase systems in improving the efficiency of protein refolding. However, comparisons between two refolding methods showed that pooling monomeric BMP-2 that was refolded under mild condition with equal volume of it refolded under severe oxidoreductase condition resulted in production of more active dimeric protein.

Conclusion: A new method for production of biologically active dimeric form of BMP-2 in E. coli expression system was established in this study.

Keywords: Bone morphogenetic protein -2, Cloning, Escherichia coli, Inclusion bodies, Protein refolding

Introduction

Bone morphogenetic proteins belong to transforming growth factor β superfamily which include TGF β, Activin, Nodals, Inhibins and Growth Differentiation Factors (GDFs) family. The members of TGF β superfamily have a vital role in embryonic development 1 and sending messages through smad signaling pathway 2. Over 20 BPM members have been identified to date 3, among them BMP-2, BMP-3, BMP-4 and BMP-7 have been studied more than any others. BMP-2 is a cysteine rich protein that plays an important role in many stages of chondrocytes differentiation and maturation rather than osteogenesis in vivo. The mature BMP-2 is a disulfide -connected homo-dimer, composed of 114 amino acids derived from the C-terminal sequence of a 396 amino acid precursor polypeptide. The intra and inter chain disulfide bonds are essential for active structure formation and ligand binding to cell surface receptors.

Cysteine rich heterologous protein production in prokaryotic systems leads to inclusion body formation and ligand binding to cell surface receptors.

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Materials and Methods

Gene synthesis and cloning

The protein coding sequence of 114 N-terminal amino acid chain for bone morphogenetic protein2 (UniProtKB-P12643) was retrieved from genbank [NM_001200.2] and optimized for expression in Origami 2 (DE3) pLys strain of E. coli (Novagen). Putting BamHI and HindIII restriction sites at amino and carboxyl ends, the construct was synthesized (GENEray Biotechnology) and subcloned in pET28a (Novagen Inc., Madison, Wis). The recombinant plasmid was transformed to competent cells and colony PCR and construct digestion were employed for cloning validation.

Protein expression and extraction

For recombinant BMP-2 expression, the transformants were grown in LB culture media containing 15 µg/ml kanamycin until OD₆₀₀=1 followed by induction with 0.2 mM IPTG for 6 hr at both 25 and 37 °C. The harvested cells were lysed by cell lytic buffer (20 mM PBS, 500 mM NaCl, 1 mM EDTA, 100 µg/ml Lysozyme, Benzonase 80 µ/ml, 0.1% Triton X100) and centrifuged at 14000 g for 30 min. The supernatant contained total soluble proteins which were applied to a nickel-charged affinity resin column (Ni-NTA Agarose, Qiagen 30210) and his-tag affinity was purified with imidazole buffers (300 mM NaCl, 50 mM phosphate buffer pH=8.0 and 50 to 300 mM imidazole) for his-tag affinity purification. The remaining pellet containing inclusion bodies was resuspended in wash buffer (20 mM PBS, 1 mM EDTA, 2% Triton X100) and centrifuged at 26000 g for 15 min and then solubilized in inclusion body dissolving buffer (6M GuHCl, 0.1 M Tris-HCl pH=8.5, 1 mM EDTA, 50 mM DTT) and incubated for 2 hr at room temperature. The protein solution was then centrifuged at 14000 g for 15 min and DTT was removed by dialysis against 5 M GuHCl, 0.1 M Tris Hcl, pH=3.5 overnight. Finally, a Bradford assay was run for protein content estimation.

Protein refolding and purification

BMP-2 refolding was carried out by dilution of protein in refolding buffer (100 mM Tris/HCl pH=8.3, 5 mM EDTA, 250 mM NaCl, 0.5 M L-arginine) to decrease protein concentration and prevent aggregation. Two oxidoreductase systems (reduced glutathione/oxidized glutathione and Cysteine/Cystine) and different refolding methods were used to analyze disulfide bond formation as well as protein dimerization (Table 1). BMP-2 refolding was started by gradual addition of denatured protein to pre-cooled refolding buffer until the concentration fell below 0.2 mg/ml while stirring at 4 °C for 24 hr. The protein concentration increased to 1 mg/ml by cross flow filtration (Amicon ultra centrifugal filter) and redox system components were removed by dialysis against urea 6 M, 0.1 M Tris/HCl and 5 mM EDTA, pH=6. Dimeric protein was eluted by heparin
column (Heparin sepharose™ 6 Fast Flow, GE Healthcare, 17-0998-01) at 0.3 and 0.7 M NaCl and purified protein was dialyzed against 20 mM ammonium acetate buffer, pH=4.8 and lyophilized.

**SDS-PAGE and western blotting**

Recombinant BMP-2 expression and refolding pattern were analyzed by non-reducing 12% SDS-PAGE. Separated proteins were electrophoretically transferred to nitrocellulose membranes by semi-dry blotting using a Dunn carbonate transfer buffer (10 mM NaCHO₃, 3 mM Na₂CO₃, 20% methanol). The membranes were blocked for 2 hr using western blocker solution (Sigma, W0138) and incubated overnight at 4°C, with anti-Histag (1:30000, Penta-His Antibody mouse monoclonal Qiagen, 1008288) as a primary antibody. The blots were washed three times with washing buffer (PBS-Tween-20, 0.05%) and incubated with the peroxidase-conjugated secondary rabbit anti mouse antibody (sigma A9044) for one hour at room temperature. The blots were visualized using ECL detection reagents (Sigma, CPS-1-120) and the films were scanned with a densitometer (GS-800, Bio-Rad).

**Biological activity test of recombinant BMP-2**

Biological activity of BMP-2 was measured by evaluation of its ability to increase the alkaline phosphatase activity (ALK) of a chondrogenic mouse embryonic carcinoma cell line, ATDC-5. The ATDC5 cell line was purchased from the RIKEN Cell Bank (RCB-0565) and a positive control was purchased from R&D company (cat no: 355-BM). ALK activity of BMP-2 induced cell line was tested using Alkaline Phosphatase Yellow Liquid Substrate System for ELISA, containing p-nitrophenyl phosphate (pNPP) (Sigma P7998).

**Cell culture**

ATDC-5 cells were cultured in Dulbecco’s modified eagle’s medium DMEM-F12 (Invitrogen, 105 21331-020) supplemented with 5% Fetal Bovine Serum (FBS; Gibco Inc. Canada) and 50 mg/ml Non-Essential Amino Acid (NEAA) (Gibco Inc. Canada) without any antibiotic.

**Assay medium**

A Serum Free Medium (SFM) was prepared for assay with RPMI 1640 (Gibco Lot no: 1668798) as a basal medium supplemented with NAEE and 0.1% bovine serum albumin.

**Assay for alkaline phosphatase activity**

Briefly, SFM contained serial dilutions of recombinant BMP2 and commercial R&D BMP2 (0-500 ng/ml) was provided in a 96 well flat-bottom plate and ATDC-5 cells were plated and incubated for 5 days at 37°C in a humidified, 5% CO₂ atmosphere. After 5 days of culturing, cells were washed with PBS and enzyme activity was assayed by pNPP -a substrate in alkaline phosphatase assay buffer- and then plate was incubated at 37°C for 20 min. Next, the absorbance was measured at 405 nm with microplate reader.

**Results**

**Analysis of cloning process**

Synthetic coding sequence of BMP-2 which was restriction enzyme digested of pGH plasmid was sub-cloned into pET28 plasmid. After transformation of competent cells with recombinant plasmid, several colonies were selected on antibiotic plate for further analyses. Colony PCR using T7 primers (Figure 1) as well as restriction enzyme digestion of recombinant plasmid (Figure 2) amplified and released expected product sizes.

**Identification of recombinant BMP-2 protein**

SDS-PAGE banding pattern analysis of proteins extracted from 37°C cultured E. coli showed that the recombinant BMP-2 was exclusively expressed as an insoluble fraction and accumulated in cytoplasm as inclusion bodies. Inclusion body extracted proteins exhibited a sharp band between 17 to 20 kDa, corre-
sponding to monomeric chain of BMP-2 (Figure 3). Similar result was also obtained following protein expression at 25°C (data not shown).

Recombinant BMP-2 refolding results were assessed via non-reducing SDS-PAGE (Figure 3). Protein dimer formation in the refolding process accounts for approximately 40-50% at best.

Western blot analysis (Figure 4) using anti-HisTag which strongly reacted with N-terminal histidine tag of recombinant BMP-2 protein, confirmed that migrated bands between 17 to 20 kDa and near 25 kDa are related to the monomeric and dimeric forms of the protein, respectively.

**Evaluation of recombinant protein refolding process**

In order to analyze the efficiency of disulfide bond formation in BMP-2 protein, two oxidoreductase systems in refolding buffer were used for refolding process (Table 1). Figure 5 illustrates electrophoretic mobility pattern of heparin affinity purified BMP-2 which was refolded by different oxidoreductase systems. A ratio of 10/1 of reduced/oxidized components was used for method one refolding process of the recombinant protein (Table 1).

The electrophoretic mobility of recombinant BMP-2 which was refolded by method two protocol (Table 1) is represented in figure 4. In this method, the unfolded protein was diluted and incubated up to 48 hr in refolding buffer containing 0.1/0.1 mM GSH/GSSG that catalyzes intra chain disulfide bond formation. Next, the solution was combined and incubated with equal volume/concentration of 5 mM/0.5 mM pretreated GSH/GSSG (product of method one) for two hours and then dialyzed. Figure 3 depicts electrophoretic pattern of refolded dimeric BMP-2 compared with that of monomeric fraction which are both eluted by 0.3 and 0.7 M NaCl, respectively.

**Analysis of protein purification methods**

Refolded BMP-2 was purified using two methods, histidine tag affinity purification and heparin affinity chromatography. His-tag affinity purification included two essential elements.

First, N-terminal histidine tag of recombinant protein was provided by expression vector and expressed as a fusion protein with BMP-2. Second, there was nickel-charged affinity resin column for protein purification.

Different concentrations of imidazole were used in elution buffer for affinity purification of BMP-2. However, no significant differences were observed in monomer/dimer separation (data not shown). Heparin affinity purification of refolded protein is presented in figure 3. Protein purification using elution buffers containing 0.3 and 0.7 M NaCl showed the best results in monomer/dimer separation.

**Biological activity of recombinant BMP-2**

The biological activity assay indicated that the refolded recombinant BMP-2 increased the ALK activity of ATDC-5 cell line as much as that of commercially available eukaryotic protein (Figure 6). Increasing the concentration of refolded BMP-2 applied onto ATDC-5 cell line resulted in induction of ALK activity in a dose-dependent manner. This is a practical assay for

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**Figure 3.** A) Nickel charged affinity resin purification of refolded BMP-2, using different concentrations of imidazole in elution buffer. Lanes 1 to 5; 30, 50, 80, 150 and 300 mM imidazole, respectively. B) Refolded and purified BMP-2 analysis using non-reduced 12% SDS-PAGE. Lane 1: refolded but unpurified protein. Lane 2 and 3: monomeric and dimeric forms of BMP-2 purified by heparin affinity method.

**Figure 4.** Western blot analysis of purified BMP-2, separated by 12% SDS-PAGE and transferred to nitrocellulose membrane by semi-dry electroblotting. Lane 1: unpurified protein, lane 2: monomeric protein and lane 3: dimeric form of BMP-2.

**Figure 5.** One-stage refolding of BMP-2 using refolding buffer containing cysteine/cystine (A & B) or GSH/GSSG (C & D) oxidoreductase systems. Lane A and C: 2/0.2 mM, lane B and D: 5/0.5 mM reduced/oxidized components’ ratio.
asurement of BMP-2 activity that can be performed with high-accuracy.

Discussion

Bacterial expression of BMP-2 resulted in production of inclusion body and therefore, protein aggregation in cytoplasm. The aggregated protein can be solubilized and extracted from inclusion body under high concentration of urea and GuHCl. Native structure of protein can be restored by decreasing urea/GuHCl concentrations and replacing them with compounds which prevent protein aggregation and promote proper folding. In the next step, these compounds should be removed gradually and protein is solubilized in appropriate buffers, capable of maintaining its active structure. Cysteine-rich proteins require a proper oxidoreductase system for disulfide bond formation or rearrangement of incorrectly formed bonds.

In this study, synthetic coding sequence of BMP-2 protein was restriction digested and subcloned in pET28a plasmid and finally transformed in Origami™ strain of E. coli. Verification of the insert and its orientation was approved by colony PCR amplification and restriction digestion analysis.

SDS-PAGE analysis of recombinant BMP-2 expression confirmed a high level of protein, aggregated as inclusion bodies and entirely not present in soluble solution. Measurement of BMP-2 activity was approved by colony PCR amplification and measurement of BMP-2 activity that can be performed with high-accuracy.

In an attempt to decrease multimeric structure and at the same time increase purified dimeric protein as well as biological activity, a different refolding procedure was designed in addition to heparin affinity purification. First, the unfolded BMP-2 was diluted gradually in protein refolding buffer containing 0.1 mM /0.1 mM of GSH/GSSG for up to 48 hr and then dialyzed. Second, the dialyzed protein was pooled with an equal volume of BMP-2 pretreated with 5 mM /0.5 mM of GSH/GSSG and incubated for two hours. Our hypothesis was that the refolding of BMP-2 under this mild condition (0.1 mM /0.1 mM GSH/GSSG) results in correct formation of intrachain disulfide bonds and therefore monomeric renaturation. Pooling with 0.5 mM /5 mM of GSH/GSSG pretreated BMP-2—which is oxidized rather than being in reduced state—resulted in improvement of protein dimerization and thus activity. Finally, the monomeric and misfolded multimeric structures of BMP-2 were successfully eluted with high purity using a heparin affinity purification column (Figure 3).

Conclusion

In view of well-established methods available for protein refolding, our study presented a reproducible, practical and cost-effective method for BMP-2 growth factor production in E. coli bacterial system which is comparable with eukaryotic systems in terms of biological activity and stability.

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References

3. Carreira AC, Zambuzzi WF, Rossi MC, Astorino Filho R, Sogayar MC, Granjeiro JM. Bone morphogenetic pro-
teins: promising molecules for bone healing, bioengineering, and regenerative medicine. Vitam Horm 2015;99:293-322.


