Apoptosis of Adipose-Derived Stem Cells Induced by Liposomal Soybean Phosphatidylcholine Extract

Reza Y. Purwoko 1*, Iis Rosliana 1, Siti Sobariah 1, Nabila Hermana 1, Silvani Permatasari 1, Dewi Wulandari 2, Puji Sari 3, Ernie H. Purwaningsih 4, Chaidir 5, Hans-Joachim Freisleben 6, Jeanne A. Pawitan 7, and Kusmarinah Bramono 8

1. érpour Medical-Spa Skin and Mesotherapy Centre, Jakarta, Indonesia
2. Department of Pharmacy, Faculty of Medicine, Universitas Indonesia, West Java, Indonesia
3. Department of Biology, Faculty of Medicine, Universitas Indonesia, West Java, Indonesia
4. Department of Pharmacy, Faculty of Medicine, Universitas Indonesia, West Java, Indonesia
5. Agro Industrial Technology Development Biomedical Laboratory, Serpong, South Tangerang, Indonesia
6. German Indonesian Medical Association, Jakarta, Indonesia
7. Department of Histology, Faculty of Medicine, Universitas Indonesia, West Java, Indonesia
8. Department of Dermatovenerology, Faculty of Medicine, University of Indonesia, West Java, Indonesia

Abstract

Background: Recently, Phosphatidylcholine (PC) has been used as an off-label treatment for lipolysis injection, which is associated with inflammatory reaction due to sodium deoxycholate, an emulsifier, so that inflammation as side effect occurs in those patients. Liposome formulation from soybean lipid was thought to be a better and safer alternative. This study aimed to analyze the mechanism of Liposomal Soybean Phosphatidylcholine (LSPC) extract from Indonesian soybeans (containing 26% PC) to induce Adipose-derived Stem Cells (ASCs) death in vitro.

Methods: Liposomes were prepared using thin film hydration method followed by a stepwise extrusion process to produce a small amount of 41.0-71.3 nm. Liposomal soybean phosphatidylcholine extract (LSPCE), liposomal purified PC (LPCC), and solution of PC+SD were used for comparison. Annexin V-fluorescein Isothiocyanate/Propidium Iodide (FITC/PI) double staining by flow cytometry and also measurement of caspase-3 activity using ELISA were used to quantify the rate of apoptosis. ASCs viability was measured using MTT assay after induction with liposomes. Morphological changes were shown using a phase-contrast, inverted microscope and Transmission-Electron Microscope (TEM).

Results: The flow cytometry results showed that cells treated with both LSPCE and LPCC showed increase in early apoptosis beginning at 6 hr after incubation, which was confirmed by caspase 3 measurement. MTT assay showed that both LSPCE and LPCC could decrease viability of cells. Cells treated with LSPCE and LPCC showed some rounded cells, which was an early sign of cell death. Cells treated with SD showed extensive membrane damage with necrosis features using TEM.

Conclusion: The results above demonstrated that LSPCE induced apoptosis of ASCs

Keywords: Adipose-derived stem cells, Apoptosis, Liposomes, Phosphatidylcholines

Introduction

Although liposuction is highly efficient for correcting unwanted localized fat deposits, injection of lipolysis has advantages of minimal downtime, less invasiveness, and nonsurgical therapies. During the past decades, lipolysis injections using Phosphatidylcholine (PC), Sodium Deoxycholate (SD), or combination of both have been frequently used in certain countries such as United States, South America, and Europe. 1,3 Phosphatidylcholine from various sources has been used as an active agent in the formulation of SD containing lipolysis injection of solutions intended to reduce subcutaneous fat 1,3, which is associated with inflammatory reaction due to the SD content 6. PC-SD has not been approved by Food and Drug Administration (FDA) but used as a lipolysis procedure due to lack of clear mechanism that is responsible for fat loss and safety concern. SD was confirmed to induce cell death and USFDA-approved with the name...
Kythera® as a treatment only for adults with moderate-to-severe fat below the chin, but it was still considered off-label because of SD side effects such as pain, redness and inflammation of the skin. SD as a detergent emulsifier causes cell and tissue necrosis and inflammation.

The effect of PC on fat cells has been studied, but its mechanism needs to be further investigated. Klein et al reported that Bovine Serum Albumin (BSA) solubilized PC has less effect on fat cell viability. A recent study confirmed the role of PC that could induce apoptosis in adipocyte cell culture, so PC alone might be developed into a safer lipolytic agent that causes less necrosis, and thus less inflammation and side effects. PC is not soluble in water, so it is a main problem to investigate PC’s efficacy. Adding SD or BSA may interfere with PC’s effect as reported in another study.

Recently, Purwoko et al have successfully prepared liposome that was made from Soybean PC (SPC) extract, which made the PC soluble without addition of other substances. This SPC extract liposome was reported in an animal study and was shown to be effective to reduce mice fat cells compared to negative control, but the mechanism needs to be further investigated. The aim of this study was to analyze the mechanism of Liposomal Soybean PC (LSPC) extract from Indonesian soybeans (containing 26% PC) to induce apoptosis of Adipose-derived Stem Cells (ASCs) in vitro.

Materials and Methods

Ethics approval
Adipose-derived stem cells were obtained from patients who underwent liposuction at érpour Medical-Spa Skin and Mesotherapy Centre, Jakarta after signing a written informed consent. This study was approved by the Ethics Committee for Medical Research, Faculty of Medicine, Universitas Indonesia (No. 255/H2.F1/ETIK/2013).

Materials
LSPC was extracted from soybean seeds of Argomulyo variety purchased from the Indonesia Research Institute for Legumes and Tuber Crops (ILETRI), Malang, East Java Province. Purified PC of >99% purity (L-α-phosphatidylcholine) was purchased from Sigma-Aldrich, USA. Solutions of PC and SD (Dermastabilon® injection) and 2% SD (Deoxylise®) were imported from Aesthetic Dermal, Girona, Spain.

Liposome preparation
Extraction and fractionation of soybean PC was done in the Agro Industrial Technology Development Biomedical Laboratory, South Tangerang, Banten Province, Indonesia. The final product consisted of 26% SPC by High Performance Liquid Chromatography (HPLC). Liposome preparation was done in the Laboratory of Medical Pharmacy, Faculty of Medicine, Universitas Indonesia, Jakarta using thin film hydration (hand-shaking) method followed by a stepwise extrusion process through polycarbonate filters with 100 nm probes in a Liposofast syringe. The characteristics showed that median diameter of liposomes from SPC extract (LSPCE) was 48.9 nm (10th-90th percentiles, 41.0-71.3 nm) with polydispersity index of 0.35. They were anionic liposomes with a mean zeta potential of -17.25 mV. Liposomes from purified PC (LPPC) were larger with a median diameter of 68.3 nm (10th-90th percentiles, 55.7-96.8 nm), with a polydispersity index of 0.16. The liposomes were cationic with a mean zeta potential of 58.23 mV. The presence and morphology of small unilamellar liposomes were further confirmed by transmission electron.

Isolation, culture, and characterization of ASCs
ASCs were harvested from liposuction aspirate. Isolation and culture of the ASCs were done according to a recently published protocol. In brief, lipoaspirate was filtered using a fine-mesh, stainless-steel tea filter and was extensively washed in Phosphate-Buffered Saline (PBS pH=7.4) until it became clear and clean. Enzymatic breakdown was induced by adding 0.075% collagenase type I (Sigma, St. Louis, Missouri, USA). The tubes were incubated at 37°C for one hour and were agituated every 5 min. The floating yellowish lipid and remnants were discarded and the infranatant was transferred into a sterile 15 ml centrifuge tube and centrifuged at 1200 rpm for 10 min. Cells were resuspended in low-glucose Dulbecco’s Modified Eagle’s Medium (DMEM) supplemented with L-glutamine, 10% Plasma-Rich Platelets (PRP), 1% amphotericin B, 1% penicillin, 1% streptomycin, and 1% heparin at 37°C in a humidified atmosphere containing 5% CO2. Cells were seeded in 12-well plates at a density of 105 cells/well. Cells were observed and the culture medium was refreshed every 2-3 days. Once the cells had grown to 80% confluence, adherent cells were dissociated (TripLE Select Enzyme, Thermofisher Scientific, USA) and passaged. Cells from the third to fifth passage were collected and used for further tests.

Characterization of ASCs was done using flow cytometry (BD Stemflow™ hMSC Analysis Kit, BD Sciences, USA). Cells from passage 5 were harvested, and washed with PBS and centrifuged at 1200 rpm for 10 min before analysis. The kit components included a series of cocktail antibodies, which are positive (CD73, CD90, CD105, and HLA-DR) for Mesenchymal Stem Cells (MSCs). Based on International Society for Cellular Therapy (ISCT), MSCs should express CD105, CD73 and CD90 (≥95%) and lack the expression of CD45, CD34, CD14 or CD79a, and CD73, CD90, and HLA-DR surface molecules (<2%) by transmission electron.

Flow cytometric (FCM) analysis of apoptosis
Cells of the third passage were seeded in a 12-well plate at a density of 150,000 cells per well. On the next day, the culture medium was replaced with media containing...
Apoptosis of ASCs Induced by LSPCE

Results

ASCs isolation, culture, and characterization

Isolations of the ASCs were achieved from six liposapirate donors. The cultured ASCs had fibroblast-like morphology and were adherent to the plastic culture ware from the second day. The cells underwent proliferation and were confluent between days 7 and 12 (Figure 1). In flow cytometric analysis, the ASCs showed phenotypes of MSCs as follows: CD73+ 99.2%; CD90+ 99.8%; and CD105 + 90.4% (see Supplement Materials). Only 1.8% of the cells expressed hematopoetic cell surface markers. Multipotency assessment of the ASCs showed that these cells were capable of differentiation into chondrocyte, osteocyte and adipocytes.

Apoptosis induced by LSPCE, LPCC, commercial PC+SD and SD

Flow cytometric analysis of early apoptosis and morphologies of ASCs treated with Liposomal Soy
Phosphatidylcholine Extract (LSPCE), Liposomal Purified PC (LPPC), commercial solution PC+SD 1000 ppm, and SD 400 ppm can be seen in figure 2.

Flow cytometric analysis showed that the mean early apoptotic rate of negative control (cells without treatment) was 1.7%. Cells treated with LSPCE showed low apoptotic rate (lower than negative control) after the 4-hr induction. However, when the induction time was prolonged to 6-8 hr, the rate was slowly increased (Figure 2A). On the other hand, LPPC induced apoptosis after 6 hr of incubation. Compared with the LSPCE, the LPPC produced higher rate of early apoptosis, but then it decreased after 8 hr of incubation (Figure 2B).

Cells treated with commercial PC+SD showed 15.60% apoptotic rate (higher than negative control) after the 2-hour induction. It was slightly down as the induction time was prolonged 4-8 hr and the number of cells was substantially reduced below 10,000 cells (Figure 2C). In other case, cells were treated with SD after 30 min until 8 hr induction showed lower apoptotic rate than negative control (Figure 2D). Cells decreased with longer time induction.

Morphological features of adipose-derived stem cells after apoptosis induction by LSPCE, LPPC, commercial PC+SD and SD can be seen in figure 3. Control cells showed morphologies that were consistent

Figure 1. Morphology of the primary culture of adipose-derived stem cells observed by inverted microscope (magnificent 100x). P0 Day 2: ASCs were fibroblast cell like and attached in well; P0 Day 5: ASCs had been proliferated; P0 Day 12: ASCs had proliferated and confluent.

Figure 2. Early apoptosis assayed using Annexin V-FITC/PI by flow cytometry. The bottom of flow cytometry graphs showed percentage of early apoptosis ASCs. Annexin V-FITC at x-axis and PI at y-axis. Early apoptosis cells observed in right bottom quadrant (Annexin V+/PI -).

A. Early apoptosis observed in ASCs treated with liposomal soybean phosphatidylcholine extract (LSPCE) 1000 ppm.
B. Percentage of early apoptosis observed in ASCs treated with liposomal purified PC (LPPC) 1000 ppm.
C. Percentage of early apoptosis observed in ASCs treated with commercial PC+SD 1000 ppm. After 6 and 8 hr induction number of cells reduced below 10,000 cells (*: 4890 cells and #: 916 cells).
D. Percentage of apoptosis observed in ASCs treated with SD 400 ppm (equals with SD concentration at commercial solution PC+SD 1000 ppm).
with MSCs, i.e. spindle-shape similar to fibroblast (Figure 3A). Cells treated with LSPCE and LPPC showed some rounded cells, which was an early sign of cell death. Cells treated with PC+SD solution showed rounded cells, retracted cells, and disrupted membranes. On the other hand, cells treated with SD were damaged (Figure 3E). Further confirmation using TEM showed that cells treated with LSPCE showed early signs of apoptosis, i.e. nearly fragmented nucleus. As comparison, cells treated with SD showed extensive membrane damage consistent with necrosis features (Figure 4).

Caspase-3 was measured to confirm the apoptotic mode of cell death. Relative concentration of caspase-3 was increasing as the concentration of tested formulations increased. Both liposomes from SPC extract and purified PC showed considerable caspase-3 activation at 1000 ppm. Highest peak was observed in cells treated with PC+SD solution (Dermastabilon®) 1000 ppm. No viable cells were detected after treatment with 500 and 1000 ppm SD. Statistical analysis showed that cells treated with 1000 ppm PC+SD differed significantly with negative control group (p=0.007) (Figure 5).

MTT assay showed that both liposomes of SPC extract and purified PC decreased the viability of ASCs. The effect was increasing with increasing the dose of liposomes (Figure 6). The half maximal effective concentration (EC50) was calculated based on the graph. The effect was stronger with liposomes of SPC extract (EC50 =500.79 ppm) than the liposomes of purified PC (EC50 =755.26 ppm). Statistical analysis showed that 750 ppm, 1000 ppm and 1500 ppm of LSPCE group differed significantly with LPPC group with the same concentration (p<0.05, independent t test).

**Discussion**

Apoptosis can be detected by many tests. Some of
the tests did not measure cell death only, but at the same time measured common physiological (non-lethal) processes. Consequently, more than one method of apoptosis analysis and right timing to apoptosis test were needed. In this study, flow cytometry, ELISA, and microscopic analysis were used to detect apoptosis. Flow cytometry is one of the best techniques to detect cell death due to the possibility to use several markers and to obtain a quantitative result. Generally, apoptosis detection timing in cell culture (5-10 hr) is faster than in normal tissue (11-14 days). Timing choices to conduct an apoptosis analysis in culture depend on the type of cell, stimulant, concentration and time of induction.

Cells treated with LSPCE showed lower early apoptotic rate compared to negative control after 4-hr incubation, thus showed normal variation. However, when the induction time was prolonged to 6 and 8 hr, the rate was slowly increased, which suggests that treatment with LSPCE caused late onset of early apoptosis. Further, LPPC could induce apoptosis after 6 hr of incubation, and produced higher rate of early apoptosis compared to the LSPCE; but then apoptosis decreased after 8 hr of incubation (Figure 2). This fact suggested that LPPC induced much higher early apoptosis, which occurred late but massive and brief in this period of time. Apoptotic rates beyond 8 hr were not measured. Therefore, cells treated with LSPCE or LPPC showed increase in early apoptosis beginning at 6 hr after incubation. However, LPPC produced higher rate of early apoptosis compared to LSPCE. Cells treated with the commercial solution of PC+SD induced apoptosis in the first few hours, and at the same time a massive non-apoptotic cell death (probably necrosis) occurred that proved the toxicity of this formulation on the ASCs. Further, the remaining cells after two hours of incubation were far from enough to be analyzed properly. Cells treated with SD only showed apoptotic rate lower than negative control and it suggested that SD did not induce apoptosis.

LSPCE showed higher caspase-3 concentration than LPPC, suggesting that apoptosis occurs at higher rate in cells treated with LSPCE. However, FCM analyses showed that apoptosis was lower in cells treated with LSPCE. This discrepancy might be due to the fact that FCM analysis was stopped after 8 hr, while caspase-3 measurement was done on other cultures that were incubated overnight. It seems that apoptosis of cells treated with LSPCE continues after 8 hr. On the other hand, LPPC had a shorter effect and probably produced less apoptotic cells. Therefore, in this study, it was shown that liposomal formulation of soybean PC can induce apoptosis in ASCs culture.

A recent study in 3T3-L1 cells proved that PC was able to induce pre-adipose and adipose cell apoptosis. Li et al also found that PC in the form of liposome (LPCC) without SD induced apoptosis of MSCs. This fact will be used as an appropriate strategy to prevent or inhibit the occurrence of obesity.

Our study showed that LSPCE induced ASCs apoptosis at morphological observation with an inverted microscope and flow cytometry analysis. Compared to PC+SD or SD only, cells treated with LSPCE or LPPC showed lower rate of apoptosis. This fact means that SD is the most potent agent to induce cell death. SD content in the preparation of PC was suspected as the main active ingredient for cell death through necrosis. El Kamshousy et al reported that injection of SD as much as 2% or 20 mg/ml not only caused fat cell necrosis, but also resulted in necrobiosis of the lower dermis tissue adjacent to the subcutaneous fat tissue. Although both LSPCE and LPPC showed lower rate of apoptosis, they are expected to be more safe. Our study showed that LSPCE induced ASCs apoptosis more effectively than LPPC because LPPC induced much higher apoptosis, but this effect occurred late and brief in the period of time. Concentration of caspase-3 was higher in cells that were treated with LSPCE than LPPC at 1000 ppm. However, analysis by flow cytometry-Annexin V assay showed that reaction to LSPCE was slower than LPPC, but the cumulative effect for 24 hr was higher than LPPC.
Our study showed that at a concentration of 1000 ppm, PC+SD worked faster and stronger than preparation of liposomes (LSPCE or LPPC). This fact might be due to the nature of PC+SD that was soluble in water and contained more PC than LSPCE. PC+SD has a possibility to work directly through Fas receptor (FasR) on cell membrane, which might be activated. Activation of the receptor can induce intracellular transduction signal to apoptosis. SD works by making holes in cell membrane and increases the permeability of mitochondrial membrane. Therefore, PC+SD can induce intrinsic apoptosis pathway. On the contrary, LPPC or LSPCE took longer time to internalize and trigger apoptosis.

Effectiveness of particles to induce cell death was affected by many factors, such as composition, surface charge, particle shape and size. Soy isoflavones composition in LSPCE that induced apoptosis had been reported by many studies and one of those studies was done, such as 5 nm titanium dioxide particle (TiO2) that was shown to be more effective to induce apoptosis in mice proteoblast than 32 nm TiO2. Therefore, the smaller size and existence of a synergistic effect between PC and isoflavones on LSPCE might induce cell death more effectively than LPPC.

**Conclusion**

In summary, our results indicated that Indonesian soybean is a potential source for therapeutic agent. From Indonesian soybean, chemically well characterized purified Indonesian soybean extract that contains PC (PSE-PC) with sufficient level of Soybean Phosphatidylcholine (SPC) can be produced and later can be prepared as physically stable LSPCE. LSPCE can induce ASCs death through apoptotic pathway with higher therapeutic potency than PC+SD. LSPCE induced less necrosis than PC+SD and SD.

**Acknowledgement**

This work was supported by grants from DRPM Universitas Indonesia. Authors thank Dr. Levina S. Pakasi for the English preparation of this manuscript.

**References**


