

## Sericin Ameliorates the Capacitation State and Chromatin Integrity of Frozen-Thawed Stallion Spermatozoa by Reducing Oxidative Stress

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### Abstract

**Background:** In the process of sperm cryopreservation, apart from cryoinjury, the production of Reactive Oxygen Species (ROS) can adversely affect the integrity of chromatin and cellular membranes. Addition of natural antioxidants to freezing medium is an approach to reduce the destructive effects of ROS on sperm.

**Methods:** In this study, during 60 min of cooling process, the ejaculates of five stallions were diluted in the following media: INRA 82 medium as Control (C), INRA 82 medium supplemented with 0.25% Sericin (S), INRA 82 medium supplemented with 1.5 mM Glutathione (G), and INRA 82 medium supplemented with 0.25% Sericin+1.5 mM Glutathione (S+G).

**Results:** In the frozen/thawed sericin supplemented group, while the integrity of DNA and the activity of catalase and Glutathione Peroxidase (GPx) were increased, the lipid peroxidation and midpiece abnormality decreased, compared with other groups ( $p < 0.05$ ). The proportions of sperms with abnormal head in group S and the sperm with distal droplet in G and S+G groups decreased, compared with group C ( $p < 0.05$ ). In CTC assay, the percentage of capacitated spermatozoa in treatment groups was lower than control ( $p < 0.01$ ).

**Conclusion:** In conclusion, the presence of sericin in freezing medium of stallion semen could improve sperm DNA integrity and its resistance to ROS and lipid peroxidation.

*Avicenna J Med Biotech 2019; 11(3): 245-252*

**Keywords:** Antioxidants, Glutathione, Sericins, Spermatozoa

### Introduction

Despite the successes achieved in semen cryopreservation in the cattle industry, there are some limitations in generalization of this technology in the equine breeding industry <sup>1</sup>. One of the most important of these limitations is the destructive effect of freeze/thaw process on sperm quality arising from physical and chemical stresses on the sperm plasma membrane. Part of these destructive effects is associated with oxidative stress due to production of Reactive Oxygen Species (ROS) <sup>2</sup>. In equine, Polyunsaturated Fatty Acids (PUFAs) of sperm plasma membrane and its DNA structure are specifically vulnerable to oxidative stress <sup>3</sup>.

Sperm is protected against ROS and Lipid Peroxidation (LPO) through two principal strategies, using enzymatic and non-enzymatic antioxidants. Glutathione Peroxidase (GPx), Superoxide Dismutase (SOD), and Catalase (CAT) are the most critical enzymatic antioxidants <sup>4,5</sup> and reduced glutathione (GSH),  $\alpha$ -tocopherol, ascorbic acid, carotenoids, ubiquinones, taurine, and

hypotaurineserve as non-enzymatic antioxidants <sup>6,7</sup>. Any disproportion between the antioxidant systems and ROS contents due to seminal plasma dilution during semen processing and/or during freeze-thaw process leads to a significant reduction in the defense system against free radicals and increases the destructive effects of LPO <sup>3</sup>. To overcome this problem, various antioxidants were added to semen extenders of different species. A considerable body of evidence indicates that extender supplementation with antioxidants before cryopreservation has improved the post-thaw motility, membrane integrity, and fertilizing ability of sperm infeline <sup>8</sup>, canine <sup>9</sup>, ovine <sup>10</sup>, caprine <sup>11</sup>, bovine <sup>12</sup>, buffalo <sup>13</sup> and equine <sup>14</sup> species.

Glutathione (L- $\gamma$ -glutamyl-L-cysteinylglycine, GSH) is a tripeptide ubiquitously found in mammalian spermatozoa and seminal plasma <sup>15</sup> which with thiol (SH) groups has antioxidant characteristics. Therefore, it has an important role to directly react with ROS and work

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Received: 13 Jun 2018  
Accepted: 9 Aug 2018

as a co-factor for GPx to reduce both hydrogen peroxide to H<sub>2</sub>O and lipoperoxides to alkyl alcohols<sup>16</sup>. Based on evidence, the glutathione concentration of semen significantly depletes throughout storage period<sup>17,18</sup>. Additionally, it has been demonstrated that supplementation of extender with low dosage of glutathione improves the spermatozoa quality in some species<sup>17,19,20</sup>.

Sericin as a water soluble component of silk which contributes to 20-30% of the total cocoon weight, has an antibacterial and UV resistant properties<sup>21,22</sup>. Its molecular weight ranges from 10 to 310 kDa, containing 18 kinds of amino acids<sup>23</sup> with high content of the hydroxyl groups of hydroxyamino acids (Serine and threonine) which can suppress *in vitro* lipid peroxidation and tyrosinase activity<sup>24</sup>. It is indicated that culture medium supplementation with 0.5% sericin improves the developmental competence of bovine embryo by protecting against oxidative stress<sup>25</sup>. It has been also demonstrated that the addition of sericin to freezing medium of various mammalian cells is comparable to the conventional medium containing serum<sup>26</sup>. Sericin as a cell membrane-impermeable cryoprotectant with inhibitory effect against ice crystal formation may protect the cells from cryodamage<sup>27</sup>. However, no information is available related to the addition of sericin to the freezing medium of stallion semen.

In the present study, the purpose was to evaluate the effects of sericin supplementation to semen extender with or without GSH on the frozen/thawed stallion spermatozoa. Therefore, motility parameters, morphology, lipid peroxidation, GPx, SOD, catalase activities, capacitation, DNA and plasma membrane integrity of spermatozoa were also assessed in frozen-thawed stallion semen.

## Materials and Methods

### Material

All chemicals and reagents were obtained from Sigma (St. Louis, MO) and Gibco (Grand Island, NY, USA), respectively, unless otherwise stated in the text.

### Semen collection and processing

Semen was collected (Three times each) from five fertile stallions, 8 to 10 years old, using an artificial vagina (Missouri model, IMV, France). After filtration, the gel-free fraction of the ejaculates was immediately transported to the laboratory for evaluation of spermatozoa quality. Only ejaculates with good wave motion (>60%) were considered for the experiments<sup>28</sup>. The samples after dilution in INRA 82 medium (0.5 L saline solution: 25 g glucose, 1.5 g lactose, 1.5 g raffinose, 0.25 g sodium citrate dihyd rate, 0.41 g potassium citrate, 50000 IU penicillin, 50 mg gentamicin, and 0.5 L skim milk) containing 2% centrifuged egg yolk (v/v) and 20 mM HEPES<sup>29</sup>, were centrifuged at 600 g for 5 min. The resulting pellets were re-suspended in freezing medium (INRA 82 medium) containing 2%

(v/v) egg yolk, 20-mM HEPES, and 2.5%, glycerol (v/v; pH=6.8) and were then considered for experimental groups.

### Study design

The diluted semen with freezing medium (ratio 1:1) were randomly divided into the following groups: C) without any supplementation, served as control, S) supplemented with 0.25% sericin, G) supplemented with 1.5 mM Glutathione, and S+G) supplemented with 0.25% sericin+1.5 mM Glutathione. Fifteen ejaculates (three ejaculates from each stallion) after freeze-thaw process were considered for evaluation of the total and progressive motility, lipid peroxidation, DNA and plasma membrane integrity, and enzymatic antioxidant activity of superoxide dismutase, catalase, and glutathione peroxidase.

### Freezing and thawing

Semen freezing was carried out according to Vidament *et al*<sup>30</sup>. In brief, the loaded 0.5 ml plastic straws with diluted sperm with a final concentration of  $2 \times 10^8$  were slowly cooled to 4 °C within 60 min. The straws were horizontally placed in racks 4 cm above the surface of Liquid Nitrogen (LN<sub>2</sub>) for 12 min, and then directly plunged in LN<sub>2</sub>. Two weeks later, the straws were then thawed in a water bath at 37°C for 40 s.

### Post thaw sperm analysis

The motility characteristics of sperm including Amplitude of Lateral Head displacement (ALH), Beat Cross Frequency (BCF), curvilinear velocity (VCL), straight line velocity (VSL), average path velocity (VAP), linearity (LIN=VSL/VCL×100), Mean Angular Displacement (MAD), wobble (WOB=VAP/VCL×100), and straightness (STR=VSL/VAP×100) were evaluated after thawing using computer assisted semen analyzers (CASA, Houshm and Fanavar, Iran). The spermatozoa were classified into four classes: class A, fast progressively motile spermatozoa; class B, weak progressively motile spermatozoa; class C, non-progressively motile spermatozoa; and class D, immotile spermatozoa. Both classes of A and B were considered as the percentage of progressive motile spermatozoa.

### Biochemical assays

The frozen-thawed sperm after homogenizing with ultrasonic homogenizer were subjected to the total protein content assessment using the Bio-Rad protein assay kit (Bio-Rad, Hercules, CA) according to the manufacturer instructions.

Lipid peroxidation was determined by measuring the amount of malondialdehyde (MDA) in the samples using the method of Buege and Aust<sup>31</sup>. Briefly, the thawed and diluted samples with distilled water, were treated with 2 ml of TBA-TCA-HCl reagent (Thiobarbituric acid 0.37%, trichloroacetic acid 15% and 0.25 N HCl) and after coverage with foil were then boiled for 15 min in water bath. The samples were then cooled and centrifuged at 3000 g for 10 min at room temperature. The absorbance of supernatant was measured with

a spectrophotometer against reference blank at 532 nm.

SOD activity in spermatozoa was assayed according to the method of Kakkar *et al*<sup>32</sup>. In brief, 50  $\mu$ l spermatozoa were added to 450  $\mu$ l distilled water, and treated with 2.45 ml reaction mixture (Xanthine, Nitroblue tetrazolium; NBT, and phosphate buffer). Subsequently, 50  $\mu$ l of xanthine oxidase (0.1 mg/ml) was added to the mixture, and the reaction was terminated by adding 50  $\mu$ l copper dichloride (6 mM) every 30 s up to 5 min. The absorbance was recorded at 560 nm and the results were reported as the inhibition rate of NBT reduction.

GPx activity was measured by the method described by Lawrence and Burk<sup>33</sup>. Briefly, 20  $\mu$ l of spermatozoa were added to 980  $\mu$ l of the reaction mixture (1 ml phosphate buffer, 100  $\mu$ l 20 mM sodium azide, 100  $\mu$ l 3 mM NADPH, 100  $\mu$ l 10 mM EDTA, 100  $\mu$ l 20 mM reduced glutathione, and 200  $\mu$ l 2 U glutathione reductase) and incubated at 37°C for 5 min. Ten  $\mu$ l of hydrogen peroxide was then added to the above mixture. The change in absorbance at 340 nm was monitored against a blank every 30 s up to 2 min. GPx activity unit is defined as  $\mu$ mol of NADPH consumed per min per mg protein.

The catalase (CAT) activity was assayed as described by Goth<sup>34</sup>. In brief, 40  $\mu$ l spermatozoa after dilution in 160  $\mu$ l of potassium phosphate buffer were incubated in 1 ml reaction mixture (65 mM hydrogen peroxide in 60 mM potassium phosphate buffer, pH=7.4) at 37°C for 1 min. The reaction was stopped by the addition of 1 ml 32.4 mM ammonium molybdate solution. The absorbance was measured at 405 nm against a blank (1 ml reaction mixture, 1 ml ammonium molybdate solution and 200  $\mu$ l of potassium phosphate buffer). One unit of CAT was defined as the amount of enzyme catalyzing the degradation of 1  $\mu$ mol hydrogen peroxide per min.

#### Chromatin structure assay

Two smears from each ejaculate were prepared on glass slides and air-dried. The smears after fixation in Carnoy's solution (3:1 methanol:glacial acetic acid) for 2 hr were rinsed with distilled water and then stained with 1% Acridine Orange (AO) for 10 min and after washing, covered with a coverslip and immediately sealed with nail polish. All slides were evaluated with an epifluorescent microscope (Olympus BX51, Japan) at 400 $\times$  magnification. Spermatozoa with normal chromatin structure were stained as green heads, while spermatozoa with abnormal DNA (Single-strand) were colored from yellow to red (Figure 1). A minimum of 200 spermatozoa per slide were evaluated and percentage of spermatozoa with intact chromatin was calculated for each sample.

#### Plasma membrane integrity

The Hypo-Osmotic Swelling Test (HOST) was used to evaluate spermatozoa membrane integrity after freeze-thaw process. Briefly, 30  $\mu$ l of each semen sample was added to 200 ml of hypo-osmotic solution (25

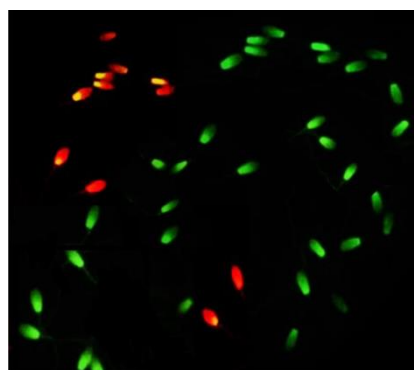


Figure 1. Stallion sperm stained with 1% Acridine Orange (AO). Sperm with normal chromatin structure display green fluorescence and those with an abnormal DNA (Single-strand) shows a yellow to red fluorescence color.

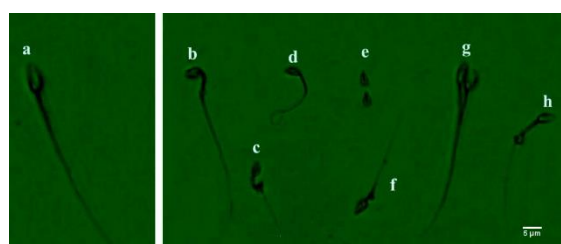


Figure 2. Sperm morphology using Diff-Quick. Sperm with normal. a) and abnormal morphologies (b-h). b) bent midpiece; c) abnormal midpiece; d) bent tail; e) tailless head; f) proximal droplet; g) abnormal head; h) distal droplet.

mM sodium citrate dihydrate and 75 mM fructose) and incubated at 37°C for 30 min. Then, the smear was prepared on glass slides and covered with a coverslip. At least 200 spermatozoa were counted under phase-contrast microscope at 400 $\times$  magnification, and percentage of spermatozoa with coiled tail (Intact membrane) was recorded for each sample.

#### Sperm morphology

To evaluate the spermatozoa morphology, samples were stained with Diff-Quick staining. Briefly, from each semen sample, duplicate smears were prepared on glass slides and after fixation, the smears were stained with staining solutions according to the instruction of Diff-Quick staining kit (Avicenna, IRI). The morphology of at least 200 spermatozoa per slide was evaluated under a bright field microscope at 100 $\times$  magnification and the percentage of normal and abnormal sperm was recorded in each treatment group (Figure 2).

#### Capacitation state of spermatozoa

The Chlortetracycline (CTC) assay was used to assess the capacitation state of spermatozoa. Briefly, 20  $\mu$ l of CTC solution was laid on a glass slide and thoroughly mixed with 20  $\mu$ l of each semen sample and after fifteen s, 5  $\mu$ l of fixative solution was added to the suspension. Afterward, 3  $\mu$ l of antifade solution was added and the slide covered with a coverslip and sealed with nail polish. The slides were stored at 4°C in dark and examined at 100 $\times$  magnification within 2 hr of

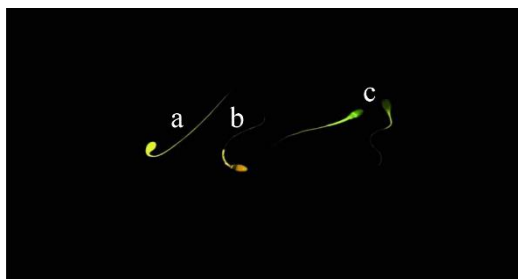


Figure 3. Capacitation state of stallion spermatozoa using chlortetracycline (CTC) staining. a) uncapacitated acrosome-intact spermatozoa (F-pattern); b) capacitated acrosome-intact spermatozoa (B-pattern); c) acrosome reacted spermatozoa (AR-pattern).

preparation under epifluorescence microscope (Olympus BX51, Japan). Two hundred sperm with duplicate from each sample were assessed according to those described by Schembri *et al*<sup>35</sup>. The patterns of staining were considered as follows: F-pattern; green fluorescence over the entire spermatozoa head, uncapacitated sperm with intact acrosome, B-pattern; fluorescence-free band on the post-acrosomal area, capacitated sperm with intact acrosome, and AR-pattern; non-fluorescence over the entire spermatozoa head or except for a thin line of fluorescence on the equatorial area, acrosome-reacted spermatozoa (Figure 3). Spermatozoa that did not fit into any of these described patterns, were not considered for analysis.

#### Statistical analysis

Normality and homogeneity of variances were confirmed using Kolmogorov–Smirnov and Levene’s tests, respectively. Statistical analyses were conducted with SPSS 20.0 (SPSS, Chicago, IL, USA). Data were analyzed by one-way ANOVA followed by LSD post hoc test, and differences were considered significant at  $p < 0.05$ . Results were presented as mean $\pm$ SEM.

## Results

### Spermatozoa parameters

As shown in table 1, supplementation of freezing medium with antioxidants had no significant effect on the spermatozoa motility parameters (total motility, progressive motility, VCL, VSL, VAP, MAD, ALH, BCF, LIN, WOB and STR). Among all classes of spermatozoa, only class B (weak progressive motile spermatozoa) was higher in Sericin (S) compared to Glutathione (G) group ( $p < 0.05$ ).

### Biochemical parameters

The effect of freezing medium supplementation with antioxidants on lipid peroxidation and enzymatic antioxidant activities of thawed stallion semen are shown in table 2. As shown, the level of lipid peroxidation in sericin supplemented group was significantly lower than other treatment groups ( $p < 0.01$ ), though, sericin in combination with glutathione had no such effect. Similarly, CAT and GPx activities of frozen-thawed semen were significantly higher when the extender medium was supplemented only with sericin compared to other groups ( $p < 0.01$ ). Nevertheless, extender supplementation with sericin or other antioxidants could not improve the percentage of SOD inhibition compared to control.

### Chromatin structure and plasma membrane integrity

As shown in figure 4, the percentage of spermatozoa with normal DNA structure was significantly higher in sericin supplemented group compared with other groups ( $p < 0.05$ ). No significant difference was observed among groups regarding plasma membrane integrity detected by hypo-osmotic swelling test ( $p < 0.05$ ).

### Morphology of spermatozoa

As shown in table 3, the abnormality of sperm heads

Table 1. The effect of supplementation of semen extender with antioxidants on sperm motility parameters in frozen–thawed stallion sperm

Parameters	Control	Sericin	Glutathione	Sericin+Glutathione
Motile spermatozoa (%)	44.74 $\pm$ 5.49	49.89 $\pm$ 4.06	44.10 $\pm$ 5.50	49.30 $\pm$ 6.70
Progressive spermatozoa (%)	32.53 $\pm$ 7.20	38.57 $\pm$ 5.50	30.92 $\pm$ 7.00	37.48 $\pm$ 9.10
Class A (%)	19.90 $\pm$ 5.70	20.28 $\pm$ 6.20	18.43 $\pm$ 7.02	23.32 $\pm$ 8.50
Class B (%)	12.64 $\pm$ 1.65 <sup>ab</sup>	18.29 $\pm$ 2.30 <sup>a</sup>	12.50 $\pm$ 0.59 <sup>b</sup>	13.76 $\pm$ 1.99 <sup>ab</sup>
Class C (%)	12.20 $\pm$ 2.00	11.31 $\pm$ 2.27	13.18 $\pm$ 2.00	11.83 $\pm$ 2.50
Class D (%)	55.27 $\pm$ 5.40	50.11 $\pm$ 4.06	55.90 $\pm$ 5.59	50.70 $\pm$ 6.70
VCL ( $\mu$ m/s)	34.97 $\pm$ 5.57	36.34 $\pm$ 5.15	34.87 $\pm$ 5.70	37.91 $\pm$ 7.49
VSL ( $\mu$ m/s)	17.48 $\pm$ 3.98	19.76 $\pm$ 4.42	16.73 $\pm$ 5.21	22.19 $\pm$ 7.01
VAP ( $\mu$ m/s)	21.77 $\pm$ 4.22	24.07 $\pm$ 4.54	20.98 $\pm$ 5.37	26.5 $\pm$ 7.16
MAD (°)	10.56 $\pm$ 3.99	8.94 $\pm$ 2.51	8.74 $\pm$ 2.35	8.33 $\pm$ 2.55
ALH ( $\mu$ m)	1.94 $\pm$ 0.16	2.05 $\pm$ 0.12	1.95 $\pm$ .19	1.97 $\pm$ 0.20
BCF (Hz)	0.31 $\pm$ 0.12	0.29 $\pm$ 0.10	0.27 $\pm$ 0.11	0.32 $\pm$ 0.13
LIN (%)	32.91 $\pm$ 4.38	39.73 $\pm$ 4.05	31.44 $\pm$ 6.14	38.08 $\pm$ 7.80
WOB (%)	45.78 $\pm$ 4.15	52.97 $\pm$ 3.57	44.69 $\pm$ 5.98	50.56 $\pm$ 7.16
STR (%)	48.90 $\pm$ 5.19	55.10 $\pm$ 3.86	47.96 $\pm$ 6.32	51.92 $\pm$ 7.62

VCL: Curvilinear velocity, VSL: Straight line velocity, VAP: Average path velocity, MAD: Mean angular displacement, ALH: Amplitude of lateral head displacement, BCF: Beat cross frequency, LIN: Linearity index (LIN=VSL/VCL $\times$ 100), WOB: Wobble (WOB=VAP/VCL $\times$ 100), and STR: Straightness index (STR=VSL/VAP $\times$ 100). a, b) Different superscripts within the same row demonstrate significant difference ( $p < 0.05$ ).

Table 2. The effect of supplementation of semen extender with antioxidants on lipid peroxidation and enzymatic antioxidant activity of frozen–thawed stallion semen

Group	CAT (U/mg pr)	GPx (U/mg pr)	MDA (nmol/mg pr)	SOD (% inhibition)
Control	43.95±1.60 <sup>b</sup>	64.97±2.50 <sup>b</sup>	29.43±1.24 <sup>a</sup>	14.49±0.42
Sericin	55.22±1.65 <sup>a</sup>	75.36±2.19 <sup>a</sup>	23.49±0.96 <sup>b</sup>	14.53±0.62
Glutathione	46.64±1.21 <sup>b</sup>	62.28±2 <sup>b</sup>	30.51±1.12 <sup>a</sup>	14.58±0.49
Sericin+Glutathione	47.75±2.35 <sup>b</sup>	63.04±3.4 <sup>b</sup>	29.03±0.83 <sup>a</sup>	14.22±0.26

CAT: Catalase, GPx: Glutathione peroxidase, MDA: Malondialdehyde, and SOD: Superoxide dismutase. a, b: Different superscripts within the same column demonstrate significant difference (p<0.01).

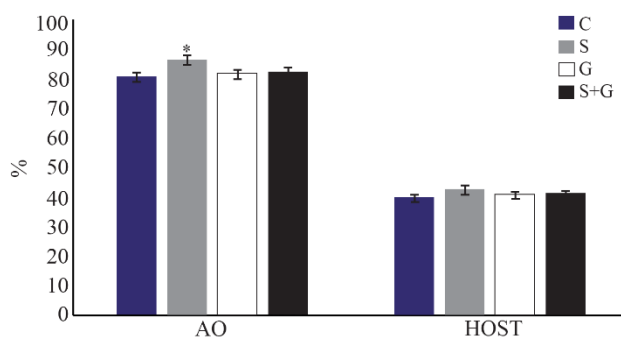


Figure 4. Effect of sericin and glutathione supplementation to the freezing medium of stallion semen on sperm chromatin structure (AO) and its plasma membrane integrity (HOST). AO: acridine orange, HOST: hypo-osmotic swelling test, C: control, S: sericin, G: glutathione, S+G: sericin+ glutathione. Values presented as %±SEM. \* p<0.05, compared to other groups.

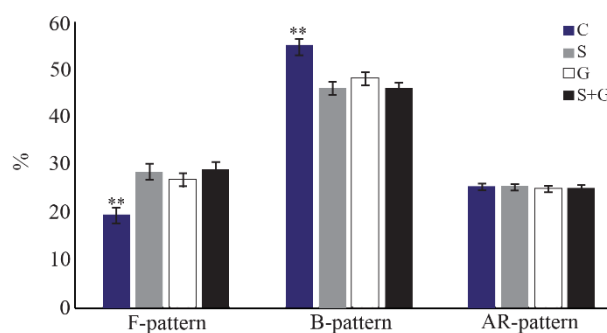


Figure 5. Effect of sericin and glutathione supplementation to the freezing medium of stallion semen on capacitation state of spermatozoa. C: control, S: sericin, G: glutathione, S+G: sericin+ glutathione. Values presented as %±SEM. \*\* p<0.01, compared to other groups.

was lower in the sericin supplemented group (5.20±0.13) compared to the control (6.20±0.48; p<0.05). The midpieces abnormality was also lower in the S group (1.53±0.08) compared to the other groups (p<0.05). The distal droplet in the G (6.40±0.40) and S+G (6.53±0.22) groups was lower than the C group (7.40±0.22; p<0.05). There were no significant differences among groups in terms of other abnormalities (p>0.05).

**Capacitation state of spermatozoa**

As shown in figure 5, the percentage of uncapacitated spermatozoa (F-pattern) in frozen–thawed semen in antioxidants supplemented groups was greater than control (p<0.01). On the other hand, the percentage of capacitated spermatozoa (B-pattern) in control was higher than treatment groups (p<0.01). There was no difference among groups in terms of acrosome-reacted spermatozoa (AR-pattern; p>0.05).

**Discussion**

The process of cryopreservation with production of oxidants and free radicals can damage the sperm. In the present study, adding sericin as a natural antioxidant, to INRA82 extender of stallion semen, alone or in combination with glutathione, slightly improved some of the sperm quality parameters. Kumar *et al* evaluated the effect of adding different concentrations of sericin to freezing medium of buffalo semen and concluded that 0.25, 0.5, and 1% of sericin ameliorated the total and progressive spermatozoa motility<sup>36</sup>. There are conflicting reports regarding the effects of glutathione on the sperm motility parameters in different species, as some of them have shown positive effects<sup>12,18,34,35</sup> while others have indicated negative effects<sup>17,19,36,37</sup>. This difference among the studies may be related to the kind of semen extender, dosage of GSH and/or inter-species differences. A dose-response study perhaps may be helpful to determine the optimal concentration of GSH.

Table 3. Sperm morphology of frozen–thawed stallion semen in the presence of antioxidants

Parameters (%)	Control	Sericin	Glutathione	Sericin+Glutathione
Normal	61.87±0.24	61.87±0.29	62.13±0.13	61.67±0.38
Abnormal heads	6.20±0.48 <sup>a</sup>	5.20±0.13 <sup>b</sup>	5.40±0.24 <sup>ab</sup>	5.67±0.23 <sup>ab</sup>
Abnormal midpieces	1.93±0.12 <sup>a</sup>	1.53±0.08 <sup>b</sup>	2.20±0.13 <sup>a</sup>	2.20±0.08 <sup>a</sup>
Bent midpieces	6.47±0.27	6.67±0.38	6.47±0.30	6.27±0.19
Proximal droplet	7.73±0.06	8.47±0.30	7.60±0.41	8.00±0.31
Distal droplet	7.40±0.22 <sup>a</sup>	6.80±0.22 <sup>ab</sup>	6.40±0.40 <sup>b</sup>	6.53±0.22 <sup>b</sup>
Bent tails	6.80±0.38	7.53±0.22	7.47±0.34	7.73±0.26
Tailless heads	1.60±0.06	1.93±0.43	2.33±0.33	1.93±0.19

a, b: Different superscripts within the same row demonstrate significant differences (p<0.05).

The result of biochemical assay showed that enzymatic antioxidant system (Except for the SOD activity) was improved in the frozen-thawed stallion semen when the extender was supplemented with sericin alone. Moreover, MDA concentration was significantly decreased by adding sericin when compared to the control and other treatment groups. This finding was in agreement with previous study where adding 0.25 and 0.5% sericin to the frozen-thawed buffalo semen had positive effects on GPx activity and LPO amelioration<sup>36</sup>.

The GPx and catalase enzymes play an important role in hydrogen peroxide removal and enhancement of GPx and catalase activities, which in turn leads to an increase in antioxidant capacity of the sperm<sup>37</sup>. In a study on the skin fibroblast cell line (AH927) treated with hydrogen peroxide, it has been shown that sericin reduces the lactate dehydrogenase and malondialdehyde activities. Similarly, in our study, sericin as a natural antioxidant, could reduce LPO and protect the spermatozoa against oxidative stress. Although the antioxidant properties of sericin have been confirmed by several studies<sup>38,39</sup>, its scavenging mechanisms have not been clearly identified. One possible mechanism may be related to the chelating effect of hydroxyl group of serine and threonine that are dominant amino acids (about 40%) in the sericin protein. However, due to the low concentration of these amino acids with thiol group, it is unlikely that this is the reason for sericin antioxidant activity<sup>24</sup>.

In the present study, plasma membrane integrity (HOST) was not affected by sericin treatment. It has been demonstrated that SOD, as an enzymatic antioxidant, plays a positive role in the spermatozoa membrane integrity following cryopreservation process. On the other hand, due to physical damage to sperm plasma membranes in the cryopreservation process and SOD leakage<sup>40</sup>, the protective effect of SOD on plasma membrane is reduced. Since in our study the sericin had no positive effect on SOD activity, no improvement was observed in sperm membrane integrity.

In the present study, the addition of GSH alone or in combination with sericin could not protect the stallion spermatozoa from free radicals and LPO which was in agreement with Zhandi and Ghadimi study who reported 2 hr exposure of miniature Caspian horse semen to GSH which could not improve the plasma membrane integrity<sup>41,42</sup>. However, in another study, the addition of 2.5 mM GSH to the freezing extender could increase the spermatozoa membrane integrity<sup>43</sup>. This discrepancy may be related to the kind of semen extender and/or dosage of GSH.

Stallion sperm DNA integrity was ameliorated in the frozen-thawed semen in the presence of sericin. ROS in a dose-dependent manner is considered as a reason for DNA damage in the stallion spermatozoa. It has been shown that catalase but not superoxide dismutase prevents the DNA damage, which indicates that

hydrogen peroxide is the main ROS that leads to DNA damage in stallion spermatozoa<sup>44</sup>. Therefore, boosting catalase activity by sericin might be the reason for improvement of the sperm DNA integrity.

Diff-Quick staining showed that the number of spermatozoa with abnormal heads and midpieces in sericin group and spermatozoa with distal droplet in glutathione and sericin+glutathione groups was lower than the one in control group. It has been demonstrated that swelling of the midpiece, mitochondrial enlargement, is a major threat of cryo damage to spermatozoa that leads to the disruption of redox metabolism, promotion of ROS generation and induction of apoptosis<sup>3</sup>. It seems sericin keeps mitochondria from cryo damage which subsequently, resulted in reduction of ROS generation.

The sperm capacitation and acrosome-reaction are promoted by cryopreservation, which may have an adverse effect on fertility. However, both G and S+G groups could not ameliorate lipid peroxidation and enzymatic antioxidant activity. The percentage of intact and capacitated spermatozoa in all treated frozen-thawed groups was higher and lower than control, respectively. It has been demonstrated that capacitation is accompanied by an increase in intracellular calcium concentration, ROS generation, cAMP concentration, and elevation of tyrosine phosphorylation. In this context, catalase can suppress the sperm capacitation through inhibition of tyrosine phosphorylation<sup>45</sup>. Thus, promotion of catalase activity in the presence of sericin may suppress the capacitation state of the frozen thawed stallion spermatozoa. It seems that GSH has another mechanism in ameliorating the capacitation state of stallion spermatozoa. Previous studies on astrocytes and PC12 cells showed that depleted GSH levels were associated with a rise in intracellular calcium levels<sup>46,47</sup>. Accordingly, a possible hypothesis was proposed for the effect of antioxidants on capacitation state of frozen-thawed stallion spermatozoa in treatment groups. In fact, by increasing the amount of GSH in antioxidant treated groups, the amount of intracellular calcium decreases, resulting in a decrease in cAMP and tyrosine phosphorylation. Finally, these events reduce the percentage of capacitated sperm in antioxidant-treated groups.

### Conclusion

In conclusion, this study indicated that supplementation of semen extender with sericin prior to cryopreservation could improve the enzymatic antioxidant activity, DNA integrity, lipid peroxidation, capacitation state, and somehow the morphologic characteristics of the stallion sperm which may be applicable to other species.

### Acknowledgement

The authors would like to express their gratitude to staff of Research Institute of Animal Embryo Technol-

ogy, Shahrekord University, for their great technical support.

### Conflict of Interest

None of the authors has any conflict of interest to declare.

### References

- Curry MR. Cryopreservation of semen from domestic livestock. *Rev Reprod* 2000;5(1):46-52.
- Ball BA, Vo AT, Baumber J. Generation of reactive oxygen species by equine spermatozoa. *Am J Vet Res* 2001; 62(4):508-515.
- Ball BA. Oxidative stress, osmotic stress and apoptosis: impacts on sperm function and preservation in the horse. *Anim Reprod Sci* 2008;107(3-4):257-267.
- Baumber J, Ball BA. Determination of glutathione peroxidase and superoxide dismutase-like activities in equine spermatozoa, seminal plasma, and reproductive tissues. *Am J Vet Res* 2005;66(8):1415-1419.
- Ball BA, Gravance CG, Medina V, Baumber J, Liu IK. Catalase activity in equine semen. *Am J Vet Res* 2000;61(9):1026-1030.
- Kefer JC, Agarwal A, Sabanegh E. Role of antioxidants in the treatment of male infertility. *Int J Urol* 2009;16: 449-457.
- Alvarez JG, Storey BT. Taurine, hypotaurine, epinephrine and albumin inhibit lipid peroxidation in rabbit spermatozoa and protect against loss of motility. *Biol Reprod* 1983;29(3):548-555.
- Thuwanut P, Chatdarong K, Techakumphu M, Axner E. The effect of antioxidants on motility, viability, acrosome integrity and DNA integrity of frozen-thawed epididymal cat spermatozoa. *Theriogenology* 2008;70(2): 233-240.
- Michael A, Alexopoulos C, Pontiki E, Hadjipavlou-Litina D, Saratsis P, Boscoc C. Effect of antioxidant supplementation on semen quality and reactive oxygen species of frozen-thawed canine spermatozoa. *Theriogenology* 2007;68(2):204-212.
- Baghshahi H, Riasi A, Mahdavi A, Shirazi A. Antioxidant effects of clove bud (*Syzygium aromaticum*) extract used with different extenders on ram spermatozoa during cryopreservation. *Cryobiology* 2014;69(3):482-487.
- Bucak MN, Sariözkan S, Tuncer PB, Ulutaş PA, Akçadağ Hİ. Effect of antioxidants on microscopic semen parameters, lipid peroxidation and antioxidant activities in Angora goat semen following cryopreservation. *Small Ruminant Res* 2009;81:90-95.
- Foote RH, Brockett CC, Kaproth MT. Motility and fertility of bull sperm in whole milk extender containing antioxidants. *Anim Reprod Sci* 2002;71(1-2):13-23.
- Reddy NSS, Mohanarao GJ, Atreja S. Effects of adding taurine and trehalose to a tris-based egg yolk extender on buffalo (*Bubalus bubalis*) sperm quality following cryopreservation. *Anim Reprod Sci* 2010;119(3-4):183-190.
- Baumber J, Ball BA, Linfor JJ. Assessment of the cryopreservation of equine spermatozoa in the presence of enzyme scavengers and antioxidants. *Am J Vet Res* 2005;66:772-779.
- Li TK. The glutathione and thiol content of mammalian spermatozoa and seminal plasma. *Biol Reprod* 1975;12: 641-646.
- Bilodeau JF, Blanchette S, Gagnon C, Sirard MA. Thiols prevent H<sub>2</sub>O<sub>2</sub>-mediated loss of sperm motility in cryopreserved bull semen. *Theriogenology* 2001;56(2):275-286.
- Gadea J, Molla M, Selles E, Marco M, Garcia-Vazquez F, Gardon J. Reduced glutathione content in human sperm is decreased after cryopreservation: effect of the addition of reduced glutathione to the freezing and thawing extenders. *Cryobiology* 2011;62(1):40-46.
- Gadea J, Sellés E, Marco MA, Coy P, Matás C, Romar R, et al. Decrease in glutathione content in boar sperm after cryopreservation: Effect of the addition of reduced glutathione to the freezing and thawing extenders. *Theriogenology* 2004;62:690-701.
- Gadea J, García-Vazquez F, Matás C, Gardón JC, Cánovas S, Gumbao D. Cooling and freezing of boar spermatozoa: supplementation of the freezing media with reduced glutathione preserves sperm function. *J Androl* 2005;26(3):396-404.
- Salmani H, Nabi MM, Vaseghi Dodaran H, Rahman MB, Mohammadi Sangcheshmeh A, Shakeri M, et al. Effect of glutathione in soybean lecithin-based semen extender on goat semen quality after freeze-thawing. *Small Ruminant Res* 2013;112:123-127.
- Zhang YQ. Applications of natural silk protein sericin in biomaterials. *Biotechnol Adv* 2002;20(2):91-100.
- Voegeli R. Sericin silk protein: unique structure and properties. *Cosmetics Toiletries* 1993;108:101-108.
- Tao W, Li M, Xie R. Preparation and structure of porous silk sericin materials. *Macromol Mater Eng* 2005;290(3):188-194.
- Kato N, Sato S, Yamanaka A, Yamada H, Fuwa N, Nomura M. Silk protein, sericin, inhibits lipid peroxidation and tyrosinase activity. *Biosci Biotechnol Biochem* 1998; 62(1):145-147.
- Isobe T, Ikebata Y, Onitsuka T, Wittayarat M, Sato Y, Taniguchi M, et al. Effect of sericin on preimplantation development of bovine embryos cultured individually. *Theriogenology* 2012;78(4):747-752.
- Sasaki M, Kato Y, Yamada H, Terada S. Development of a novel serum-free freezing medium for mammalian cells using the silk protein sericin. *Biotechnol Appl Biochem* 2005;42(Pt 2):183-188.
- Ohnishi K, Murakami M, Morikawa M, Yamaguchi A. Effect of the silk protein sericin on cryopreserved rat islets. *J Hepatobiliary Pancreat Sci* 2012;19(4):354-360.
- Freitas ML, Bouéres CS, de Oliveira FG, de Oliveira Viu M, de Oliveira RA. Comparison of two different centrifugation extenders for preservation of frozen equine semen. *Equine Vet Educ* 2015;27(8):410-413.

29. Khlifaooui M, Battut I, Bruyas JF, Chatagnon G, Trimèche A, Tainturier D. Effects of glutamine on post-thaw motility of stallion spermatozoa: an approach of the mechanism of action at spermatozoa level. *Theriogenology* 2005;63(1):138-149.
30. Vidament M, Dupere A, Julienne P, Evain A, Noue P, Palmer E. Equine frozen semen: freezability and fertility field results. *Theriogenology* 1997;48:907-917.
31. Buege JA, Aust SD. Microsomal lipid peroxidation. *Methods Enzymol* 1978;52:302-310.
32. Kakkar P, Das B, Viswanathan P. A modified spectrophotometric assay of superoxide dismutase. *Indian J Biochem Biophys* 1984;21(2):130-132.
33. Lawrence RA, Burk RF. Glutathione peroxidase activity in selenium-deficient rat liver. *Biochem Biophys Res Commun* 1976;71:952-958.
34. Goth L. A simple method for determination of serum catalase activity and revision of reference range. *Clin Chim Acta* 1991;196(2-3):143-151.
35. Schembri M, Major D, Maxwell W, Suttie J, Evans G. Capacitation-like changes in equine spermatozoa throughout the cryopreservation process. *Reprod Fertil Dev* 2002;14:225-233.
36. Kumar P, Kumar D, Sikka P, Singh P. Sericin supplementation improves semen freezability of buffalo bulls by minimizing oxidative stress during cryopreservation. *Anim Reprod Sci* 2015;152:26-31.
37. Hu JH, Zan LS, Zhao XL, Li QW, Jiang ZL, Li YK, et al. Effects of trehalose supplementation on semen quality and oxidative stress variables in frozen-thawed bovine semen. *J Anim Sci* 2010;88(5):1657-1662.
38. Takechi T, Wada R, Fukuda T, Harada K, Takamura H. Antioxidant activities of two sericin proteins extracted from cocoon of silkworm (*Bombyx mori*) measured by DPPH, chemiluminescence, ORAC and ESR methods. *Biomed Rep* 2014;2(3):364-369.
39. Fan JB, Wu LP, Chen LS, Mao XY, Ren FZ. Antioxidant activities of silk sericin from silkworm *Bombyx mori*. *J Food Biochem* 2009;33:74-88.
40. Lasso JL, Noilws EE, Alavarez JG, Storey BT. Mechanism of superoxide dismutase loss from human sperm cells during cryopreservation. *J Androl* 1994;15(3):255-265.
41. Jones R, Mann T, Sherins R. Peroxidative breakdown of phospholipids in human spermatozoa, spermicidal properties of fatty acid peroxides, and protective action of seminal plasma. *Fertil Steril* 1979;31(5):531-537.
42. Zhandi M, Ghadimi V. Effect of glutathione-supplemented INRA82 extender on miniature Caspian stallion sperm quality during storage at 5 C. *J Equine Vet Sci* 2014;34:606-610.
43. de Oliveira RA, Wolf CA, de Oliveira Viu MA, Gambarini ML. Addition of glutathione to an extender for frozen equine semen. *J Equine Vet Sci* 2013;33:1148-1152.
44. Baumber J, Ball BA, Linfor JJ, Meyers SA. Reactive oxygen species and cryopreservation promote DNA fragmentation in equine spermatozoa. *J Androl* 2003;24(4):621-628.
45. Aitken R, Paterson M, Fisher H, Buckingham D, Van Duin M. Redox regulation of tyrosine phosphorylation in human spermatozoa and its role in the control of human sperm function. *J Cell Sci* 1995;108(Pt 5):2017-2025.
46. Jurma OP, Hom DG, Andersen JK. Decreased glutathione results in calcium-mediated cell death in PC12. *Free Radic Biol Med* 1997;23(7):1055-1066.
47. Abramov AY, Canevari L, Duchon MR. Changes in intracellular calcium and glutathione in astrocytes as the primary mechanism of amyloid neurotoxicity. *J Neurosci* 2003;23(12):5088-5095.