Sortilin as a Novel Diagnostic and Therapeutic Biomarker in Chronic Lymphocytic Leukemia

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Abstract

Background: The overexpression of sortilin/neurotensin receptor has previously been reported in various human solid tumors but not in hematological malignancies. Here, we report the overexpression of sortilin in leukemic cells from patients with Chronic Lymphocytic Leukemia (CLL).

Methods: Flow cytometry was used to compare the expression of sortilin in CLL patients (n=52) and healthy individuals (n=26). Also, in vitro apoptosis induction was assessed in CLL Peripheral Blood Mononuclear Cell (PBMCs) following directly targeting of sortilin.

Results: The results showed a significant expression of sortilin on the surface of CLL PBMCs (range from 2.2 to 71.5%) in comparison to healthy individuals (range from 0.03 to 7.4%) (p\leq 0.0001). The optimal cut-off value of sortilin expression was determined at 7.2% with high sensitivity and specificity. Treatment of leukemic cells with anti-sortilin antibody could induce apoptosis without any effect on normal cells.

Conclusion: Apoptosis induction in CLL cells together with a significant correlation between the expression of sortilin and CD23 represent a possible functional role of sortilin in leukemogenesis of CLL cells. Therefore, sortilin might be considered as a promising novel biomarker in diagnosis, monitoring, and therapy of patients with CLL.

Keywords: Apoptosis, Biomarker, Chronic Lymphocytic Leukemia, Monoclonal antibody, Sortilin

Introduction

B cell Chronic Lymphocytic Leukemia (B-CLL) is a clonal lymphoproliferative disorder characterized by accumulation of mature-appearing neoplastic lymphocytes with co-expressing CD5, CD19, and CD23. The majority of CLL patients live for a long time without any treatment. However, CLL has still to be considered as an incurable disease. Due to low-efficient therapy and risk of side effects, it is often advised waiting until the disease become progressive and the bothersome symptoms appear. Therefore, the discovery of molecular biomarkers enabling us to predict disease progression to start therapeutic interventions would be advantageous. Indeed, improvement in our knowledge about the biology of CLL will improve the therapeutic options.

Sortilin is a 95 kDa transmembrane glycoprotein with a deregulated expression in many human cancers and also neurological disorders. Human sortilin is encoded by SORT1 gene located on chromosome 1 and classified as a member of mammalian vacuolar protein sorting 10p domain (Vps10pD) family. As a multifunctional receptor, sortilin mediates transport of proteins such as neurotensin and neurotrophin to cell membrane or lysosomes, directing cell survival and tumorigenesis.

Several studies have reported that sortilin is deregulated in various human carcinomas including breast, colon, prostate, lung, and melanoma. We have also previously reported the overexpression of sortilin in ovarian carcinoma using a developed specific mAb (clone 2D8). Here, we used this antibody to study the overexpression of sortilin in CLL patients in comparison with healthy individuals. The induction of apoptosis in CLL Peripheral Blood Mononuclear Cells (PBMCs) following 2D8 mAb treatment showed that sortilin may function as a survival factor in CLL. In this study, we attempted to evaluate sortilin/neurotensin re-
ceptor as a new biomarker in CLL and also determine the application of anti-sortilin antibody in targeting CLL leukemic cells.

**Materials and Methods**

**Patients and healthy individuals**

Blood samples were collected from untreated CLL patients (n=52) referred to Firoozgar Hospital (Tehran, Iran) and healthy individuals (n=26). Patients and healthy individuals were informed for the content of study and consented to provide blood sample for research purposes. PBMCs were isolated using Ficoll-Paque plus (GE Healthcare, Little Chalfont, UK) according to the manufacturer instructions. The study was conducted in accordance with the 1964 Helsinki Declaration and was approved in the ethical committee of Avicenna Research Institute (ARI).

**Cell lines**

Cell lines including 232-B4, I83-E95, WA-C3CD5+, Jurkat, RPMI 8226, Caov-4 and SKOV3 (National Cell Bank of Iran, Tehran, Iran) as well as Lymphoblastoid Cell Line (LCL) were cultured in RPMI 1640 media (10% FBS) (Gibco) in 24-well plates for 2 hours, and then treated with 10 μg/ml of 2D8 or isotype control mAbs (ARI, Tehran, Iran). Afterwards, FITC-conjugated sheep anti-mouse Ig (1:50) (ARI) was added. Primary antibodies were CD5-FITC/CD19-PE, CD23-PE (DAKO, Troy, MI), CD38-PE (10 μl) and ZAP-70-PE (10 μl) (BD Biosciences, San Jose, CA). To analyze the expression of ZAP-70, PBMCs were fixed and permeabilized using Bioscience kit (San Diego, CA). Fluorescence intensity was analyzed using BD Accuri™ C6 Plus flow cytometer (BD Biosciences).

**Immunofluorescent staining**

Thirty thousand cells were seeded and fixed with acetone following blocking with 5% normal sheep serum for 30 minutes. The slides were incubated with2D8 or isotype control mAbs (ARI, Tehran, Iran). The nuclei were stained with 1 μg/ml of 4′,6-Diamidino-2-phenylindole dihydrochloride (DAPI, Sigma). The slides were observed under an Olympus BX51 fluorescent microscope (Tokyo, Japan).

**Apoptosis assay**

PBMCs were purified from CLL patients (n=6) and healthy individuals (n=6) under sterile conditions. Cells (4x10⁶) were cultured in 400 μl of RPMI-1664 (5% FBS) (Gibco) in 24-well plates for 2 hr, and then treated with 10 μg/ml of 2D8 or isotype control mAbs. Staurosporine (2 μM) (Sigma) was served as positive control. The controls including isotype control mAb, and staurosporine were used simultaneously in other study. After 14 hr incubation, cells were removed from the culture medium and stained with Annexin V FITC and Propidium Iodide (PI) (BD Biosciences) followed by FACS analysis using BD Accuri™ C6 Plus flow cytometer. To calculate the percentage of apoptotic cells, the cell percentage in early (i.e. Annexin V+, PI-) and late apoptosis (i.e. Annexin V+, PI+) were added together.

**Statistical analysis**

Statistical analysis was performed using Student’s t-test for parametric data and Mann–Whitney U test for nonparametric data. Receiver operating characteristic (ROC) curve was used to determine the optimal cut-off value of sortilin expression. Analyses were performed using GraphPad Prism 6 software and statistical significance was defined as p≤0.05.

**Results**

**Sortilin expression in CLL and healthy PBMCs**

The expression of sortilin on the cell surface of purified PBMCs from 52 CLL patients (median age 59 years, range 40-81) was compared to 26 healthy individuals (median age 46.5 years, range 25-70). The biological characteristics and immunophenotyping of CLL patients are summarized in table 1.

Flow cytometry results showed that 2D8 mAb specifically detected sortilin on the surface of CLL cells, but not healthy PBMCs (Figure 1A). Results demonstrated that the median expression of sortilin in CLL patients was 24.6% (range from 2.2% to 71.5%) in comparison with healthy individuals which was 4.2% (range from 0.03% to 7.4%) (p<0.0001) (Figure 1B). However, no significant difference in sortilin expression was observed between progressive and non-progressive cases.

**Table 1. Clinical characteristics of CLL patients**

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>N=52</th>
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<tbody>
<tr>
<td>Gender</td>
<td>Male/Female 30/22</td>
</tr>
<tr>
<td>Age</td>
<td>Median 62, Range 42-85</td>
</tr>
<tr>
<td>Treatment history</td>
<td>(N) Prior treatment 0</td>
</tr>
<tr>
<td>Rai stage</td>
<td>0 (n=38), I (n=7), II (n=5), III and IV (n=0) 50</td>
</tr>
<tr>
<td>Lymphocyte count</td>
<td>(10⁹/l) Median (range) 30 (11-150)</td>
</tr>
<tr>
<td>CD23 (%)</td>
<td>Mean±SEM 58±3, Median (range) 63 (2-98)</td>
</tr>
<tr>
<td>CD5⁺/19⁺ (%)</td>
<td>Mean±SEM 77±1.8, Median (range) 78 (47-97)</td>
</tr>
<tr>
<td>ZAP-70 (%)</td>
<td>Greater than 20% 1, Lower than 20% 51</td>
</tr>
<tr>
<td>CD38 (%)</td>
<td>Greater than 30% 16, Lower than 30% 36</td>
</tr>
</tbody>
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or even between different stages (0, I and II) of the malignancy. The commercial polyclonal antibody did not detect any cell surface expression of sortilin neither in healthy nor in leukemic cells.

**Determining cut-off value**

The cut-off value of sortilin expression in CLL patients was determined at 7.2% (AUC: 0.98, sensitivity: 96.1%, specificity: 94.2%) ($p \leq 0.0001$) (Figure 2). Considering cut-off value of sortilin expression (7.2%), 3 out of 52 patients (5.7%) were identified as negative, and only one out of 26 healthy individuals (3.8%) was presented as false positive.

**Immunofluorescent staining**

The expression of sortilin in CLL patients was also investigated by immunofluorescent staining. Figure 3 shows that sortilin is expressed in leukemic cells but not in healthy PBMC. MAb 2D8 also detected sortilin in CLL cell lines including 232-B4, WA-C3CD5+ and I83-E95 (Figure 3). Caov-4 ovarian carcinoma cell line was used as positive control.

**Western blot**

Cells were lysed in a lysis buffer (1% Triton X-100, 50 mM Tris, pH=7.4, 150 mM NaCl, 5 mM EDTA, 1 mM NaF, 20 mM Na3P2O7, 1% glycerol, 0.1% sodium deoxycholate) containing 10% phosphatase inhibitor and...
sequent electro-transferring to PVDF membrane. The yacrylamide gel electrophoresis (SDS-PAGE) with sub-
loaded to each well of 10% Sodium dodecyl sulfate pol-
thiothreitol (5 mM) to samples, 15 µg of cell lysate were
mammals were probed with primary antibodies in-
were added. The membranes were developed using an
anti-mouse Ig (1:2500) or anti-rabbit Ig (1:3500) (ARI)
According to the manufacturer's instruc-
To the best of our knowledge, this is the first time
reporting cell surface expression of sortilin in malignant
result shows an ectopic expression of sortilin by local-
izing the protein on the surface of malignant B cells
1% protease inhibitor cocktail. Following addition of di-
MCL and MZL; however, no statistically difference
Conclusion: 5.6%) (p=0.01) (Figure 5B). Isotype control mAb
results shows an ectopic expression of sortilin by local-
ating cancer.
Here we report overexpression of sortilin on the
expression in leukemia cells (r=0.27, p=0.045) (Figure 4). However, no correlation between sortilin and other CLL immunophenotypic markers in
including CD38 and ZAP70 was identified (data not
Apoptosis induction following incubation with 2D8
mAb was compared between CLL (n=6) and healthy
PBMCs. Therefore, sortilin might be categorized as a di-
agnostic biomarker in CLL patients. As there was no
significant difference in sortilin expression between two
progressive and non-progressive cases, it may
not be classified as a prognostic indicator of CLL. This
might imply that sortilin expression is a genuine phe-
omenon of malignant B cells initiated since the for-
mation of leukemic cells.
Expression of sortilin was studied in different stages
(0, I and II) of CLL; however, no statistically difference
was found. Perhaps more comprehensive studies could
assist in defining the sortilin as an adequate staging
marker in CLL patients. We found a significant relation-
ship between expression of sortilin and CD23 as an im-
munophenotypic marker of CLL. Since CD23 contrib-
utes to accumulation of long-lived B CLL cells, it may
conclude that sortilin can play a role in B cell survival
akin to CD23. CD23 is a cell surface protein considered
to be important in differentiation of CLL from Mantle
Cell Lymphoma (MCL) and Marginal Zone Lymphoma
(MZL) 21. Therefore, it is wise to investigate the lack/
presence of sortilin in patients with MCL and MZL;
which may add another tumor marker to the current di-
gnostic panel for differentiating CLL from other Non-
Hodgkin’s lymphomas.
Although it is not obvious why CLL cells harbor cell
surface sortilin, several studies suggest that sortilin may
also undertake vital tasks in the pathophysiology of
other cancers. Recent finding suggests a role of sortilin
in angiogenesis through releasing of exosomes to the ex-
cellular media 22. A complex composed of sortilin and
results shows an ectopic expression of sortilin by local-
izing the protein on the surface of malignant B cells
which differs from its subcellular localization either in-
tracellular or secreted forms in normal B cells 19. The
Western blot data using 2D8 mAb (Figure 6) clearly
shows expression of a 95 kDa band in CLL lysates but
not in normal healthy B cells (left panel) confirming our
immunocytochemistry data in normal B cells (Figure 3).
Additionally, our antibody detects a strong band of 40
kDa in CLL cells with a faint band in normal B cells.
Furthermore, using a commercially available anti-
sortilin antibody in Western blot experiments a 60 kDa
band of sortilin variant (in addition to the 95 kDa band)
was observed in CLL cells but not in normal healthy in-
dividuals (Figure 6, right panel). This may suggest that
the structure of sortilin protein expressed in malignant
B cells differs from its normal counterpart. Such differ-
ence has been reported for ectopic expression of a 38
kDa proline/arginine rich end leucine-rich repeat protein
(PRELP) in leukemic B cells from CLL patients 19. It
seems that variation in the structure of ectopically ex-
pressed proteins may affect the subcellular localization,
representing a hallmark of cancer cells. This notion
should be considered in discovery of new markers tar-
geting cancer.

Correlation between sortilin and CD23
A significant correlation was observed between sort-
ilin and CD23expression in leukemic cells (r=0.27,
p=0.045) (Figure 4). However, no correlation
between sortilin and other CLL immunophenotypic markers in
cluding CD38 and ZAP70 was identified (data not
shown).

Apoptosis assay
Apoptosis induction following incubation with 2D8
mAb was compared between CLL (n=6) and healthy
PBMCs (Figure 5). Figure 5A is the representative
of one out of six individuals in each group of CLL pa-
tients and healthy controls. Results demonstrated
that 2D8 mAb induced apoptosis in CLL cells (median:
31.5%) with very low effect on healthy individuals (me-
dian: 5.6%) (p≤0.01) (Figure 5B). Isotype control mAb
did not induce significant apoptosis in CLL (6.5%) or
healthy individuals (5.1%). Stauroporine, as positive
control, induced apoptosis in both CLL (52.5%) and
healthy PBMCs (60%) (p=0.05).

Discussion
To the best of our knowledge, this is the first time
reporting cell surface expression of sortilin in malignant
B cells of CLL patients. We used three different protein
readout systems to verify our findings. Flow cytometry

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Expression of Sortilin in CLL

two tyrosine kinase receptors containing TrkB and EGFR in exosome structure mediated communication and signaling events between the lung cancer cells and its target, the endothelial cells 22. The immunohistochemistry experiments performed in a cohort of breast cancers patients showed an increase in the expression of sortilin which was associated with the breast cancer aggressiveness and invasion 5. In other study, it was shown that activation of matrix metalloproteinases in HT29 colorectal cancer cell line led to cleavage of the luminal part of sortilin resulting in release of its soluble form (sSortilin) 23. Subsequently, activation of Focal Adhesion Kinase pathway by sSortilin regulates numerous downstream intracellular pathways mediating either cell migration or metastasis 23. Sortilin as a neurotrophin transporter is implicated in cell proliferation and anti-apoptotic effect of Brain-Derived Neurotrophic Factor (BDNF) in colorectal cancer cell 3. The phenomenon is accelerated by high affinity binding of sortilin to pro-BDNF as an apoptosis inducer in this type of malignancy. Knocking down of sortilin using siRNA treatment showed that sortilin plays a survival role in Caov-4 ovarian carcinoma cell line 13. In the same way, sortilin silencing resulted inhibition of cell survival and migration through decreased activation of extracellular signal-regulated kinases (ERK) in CAL-62 thyroid cancer cell line 24.

To investigate the potency of sortilin as a therapeutic biomarker in CLL, we explored the apoptosis induction by targeting sortilin using 2D8 mAb. As shown in figure 5A, treatment with 2D8 mAb considerably led to apoptosis in CLL cells but not in healthy PBMCs (p≤0.01). It is not exactly clear how 2D8 mAb induces cell de-
struction; however, it is obvious that this antibody directly induces apoptosis in CLL PBMCs. Mechanisms like antigen crosslinking, blockade of ligand-receptor growth or survival pathways, or activation of death receptors potentially may elicit the induction of direct apoptosis in targeted cells 25. Previously, it was shown that sortilin forms a functional complex by heterodimerization with Neurotensin Receptor 1 (NT-R1), mediating internalization of neurotensin growth signaling pathways 26,27. Therefore, it is also possible that 2D8 mAb may induce direct apoptosis in CLL cells by blockade of this type of heterodimerization. Therefore, it is necessary to elucidate the expression of NTR1, its heterodimerization with NTR3, and also endogenous expression of neurotensin in CLL cells to firmly state this hypothesis. In overall, beyond the mechanism of action, mAbs can induce targeted cell killing alone or can enhance target cell susceptibility to chemo- or radio-therapeutics by affecting the modulation of anti-apoptotic pathways 25.

Here we suggest a combination of chemo- and immunotherapy regimens containing specific mAbs targeting sortilin which might increase the efficacy of responses in CLL. Akin to the improved response achieved from combination of anti-CD20 antibody (of atumumab) plus fludarabine and cyclophosphamide in patients with relapsed CLL 28. Since CLL is a complex malignancy with a very heterogeneous pattern, and many signaling pathways may act in parallel in cancer cells; a combination of multi-targeting antibody-based therapy is necessary 29.

Based on a report on expression profile of sortilin in normal resting B cells and few malignant B cell lines, no surface expression of sortilin was detected either in normal or malignant B cells 18. Although sortilin is expressed in normal B cells but its localization differs in normal and malignant conditions. Our results clearly show that our anti-sortilin antibody detects sortilin on the surface of malignant B cells. Such discrepancy in findings might be due to the nature of immunogen used for antibody generation, as our antibody generated against the very N-terminal part of sortilin encompassing signal peptide 12.

**Conclusion**

In conclusion, our results from apoptosis induction in CLL cells by targeting sortilin might be attributed to the significant role of sortilin in CLL survival. Therefore, we believe that this is an important finding, which may potentially contribute to diagnosis, monitoring, and therapy of patients with CLL.

**Acknowledgement**

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**Conflict of Interest**

The authors declare no conflict of interest.

**References**


