

Altered miR-223 Expression in Sputum for Diagnosis of Non-Small Cell Lung Cancer

Abouzar Bagheri¹, Hamid Reza Khorram Khorshid¹, Seyed Javad Mowla², Hassan Ali Mohebbi³, Azam Mohammadian⁴, Mehdi Yaseri⁵, Masoud Solaymani-Dodaran⁶, Masih Sherafatian², and Mahmood Tavallaie^{7*}

1. Genetic Research Center, University of Social Welfare and Rehabilitation Sciences, Tehran, Iran

2. Department of Molecular Genetics, Faculty of Biological Sciences, Tarbiat Modares University, Tehran, Iran

3. Trauma Research Center, Baghiyatallah University of Medical Sciences, Tehran, Iran

4. Department of Chemistry, Faculty of Science, Ferdowsi University of Mashhad, Mashhad, Iran

5. Department of Epidemiology and Biostatistics, Tehran University of Medical Sciences, Tehran, Iran

6. Iran University of Medical Sciences, Tehran, Iran

7. Human Genetic Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran

Abstract

Background: Diagnosis of Non-small Cell Lung Cancer (NSCLC) at an early stage is a daunting challenge due to the deficiency of specific noninvasive markers. MicroRNAs (miRNAs) play important roles in the initiation and progression of NSCLC. Measuring miRNA expression levels could provide a potential approach for the diagnosis of NSCLC. Our goals were to examine miR-223, miR-212, miR-192, miR-3074, SNORD33 and SNORD37 expression levels in tissue and sputum of NSCLC patients and cancer free subjects for molecular diagnosis of NSCLC.

Methods: Relative expressions of miR-223, miR-212, miR-192, miR-3074, SNORD33 and SNORD37 were examined with quantitative real-time RT-PCR assay in tissue and sputum obtained from 17 NSCLC patients and 17 controls.

Results: miR-3074 was upregulated in tissue samples of NSCLC patients compared with control group. miR-223 was upregulated, miR-212 and SNORD37 were downregulated in sputum samples of patients compared with controls. miR-223 quantification produced 82% sensitivity and 95% specificity with areas under the ROC curve at 0.90 in detection of NSCLC.

Conclusion: miR-223 clearly discriminated cancer patients from cancer-free subjects and our results suggest that miR-223 could be a diagnostic useful biomarker. The measurement of altered miRNA expression in sputum samples manifested the potential noninvasive approach for detection of lung cancer.

* Corresponding author:
Mahmood Tavallaie, Ph.D.,
Human Genetic Research Center,
Baqiyatallah University of
Medical Sciences, Tehran, Iran
Tel: +98 9121055684
Fax: +98 21 88053609
E-mail:
tavalla.mah@gmail.com
Received: 19 Oct 2016
Accepted: 20 Dec 2016

Avicenna J Med Biotech 2017; 9(4): 189-195

Keywords: MicroRNAs, Non-small cell lung carcinoma, Sputum

Introduction

Lung cancer is the most common cause of cancer death worldwide¹. Unfortunately, initial symptoms of lung problems appear when the disease is in an advanced stage². Non-small Cell Lung Cancers (NSCLC) account for ~80-85% of all lung cancer cases³. More than 75% of NSCLCs are diagnosed when the disease is locally advanced or metastatic. This fact represents a current 5-year survival of less than 15%^{4,5}. Therefore, finding NSCLC in early stages is a realistic approach to reduce the mortality associated with NSCLC. While computed tomography seems hopeful in detection of NSCLC at a smaller size compared to a chest X-ray, the improved sensitivity is related to an increased false-positive rate. The fluorescence bronchoscopy exceeds at diagnosing centrally-located lung tumors. However,

it is an invasive technique^{6,7}. The development of highly valid and noninvasive diagnostic procedure would simplify the early detection of NSCLC, which is clinically meaningful.

Small non-coding RNAs (sncRNAs) mainly consist of microRNAs (miRNAs) and small nucleolar RNAs (snoRNAs)⁸. miRNAs can post-transcriptionally regulate the expression of myriad of different target genes including more than 30% of protein coding genes⁹, thereby managing an extensive spread of biological functions such as cellular proliferation¹⁰, apoptosis¹¹ and differentiation¹². Scientific emerging evidences suggest the potential involvement of altered miRNA expressions in the pathogenesis of human cancers¹³⁻¹⁵. miRNAs may function as tumor suppressors or onco-

genes, thus dysregulated expressions participate in cancer development and progression^{16,17}. Consequently, miRNAs can potentially be useful in the detection, classification, prognosis, and therapy of human malignancies¹⁸.

Recently, new and unexpected functions of other types of small ncRNAs have been discovered and investigators found that snoRNA expression in cancers is as variable as miRNA expression¹⁹. Some snoRNAs could be processed to produce molecules like miRNA which drive post-translational gene silencing on complementary mRNAs²⁰⁻²². Expression of snoRNAs could be detected in biological fluids, making them potentially applicable biomarkers²³.

Sputum is the most fluently accessible specimen that includes the pathogenically relative cell types; furthermore, collecting sputum is noninvasive, fast, and economical, which are prominent features to be an ideal sample type for population screening. These characteristics cause molecular analysis of sputum to be an important target for the investigation of lung cancer biomarkers²⁴.

It is hypothesized that simultaneous assessment of a panel of ncRNAs could provide a highly sensitive and specific diagnostic test for detection of NSCLC. To verify the hypothesis, a panel of significant ncRNAs, including miR-223, miR-212, miR-192, miR-3074, SNORD33 and SNORD37 was selected to analyze tissue and sputum of NSCLC patients.

Materials and Methods

To determine the clinical significance of dysregulated expressions of ncRNAs in tissue and sputum for diagnosis of NSCLC, expression changes of 6 cancer-associated ncRNAs, miR-223, miR-212, miR-192, miR-3074, SNORD33 and SNORD37, in 17 NSCLC patients and 17 cancer free subjects were evaluated.

Sample collection

Lung tissue and sputum samples were collected from patients at Masih Daneshvari and Baghiat Allah Hospitals (Tehran, Iran). The research has been performed in accordance with the Declaration of Helsinki and has been approved by Ethics Committee of the University of Social Welfare and Rehabilitation Sciences, Tehran, Iran. All the participants agreed to the research plan and signed the written consent form and ethics permission was obtained for the research on samples.

Subjects in this study, 17 NSCLC patients 51-73 years old, had histopathologically confirmed primary NSCLC, stages I-IV, and medical history information and 17 cancer-free controls were sex, and age matched

to the patients group (Table 1).

Lung tissue specimens were immediately immersed in RNAlater buffer (Applied Biosystems, USA) and stored at -80°C for RNA extraction. Prior to the collection of a sputum sample, patients rinsed their mouths with water, breathed deeply, held their breath and coughed. All expectorated sputum were collected into a sterile plastic sample container that was then sealed and stored at -80°C until further processing. Routine sputum cytology was not performed on the collected sputum samples because previous studies have shown that sputum cytology has a high rate of both false positives and false negatives^{25,26}.

RNA isolation

1 ml of TRIzol (Ambion, USA) and 750 µl of TRIzol-LS (Ambion, USA) were added to the individual homogenized tissue and sputum samples, respectively. Samples were then reacted at room temperature for 5 min. Chloroform was added to extract RNA then 500 µl of isopropanol was added to precipitate RNA, which was then washed with 75% EtOH. RNA was then dissolved in nuclease-free water. The concentration and purity of the isolated RNA were determined by a NanoDrop and the integrity of the RNA was verified using RNase-free agarose gel electrophoresis.

cDNA synthesis and real-time RT-PCR

Poly (A) tailing of RNA was performed by *Escherichia coli* (*E. coli*) poly (A) polymerase kit (New England Biolabs, UK), then reverse transcription reaction was carried out by anchored oligo (dT) primer (Table 2) and a reverse transcriptase kit (Thermo Scientific, USA). cDNA synthesis parameters were as follows: 42°C for 60 min and 70°C for 10 min. Quantitative real-time RT-PCR was performed using EvaGreen master mix (Solis BioDyne, Estonia) and specific primers for miR-223, miR-212, miR-192, miR-3074, SNORD33 and SNORD37 (Table 2). The PCR parameters were as follows: initial denaturation (one cycle at 95°C for 15 min); 40 cycles of denaturation, amplification, and quantification (95°C for 15 s, 58-64°C for 30 s, and 72°C for 5 s); and the melting curve (starting at 65°C and gradually increasing to 95°C). The miRNA expression was normalized to the levels of U6, and expression differences were calculated according to the standard curve and efficiency established for each primer set.

Statistical analysis

All statistical analysis performed by R (R Core Team (2014), R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria, URL <http://www.R-project.org>). P-value of <0.05 was considered statistically sig-

Table 1. Characteristics of 17 NSCLC patients and 17 cancer-free controls

	Sex		Age (year)	Histologic types		Stage			
	Male	Female	-	SCC	AC	I	II	III	IV
Cancer patients	15	2	51-73	6	11	2	3	5	7
Cancer-free controls	15	2	48-71	--	--	--	--	--	--

Table 2. Primer sequences used in real-time RT-PCR analysis

Name	Forward	Reverse
miR-192	GTGAGCTGACCTATGAATTGACA	GCGAGCACAGAATTAATACGAC
miR-3074	ACCATTCTGCTGAACTGAG	GCGAGCACAGAATTAATACGAC
SNORD37	CACGATGTCTACTGAAGAAAGCCTG	GCGAGCACAGAATTAATACGAC
SNORD33	TTTCCCGACCATGAGATGAC	GCGAGCACAGAATTAATACGAC
U6	TTTCGCAAGGATGACACGC	GCGAGCACAGAATTAATACGAC
miR-223	(Pg4487-03, Parsgenome, Iran)	
miR-212	(Pg4487-03, Parsgenome, Iran)	
Anchored Oligo (dT) Primer	GCGAGCACAGAATTAATACGACTCACTATAGG (32bp) (T)12VN*	

* V= G, A, C; N= G, A, T, C.

Table 3. Evaluated ncRNAs in NSCLC patients and cancer-free controls in sputum samples

ncRNA	p-value*	Fold change (cancer/control)	AUC (95% CI)	Cutoffs	Sensitivity	Specificity
miR-223	<0.05	19.87	0.90 (0.81-0.99)	4.83	82%	95%
miR-212	<0.05	0.21	0.69 (0.53-0.85)	0.65	45%	91%
miR-3074	>0.05	0.89	0.65 (0.51-0.79)	0.44	93%	40%
miR-192	>0.05	1.13	0.61 (0.47-0.76)	0.4	96%	30%
SNORD33	>0.05	1.67	0.68 (0.55-0.82)	0.56	43.3%	96.3%
SNORD37	<0.05	0.07	0.82 (0.72-0.93)	0.78	93.3%	63.3%

* The p-values are based on Mann-Whitney test.

nificant. Kolmogorov Smirnov test as well as Q-Q plot were applied to check the normal distribution of data. To compare the ncRNAs between cancer patients and controls, Mann-Whitney test was used. Also, to test the difference adjusted for the effect of age, sex, smoking and clinicopathologic characteristics, logistic regression was used. Furthermore, Receiver-Operator Characteristic (ROC) curve analysis was undertaken for each gene in the sputum specimens from cancer patients and cancer-free controls. Also, the Areas Under the ROC Curve (AUROCs) were calculated and the optimal threshold was chosen based on Youden's J statistics, then sensitivity and specificity as well as diagnostic accuracy levels to distinguish control individuals from cancer patients, and corresponding thresholds were calculated for each ncRNA. To obtain the best combination of ncRNAs that can distinguish the cancer patients from controls, logistic regression was used.

Results

ncRNAs as biomarkers in sputum samples of NSCLC patients

miR-223 significantly increased in sputum of cancer patients compared to non-cancers ($p < 0.05$). miR-223 overexpression resulted in 82% (95% CI, 0.63-1.00) sensitivity and 95% (95% CI, 0.86-1.00) specificity in the diagnosis of NSCLC. miR-212 significantly decreased in sputum of patients compared to controls ($p < 0.05$). miR-212 underexpression resulted in 68% (95% CI, 0.46-0.90) sensitivity and 64% (95% CI, 0.41-0.87) specificity in the diagnosis of NSCLC. SNORD37 significantly decreased in sputum samples of NSCLCs compared to controls and resulted in 93.3% (95% CI, 0.46-0.90) sensitivity and 63.3% (95% CI, 0.41-0.87) specificity in the diagnosis of NSCLC

($p < 0.05$). miR-192, miR-3074 and SNORD33 did not alter in sputum of patients compared to controls ($p > 0.05$), (Table 3). Prevalence of miR-223, miR-212, miR-192, miR-3074, SNORD33 and SNORD37 expressions detected in sputum was not associated with patient age, gender, histological tumor type and stage ($p > 0.05$).

ncRNAs as biomarkers in tissue samples of NSCLC patients

miR-223, miR-212, miR-192, SNORD33 and SNORD37 did not alter in tissue samples of cancer patients compared to non-cancers. Expression of miR-3074 significantly increased in tissue samples of cancer patients compared to non-cancers. miR-3074 overexpression resulted in 53% (95% CI, 0.63-1.00) sensitivity and 86% (95% CI, 0.86-1.00) specificity in the diagnosis of NSCLC ($p < 0.05$) (Table 4). The prevalence of miR-223, miR-212, miR-192, miR-3074, SNORD33 and SNORD37 expression in tissue samples was not associated with patient age, gender, histological tumor type and stage ($p > 0.05$).

Genetic changes in NSCLC patients and cancer-free individuals

The best AUC in our research belonged to miR-223 in sputum samples. Sensitivity and specificity of three significant biomarkers (miR-223, miR-212 and SNORD37) as a panel were not distinguishable from miR-223 alone in the diagnosis of NSCLC, sensitivity 82% (95% CI, 0.63-1.00) and specificity 95% (95% CI, 0.86-1.00) and AUROC at application of combined miR-223, miR-212 and SNORD37 in comparison to solitary miR-223 was not significant. Figure 1 shows ROC curve with corresponding AUROC for miR-223, miR-212 and SNORD37 expressions in sputum from cancer patients versus non-cancers.

Noninvasive Diagnosis of Non-Small Cell Lung Carcinoma

Table 4. Evaluated ncRNAs in NSCLC patients and cancer-free controls in tissue samples

ncRNA	p-value *	Fold change (cancer/control)	AUC (95% CI)	Cutoffs	Sensitivity	Specificity
miR-223	>0.05	0.92	0.65 (0.50-0.79)	0.51	70%	56.7%
miR-212	>0.05	0.65	0.62 (0.47-0.77)	0.55	46.7%	93.3%
miR-3074	<0.05	3.6	0.73 (0.58-0.84)	0.71	86%	53.3%
miR-192	>0.05	0.79	0.47 (0.32-0.63)	0.53	73.3%	46.7%
SNORD33	>0.05	1.31	0.68 (0.53-0.83)	0.48	66.7%	86.7%
SNORD37	>0.05	1.07	0.55 (0.4-0.7)	0.5	80%	40%

* The p-values are based on Mann-Whitney test.

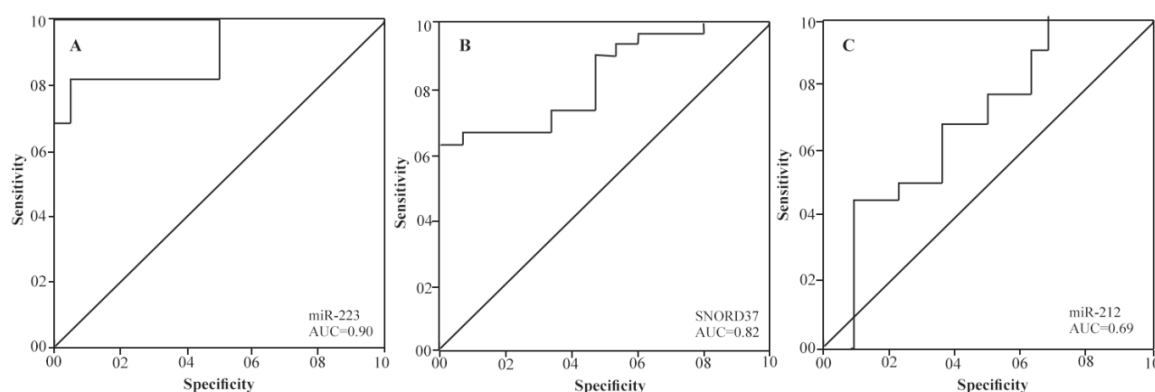


Figure 1. Receiver-operator characteristic (ROC) curve analysis of expression levels of the two miRNAs in sputum of 17 patients diagnosed with NSCLC and 17 healthy individuals. The area under the ROC curve (AUC) for each biomarker conveys its accuracy in distinguishing cancer-free subjects from cancer patients in terms of sensitivity and specificity. Significant genes produce (A, miR-223) 0.90 (95% CI, 0.81-0.99), (B, SNORD37) 0.82 (95% CI, 0.72-0.93) and (C, miR-212) 0.69 (95% CI, 0.53-0.85) AUC values.

Discussion

Early diagnosis of NSCLC could change the disease outcome; actually, the survival rate will increase significantly. Many imaging and cytology-based strategies have been employed to augment early detection; however, because of low sensitivity or supererogatory cost, none has yet been highly efficient. Our current study clearly shows for the first time that alteration of miR-223 expression in sputum would provide a useful biomarker for noninvasive diagnosis of NSCLC.

Elevated miR-223 expression occurs in sputum of NSCLC patients with equal frequency among all histologic types of lung tumor, suggesting that the genetic changes are not specific to histologic type. Detection of the abnormality may be useful in determining different types of NSCLC that is really important since lung adenocarcinomas, which originate from the smaller peripheral airways, are difficult to be detected by bronchoscopy or sputum cytology and have become more prevalent than other types of lung cancer.

Ever since the first miRNA was discovered in *C. elegans* and was found to have an essential role in the worm development^{27,28}, it is a widely accepted concept that miRNAs are remarkable regulatory factors in development, apoptosis, and disease generation and progression²⁹. miRNAs participate in keeping the balance of genes regulating pathways that determine the cells' fate. Deregulation of miRNAs incredibly withers this balance, thereby contributing to oncogenesis from initiation to metastasis.

Since there is no single validated molecular biomarker for early lung cancer detection and spectrum of biomarkers is needed for early diagnosis, a panel of biomarkers including miR-223, miR-212, miR-3074, miR-192, SNORD33 and SNORD37 were the candidates in our study.

miR-3074 is a less investigated member of miR-23b cluster that its dysregulation has been reported in various cancers³⁰⁻³⁴. In our research, just expression of miR-3074 was significantly different in tissue samples of NSCLC patients compared with control group. However, in sputum samples, 3 biomarkers miR-223, miR-212 and SNORD37 significantly altered between cancer and control group.

miR-223 has been found to affect the cell cycle by regulating E2F1³⁵, migration and invasion in cancer cells by targeting EPB41L3³⁶, proliferation and tumor growth of cells by targeting IGF1R and downstream Akt/mTOR/p70S6K signaling pathway^{37,38}. miR-223 could act as a signal in the crosstalk between tumor and immune cells in the tumor microenvironment which leads to increased invasiveness in the cancer cells³⁹ or mediating immune evasion mechanisms⁴⁰. miR-223 affects different target genes at multiple cancers like Artemin (oesophageal carcinoma)⁴¹, C/EBP β (leukaemia)⁴², E2F1 (leukaemia)³⁵, EPB41L3 (gastric cancer)³⁶, Fbxw7/Cdc4 (leukaemia, gastric cancer, oesophageal squamous cell carcinoma)^{43,44}, FOXO1 (colorectal cancer cells)⁴⁵, HSP90B1 (osteosarcoma)⁴⁶, IGF1R (HeLa, leukemia and hepatoma cells)^{37,47}, SEPT6

(prostate cancer)⁴⁸, LMO2 (Leukaemia/lymphoma)⁴² and NFI-A (Leukaemia/lymphoma)⁴⁹. Because of important roles of miR-223, this biomarker was selected for investigation in NSCLC patients. In our research, miR-223 had the highest AUC between candidate biomarkers and significantly increased in patient group, but AUC 0.9 was not enough to be applied individually for diagnosis of NSCLC.

miR-212 was found to be dysregulated in many cancers: oral squamous cell carcinoma⁵⁰, colorectal carcinoma⁵¹, gastric cancer⁵², NSCLC⁵³ and head and neck squamous cell carcinoma⁵⁴ and recently, important biological functions in lung cancer cells for miR-212 has been proved¹⁷. It had been reported that miR-212 was involved in cell cycle¹⁷, DNA methylation⁵², cell apoptosis⁵³, and signaling pathways^{55,56}. miR-212 significantly decreased in sputum of our lung cancer patients but sensitivity and specificity in the diagnosis of NSCLC were not sufficient for clinical trials; in addition, miR-212 did not improve compound sensitivity and specificity.

Several investigations suggest that snoRNAs exhibit differential expression in lung tumor and can affect cell transformation, tumorigenesis, and metastasis of NSCLC. SNORD33 and SNORD37 are located on chromosome 19q13.3 and 19p13.3, respectively that contain potential oncogenes involved in malignancies, including lung cancer^{20,57,58}. This study for the first time showed that SNORD37 significantly decreased in sputum samples of NSCLC patients and produced AUC=0.82 in distinguishing patient group from normal individuals. However, SNORD37 did not increase final sensitivity and specificity as a panel with miR-223 and miR-212.

Another characteristic of miRNAs unlike mRNAs is prominent stability in different kinds of biological specimens like urine, serum, plasma, saliva, sputum, formalin-fixed, paraffin-embedded clinical tissues and fresh snap-frozen materials⁵⁹⁻⁶⁴. This prominent stability is due to their resistance to endogenous and exogenous RNase activity, extreme temperatures and pH, long storage in frozen conditions, and repeated freeze-thaw cycles^{61,63,65}. These features introduce miRNA as a great target for different aspects of biological and medical investigations. Although miRNA has recently emerged as a powerful molecular biomarker for detection of diseases like cancers, its potential as a sputum-based biomarker has not been fully explored.

Sputum has the benefits as a potential surrogate substance for molecular genetic diagnosis of lung cancer, because its non-invasive procurement would allow the comprehensive analysis of tumors without the requirement of invasive procedures, such as biopsy or surgery, and the fact that it contains clinically worthy lung and lower respiratory tract bronchial epithelial cells adds to its benefit. Furthermore, sputum has low cost and sample management, including sample collection and processing is simple⁶⁶.

Conclusion

Although assessment of miR-223 expression in sputum seems to be hopeful in the noninvasive detection of lung cancer, 82% sensitivity and 95% specificity are not efficient for routine clinical application. In this study, although the sputum and tissue levels of some biomarkers in NSCLC patients were analyzed at different stages, the number of patients was small and the number of biomarkers tested was limited. In the future investigation, more samples especially early-stage samples should be accessed to evaluate the role of sputum miRNAs associated with NSCLC. The outcome might indicate the need to develop a strategy for simultaneous evaluation of a panel of tumor-specific miRNA biomarkers in sputum in order to attain an extremely sensitive and specific diagnostic test for lung cancer.

Acknowledgement

We would like to thank Masih Daneshvari and Baghiat Allah Hospitals (Iran) for providing the sputum and tissue samples. The study was supported by the University of Social Welfare and Rehabilitation Sciences, Tehran, Iran.

Conflict of Interest

All the authors declare that they have no competing interests.

References

1. Garon EB, Rizvi NA, Hui R, Leighl N, Balmanoukian AS, Eder JP, et al. Pembrolizumab for the treatment of non-small-cell lung cancer. *N Engl J Med* 2015;372(21): 2018-2028.
2. D'Urso V, Doneddu V, Marchesi I, Collodoro A, Pirina P, Giordano A, et al. Sputum analysis: non-invasive early lung cancer detection. *J Cell Physiol* 2013;228(5):945-951.
3. D'addario G, Früh M, Reck M, Baumann P, Klepetko W, Felip E, et al. Metastatic non-small-cell lung cancer: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 2010;21 Suppl 5:v116-v119.
4. Lewis PD, Lewis KE, Ghosal R, Bayliss S, Lloyd AJ, Wills J, et al. Evaluation of FTIR spectroscopy as a diagnostic tool for lung cancer using sputum. *BMC Cancer* 2010;10:640.
5. National Lung Screening Trial Research Team, Aberle DR, Berg CD, Black WC, Church TR, Fagerstrom RM, et al. The national lung screening trial: overview and study design. *Radiology* 2011;258(1):243-253.
6. Toloza EM, Harpole L, McCrory DC. Noninvasive staging of non-small cell lung cancer: a review of the current evidence. *Chest* 2003;123(1 suppl):137S-146S.
7. Hirsch FR, Franklin WA, Gazdar AF, Bunn PA Jr. Early detection of lung cancer: clinical perspectives of recent advances in biology and radiology. *Clin Cancer Res* 2001;7(1):5-22.

8. Su Y, Guarnera MA, Fang H, Jiang F. Small non-coding RNA biomarkers in sputum for lung cancer diagnosis. *Mol Cancer* 2016;15(1):36.
9. MacFarlane LA, Murphy PR. MicroRNA: biogenesis, function and role in cancer. *Curr Genomics* 2010;11(7): 537-561.
10. Bueno MJ, Pérez de Castro I, Malumbres M. Control of cell proliferation pathways by microRNAs. *Cell Cycle* 2008;7(20):3143-3148.
11. Jovanovic M, Hengartner MO. miRNAs and apoptosis: RNAs to die for. *Oncogene* 2006;25(46):6176-6187.
12. Lee CT, Risom T, Strauss WM. MicroRNAs in mammalian development. *Birth Defects Res C Embryo Today* 2006;78(2):129-139.
13. Guz M, Rivero-Müller A, Okoń E, Stenzel-Bembenek A, Polberg K, Słomka M, et al. MicroRNAs-role in lung cancer. *Dis Markers* 2014;2014:218169.
14. Visone R, Croce CM. MiRNAs and cancer. *Am J Pathol* 2009;174(4):1131-1138.
15. Iorio MV, Croce CM. MicroRNA dysregulation in cancer: diagnostics, monitoring and therapeutics. A comprehensive review. *EMBO Mol Med* 2012;4(3):143-159.
16. Yanaihara N, Caplen N, Bowman E, Seike M, Kumamoto K, Yi M, et al. Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* 2006;9(3):189-198.
17. Jiang X, Chen X, Chen L, Ma Y, Zhou L, Qi Q, et al. Upregulation of the miR-212/132 cluster suppresses proliferation of human lung cancer cells. *Oncol Rep* 2015; 33(2):705-712.
18. Balgkouranidou I, Liloglou T, Lianidou ES. Lung cancer epigenetics: emerging biomarkers. *Biomark Med* 2013;7 (1):49-58.
19. Gee H, Buffa FM, Camps C, Ramachandran A, Leek R, Taylor M, et al. The small-nucleolar RNAs commonly used for microRNA normalisation correlate with tumour pathology and prognosis. *Br J Cancer* 2011;104(7):1168-1177.
20. Liao J, Yu L, Mei Y, Guarnera M, Shen J, Li R, et al. Small nucleolar RNA signatures as biomarkers for non-small-cell lung cancer. *Mol Cancer* 2010;9:198.
21. Brameier M, Herwig A, Reinhardt R, Walter L, Gruber J. Human box C/D snoRNAs with miRNA like functions: expanding the range of regulatory RNAs. *Nucleic Acids Res* 2011;39(2):675-686.
22. Ender C, Krek A, Friedländer MR, Beitzinger M, Weinmann L, Chen W, et al. A human snoRNA with microRNA-like functions. *Mol Cell* 2008;32(4):519-528.
23. Crea F, Clermont PL, Parolia A, Wang Y, Helgason CD. The non-coding transcriptome as a dynamic regulator of cancer metastasis. *Cancer Metastasis Rev* 2014;33(1):1-16.
24. Kim CE, Tchou-Wong K-M, Rom WN. Sputum-based molecular biomarkers for the early detection of lung cancer: limitations and promise. *Cancers (Basel)* 2011;3(3): 2975-2989.
25. Witt BL, Wallander ML, Layfield LJ, Hirschowitz S. Respiratory cytology in the era of molecular diagnostics: a review. *Diagn Cytopathol* 2012;40(6):556-563.
26. Roa WH, Kim JO, Razzak R, Du H, Guo L, Singh R, et al. Sputum microRNA profiling: a novel approach for the early detection of non-small cell lung cancer. *Clin Invest Med* 2012;35(5):271.
27. Lee RC, Feinbaum RL, Ambros V. The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell* 1993;75(5):843-854.
28. Wightman B, Ha I, Ruvkun G. Posttranscriptional regulation of the heterochronic gene *lin-14* by *lin-4* mediates temporal pattern formation in *C. elegans*. *Cell* 1993;75 (5):855-862.
29. Haneklaus M, Gerlic M, O'Neill LA, Masters SL. miR-223: infection, inflammation and cancer. *J Int Med* 2013;274(3):215-226.
30. Fuse M, Kojima S, Enokida H, Chiyomaru T, Yoshino H, Nohata N, et al. Tumor suppressive microRNAs (miR-222 and miR-31) regulate molecular pathways based on microRNA expression signature in prostate cancer. *J Hum Genet* 2012;57(11):691-699.
31. Hidaka H, Seki N, Yoshino H, Yamasaki T, Yamada Y, Nohata N, et al. Tumor suppressive microRNA-1285 regulates novel molecular targets: aberrant expression and functional significance in renal cell carcinoma. *Oncotarget* 2012;3(1):44-57.
32. Jin L, Wessely O, Marcusson EG, Ivan C, Calin GA, Alahari SK. Prooncogenic factors miR-23b and miR-27b are regulated by Her2/Neu, EGF, and TNF- α in breast cancer. *Cancer Res* 2013;73(9):2884-2896.
33. Chiyomaru T, Seki N, Inoguchi S, Ishihara T, Mataka H, Matsushita R, et al. Dual regulation of receptor tyrosine kinase genes EGFR and c-Met by the tumor-suppressive microRNA-23b/27b cluster in bladder cancer. *Int J Oncol* 2015;46(2):487-496.
34. Zhao G, Liu L, Zhao T, Jin S, Jiang S, Cao S, et al. Upregulation of miR-24 promotes cell proliferation by targeting NAIF1 in non-small cell lung cancer. *Tumour Biol* 2015;36(5):3693-3701.
35. Pulikkan JA, Dengler V, Peramangalam PS, Zada AAP, Müller-Tidow C, Bohlander SK, et al. Cell-cycle regulator E2F1 and microRNA-223 comprise an autoregulatory negative feedback loop in acute myeloid leukemia. *Blood* 2010;115(9):1768-1778.
36. Li X, Zhang Y, Zhang H, Liu X, Gong T, Li M, et al. miRNA-223 promotes gastric cancer invasion and metastasis by targeting tumor suppressor EPB41L3. *Mol Cancer Res* 2011;9(7):824-833.
37. Jia CY, Li HH, Zhu XC, Dong YW, Fu D, Zhao QL, et al. MiR-223 suppresses cell proliferation by targeting IGF-1R. *PLoS one* 2011;6(11):e27008.
38. Nian W, Ao X, Wu Y, Huang Y, Shao J, Wang Y, et al. miR-223 functions as a potent tumor suppressor of the lewis lung carcinoma cell line by targeting insulin-like growth factor-1 receptor and cyclin-dependent kinase 2. *Oncol Lett* 2013;6(2):359-366.

39. Yang M, Chen J, Su F, Yu B, Su F, Lin L, et al. Microvesicles secreted by macrophages shuttle invasion-potentiating microRNAs into breast cancer cells. *Mol Cancer* 2011;10:117.
40. Liu Q, Zhang M, Jiang X, Zhang Z, Dai L, Min S, et al. miR-223 suppresses differentiation of tumor-induced CD11b+ Gr1+ myeloid-derived suppressor cells from bone marrow cells. *Int J Cancer* 2011;129(11):2662-2673.
41. Li S, Li Z, Guo F, Qin X, Liu B, Lei Z, et al. miR-223 regulates migration and invasion by targeting Artemin in human esophageal carcinoma. *J Biomed Sci* 2011;18:24.
42. Sun W, Shen W, Yang S, Hu F, Li H, Zhu TH. miR-223 and miR-142 attenuate hematopoietic cell proliferation, and miR-223 positively regulates miR-142 through LMO2 isoforms and CEBP- β . *Cell Res* 2010;20(10):1158-1169.
43. Li J, Guo Y, Liang X, Sun M, Wang G, De W, et al. MicroRNA-223 functions as an oncogene in human gastric cancer by targeting FBXW7/hCdc4. *J Cancer Res Clin Oncol* 2012;138(5):763-774.
44. Kurashige J, Watanabe M, Iwatsuki M, Kinoshita K, Saito S, Hiyoshi Y, et al. Overexpression of microRNA-223 regulates the ubiquitin ligase FBXW7 in oesophageal squamous cell carcinoma. *Br J Cancer* 2012;106(1):182-188.
45. Wu L, Li H, Jia CY, Cheng W, Yu M, Peng M, et al. MicroRNA-223 regulates FOXO1 expression and cell proliferation. *FEBS Lett* 2012;586(7):1038-1043.
46. Li G, Cai M, Fu D, Chen K, Sun M, Cai Z, et al. Heat shock protein 90B1 plays an oncogenic role and is a target of microRNA-223 in human osteosarcoma. *Cell Physiol Biochem* 2012;30(6):1481-1490.
47. Lu TX, Lim EJ, Besse JA, Itskovich S, Plassard AJ, Fulkerson PC, et al. MiR-223 deficiency increases eosinophil progenitor proliferation. *J Immunol* 2013;190(4):1576-1582.
48. Wei Y, Yang J, Yi L, Wang Y, Dong Z, Liu Z, et al. MiR-223-3p targeting SEPT6 promotes the biological behavior of prostate cancer. *Sci Rep* 2014;4:7546.
49. Zardo G, Ciolfi A, Vian L, Starnes LM, Billi M, Racanicchi S, et al. Polycombs and microRNA-223 regulate human granulopoiesis by transcriptional control of target gene expression. *Blood* 2012;119(17):4034-4046.
50. Scapoli L, Palmieri A, Muzio LL, Pezzetti F, Rubini C, Girardi A, et al. MicroRNA expression profiling of oral carcinoma identifies new markers of tumor progression. *Int J Immunopathol Pharmacol* 2010;23(4):1229-1234.
51. Wong TS, Liu XB, Wong BY, Ng RW, Yuen AP, Wei WI. Mature miR-184 as potential oncogenic microRNA of squamous cell carcinoma of tongue. *Clin Cancer Res* 2008;14(9):2588-2592.
52. Wada R, Akiyama Y, Hashimoto Y, Fukamachi H, Yuasa Y. miR-212 is downregulated and suppresses methyl-CpG-binding protein MeCP2 in human gastric cancer. *Int J Cancer* 2010;127(5):1106-1114.
53. Incoronato M, Garofalo M, Urso L, Romano G, Quintavalle C, Zanca C, et al. miR-212 increases tumor necrosis factor-related apoptosis-inducing ligand sensitivity in non-small cell lung cancer by targeting the antiapoptotic protein PED. *Cancer Res* 2010;70(9):3638-3646.
54. Hatakeyama H, Cheng H, Wirth P, Counsell A, Marcrom SR, Wood CB, et al. Regulation of heparin-binding EGF-like growth factor by miR-212 and acquired cetuximab-resistance in head and neck squamous cell carcinoma. *PLoS One* 2010;5(9):e12702.
55. Li Y, Zhang D, Chen C, Ruan Z, Li Y, Huang Y. MicroRNA-212 displays tumor-promoting properties in non-small cell lung cancer cells and targets the hedgehog pathway receptor PTCH1. *Mol Biol Cell* 2012;23(8):1423-1434.
56. Ucar A, Vafaizadeh V, Jarry H, Fiedler J, Klemmt PA, Thum T, et al. miR-212 and miR-132 are required for epithelial stromal interactions necessary for mouse mammary gland development. *Nat Genet* 2010;42(12):1101-1108.
57. Mannoor K, Liao J, Jiang F. Small nucleolar RNAs in cancer. *Biochim Biophys Acta* 2012;1826(1):121-128.
58. Wang X, Zhang Y, Nilsson CL, Berven FS, Andr en PE, Carlsohn E, et al. Association of chromosome 19 to lung cancer genotypes and phenotypes. *Cancer Metastasis Rev* 2015;34(2):217-226.
59. Park NJ, Zhou H, Elashoff D, Henson BS, Kastratovic DA, Abemayor E, et al. Salivary microRNA: discovery, characterization, and clinical utility for oral cancer detection. *Clin Cancer Res* 2009;15(17):5473-5477.
60. Li J, Smyth P, Flavin R, Cahill S, Denning K, Aherne S, et al. Comparison of miRNA expression patterns using total RNA extracted from matched samples of formalin-fixed paraffin-embedded (FFPE) cells and snap frozen cells. *BMC Biotechnol* 2007;7:36.
61. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci USA* 2008;105(30):10513-10518.
62. Chen X, Ba Y, Ma L, Cai X, Yin Y, Wang K, et al. Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res* 2008;18(10):997-1006.
63. Xie Y, Todd NW, Liu Z, Zhan M, Fang H, Peng H, et al. Altered miRNA expression in sputum for diagnosis of non-small cell lung cancer. *Lung Cancer* 2010;67(2):170-176.
64. Miah S, Dudzic E, Drayton R, Zlotta A, Morgan S, Rosario D, et al. An evaluation of urinary microRNA reveals a high sensitivity for bladder cancer. *Br J Cancer* 2012;107(1):123-128.
65. Gilad S, Meiri E, Yogev Y, Benjamin S, Lebanony D, Yerushalmi N, et al. Serum microRNAs are promising novel biomarkers. *PLoS One* 2008;3(9):e3148.
66. Ulivi P, Zoli W. miRNAs as non-invasive biomarkers for lung cancer diagnosis. *Molecules* 2014;19(6):8220-8237.