CRISPR/Cas9 System for Efficient Genome Editing and Targeting in the Mouse NIH/3T3 Cells

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Abstract

Background: The Clustered, Regularly Interspaced, Short Palindromic Repeats (CRISPR) and CRISPR-associated protein (Cas) system has been used as a powerful tool for genome engineering. In this study, the application of this system is reported for targeting Rag genes to produce mutant mouse NIH/3T3 cell line. The Rag1 and Rag2 genes are essential for generation of mature B and T lymphocytes. Disruption of Rag genes causes disease like Severe Combined Immunodeficiency syndrome (SCID). Here, the efficiency and specificity of CRISPR system were tested with highly active sgRNAs to generate novel mutations in the NIH/3T3 mouse cell line.

Methods: Four single guide RNAs were designed to target sequences in the coding region of the Rag1 and Rag2 genes. Four sgRNA-Cas9 plasmids were tested to target Rag1 and Rag2.

Results: Based on T7 endonuclease assay and sequencing analysis, the expression of sgRNAs targeting two sites in Rag1 resulted in deletion of the intervening DNA fragment. The expression of sgRNAs with Cas9 targeting two sites in Rag2 gene resulted in indel mutations at both sites. In this report, fragment deletion in Rag1 gene was detected in about 50% of transfected cells.

Conclusion: Therefore, CRISPR/Cas9 system can be highly efficient and specific when sgRNAs are designed rationally and provides a powerful approach for genetic engineering of cells and model animals.

Keywords: Cell line, Deletion, Gene editing, Mice

Introduction

Recently, the Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) system associated to the Cas9 endonuclease (CRISPR/Cas9) has been developed as a specific and effective tool for genome engineering. Compared to other genome editing technologies based on endonucleases such as Zinc Finger Nuclease (ZFN) and Transcription Activator-Like Effector Nuclease (TALEN) that are rather complex to design and need to be assembled for each target sequence, the CRISPR/Cas9 is inexpensive and easy to carry out. 1-5.

This system is based on the base-pairing of the genomic sequence, adjacent to an obligate protospacer adjacent motif (PAM) NGG, with a short complimentary RNA sequence which is then cleaved by the Cas9 protein in a sequence-specific manner. 5-7. Double Strand Breaks (DSBs) induced by this system can be repaired in one of the two ways: Non-Homologous End Joining (NHEJ) or Homology-Directed Repair (HDR), resulting in insertions or deletions in desired genomic regions. Deletions may result from introducing two sgRNAs along with Cas9. The sgRNAs induce two DSBs and deletion of the intervening fragment may occur. There are several advantages of genomic deletions compared to single-site small indels including: 1) easy identification by conventional PCR, while identification of small indels or point mutations may require time consuming techniques such as Restriction Fragment Length Polymorphism (RFLP), immunoblotting, T7 Endonuclease (T7EN1) cleavage assay and 2) predictability loss of gene function. 8

The CRISPR-Cas system has been used for genome editing in several organisms 5,9-13 and mammalian cell lines 14-17. Although genome editing by CRISPR-Cas
system has been demonstrated to be non-specific in some cases \(^\text{15}\), it can be improved in specificity if gRNAs are designed correctly.

Currently, a variety of animal models with natural immune system defects can be applied for the immunology and genetic studies \(^\text{19}\). The Rag1 and Rag2 genes play an important role in the rearrangement and recombination of immunoglobulin and T-cell receptor genes during the process of V(D)J recombination. In mouse models with RAG-1 and RAG-2 deficiency, deletion of the Rag-1/Rag-2 genes cause the arrest of rearrangement of B-cell receptors (Immunoglobulin production) and T-cell receptors and lack of the T and B cell differentiation.

In recent years, programmable site-specific nucleases have been used to generate recombination-activating genes (RAG1/RAG2) immunodeficient mouse models \(^\text{20-22}\). As mentioned above, the RAG proteins are essential for the V(D)J recombination process, which generates diversity of immune systems. RAG-1/RAG-2 deficient mice do not produce mature B and T lymphocytes. The loss of RAG function leads to a Severe Combined Immunodeficiency (SCID) phenotype \(^\text{23}\).

In this study, knock-out mouse cell line models of RAG1/RAG2 were created. The following procedures were performed; identification of appropriate target sites on Rag genes to design sgRNAs based on rules handling on-target efficacy, construction of CRISPR/CAS9 plasmids by cloning strategy, co-transfection of CRISPR/CAS9 plasmids and puromycin resistance vector into mouse model NIH/3T3 cell lines and finally analysis of mutagenesis induced by CRISPR system in transfected cell line.

**Materials and Methods**

**Target site selection**

Target sequences G/A-(N19)-NGG were selected for U6 plasmid-base transcription. The target sequence can be present on either DNA strand. To choose the most specific target for sgRNA, the potential for off-target effects should be minimized. For this purpose, there are several computational tools allowing the design of sgRNA target sequences with minimum potential for off-targets \(^\text{24-26}\).

In our study, assessment of off-target searching was performed using the online CRISPR RGEN Tools program (https://www.rgenome.net). Therefore, target selection was done based on following criteria; 5’-G-Ny-GGG-3’ matching and selection of target sequences in coding region nearby gene promoter. Finally, four target sequences in coding regions of Rag1 and Rag2 genes were selected where two sequences targeted the Rag1 gene and two other sequences targeted the Rag2 gene, respectively.

**sgRNA design**

Finally, based on the mentioned rules for target selection, sgRNA sequences were ordered in the form of phosphorlated forward and reverse oligonucleotides flanked by desired restriction sites. These sgRNA oligonucleotides can be annealed to generate linkers compatible with restriction sites of digested sgRNA/Cas9 expressing vector.

**Construction of sgRNA and Cas9 expressing vector**

In our experiment, two kinds of CRISPR expressing vectors were used: pX330 vector (Addgene#42230) and pLenti-Cas-Guide vector (Origene, GE100010) to express codon optimized spCas9 under CMV (in pX330) or CBh (in pLenti-Cas-Guide) promoter and sgRNA under U6 promoter. Antibiotic resistance genes used as the selection markers were ampicillin and chloramphenicol in pX330 and pLenti-Cas-Guide vectors, respectively.

In order to target four sequences in coding regions of Rag1 and Rag2 genes, desired sgRNA/Cas9 vectors were constructed. To clone a sgRNA into pX330 expression vector, the vector was digested with BbsI (Thermo scientific, ER1011). Similarly, the pLenti-Cas-Guide vector was digested with BamHI (Thermo scientific, ER0051) and BsmBI (Thermo scientific, ER0451) restriction enzymes, then both vectors were treated with Alkaline Phosphatase (Thermo scientific, EL0011). Complementary oligos of RAG1-F1 RAG1-R1, RAG1-F2 RAG1-R2, RAG2-F1 RAG2-R1 and RAG2-F2 RAG2-R2 (Table 1) for each target sequence were heated at 95°C for 5 min, and annealed by decreasing the temperature from 0.5°C/C to 22°C using a thermocycler (Eppendorf, USA). Then, the short double strand DNA fragments (RAG1-F1R1, RAG2-F1R1 and RAG2-F2R2) were ligated into linearized BbsI site of pX330 and RAG1-F2R2 was ligated into linearized

<table>
<thead>
<tr>
<th>sgRNA names</th>
<th>Sequence 5' to 3'</th>
<th>Restriction site</th>
</tr>
</thead>
<tbody>
<tr>
<td>RAG1-F1</td>
<td>CACCGTGCGACGAGTTCGCCG</td>
<td>EcoRI</td>
</tr>
<tr>
<td>RAG1-R1</td>
<td>AAAACGCGGAGACGGAACGTCGAC</td>
<td>EcoRI</td>
</tr>
<tr>
<td>RAG1-F2</td>
<td>GATCCTATGCGAGATTCGTCGAC</td>
<td>EcoRI</td>
</tr>
<tr>
<td>RAG1-R2</td>
<td>AAAACCCCGAGCAATTCGCCATGC</td>
<td>EcoRI</td>
</tr>
<tr>
<td>RAG2-F1</td>
<td>CACCGGAATGGCGGATATCCGGTTC</td>
<td>EcoRI</td>
</tr>
<tr>
<td>RAG2-R1</td>
<td>AAAGGAAAGCTGATACGCCATC</td>
<td>EcoRI</td>
</tr>
<tr>
<td>RAG2-F2</td>
<td>CACCGGTATATGCGAAGGGAAAGCA</td>
<td>EcoRI</td>
</tr>
<tr>
<td>RAG2-R2</td>
<td>AAAGCTGCTTCCCTCGACTATAC</td>
<td>EcoRI</td>
</tr>
</tbody>
</table>
 BamHI and BsmBI sites of pLenti-Cas-Guide vector. Ligation products were transformed into *Escherichia coli* (E. coli) DH5α competent cells to get clones. *E. coli* were grown in LB (Luria-Bertani) medium containing desired antibiotics as selective markers.

**Restriction enzyme digestion and sequencing**

In order to confirm ligation and correct direction of inserts, digestion with restriction enzymes and sequencing were performed. For the restriction enzyme digestion, the cloned pX330 vector (In which BbsI restriction site has been removed after cloning) was digested with BbsI restriction enzyme and the cloned pLenti-Cas-Guide vector (Since RAG1-F2R2 sgRNA contains *EcoRI* restriction site, so the new construct presents with an additional *EcoRI* restriction site) was digested with *EcoRI*. For the sequencing, desired primers for regions surrounding the target sites in Rag1 and Rag2 genes were designed (Table 2). The sequencing results were analyzed by sequence alignment program clustalW2 to confirm correct direction of inserts.

**Transfection of Cas9-sgRNA vector into NIH/3T3 cell line**

Among reagents for lipofection, lipofectamine 2000 has the highest efficiency for NIH/3T3 transfection. In our experiments, pLKO.1-puro vector (Addgene #10878) containing puromycin resistance gene was used for the selection of cells that have been transfected by sgRNA-Cas9 vector because pX330 and pLenti plasmids do not contain any markers for selection. Therefore, NIH/3T3 cell lines were transiently co-transfected with pLKO.1-puro vector and Cas9-sgRNA vector using lipofectamine 2000.

**Cell culture and Cas9-sgRNA vector transfection**

NIH/3T3 cell lines were grown in DMEM media supplemented with 10% FBS. Cells were co-transfected with sgRNA-Cas9 expressing vector and pLKO.1 vector using lipofectamine 2000 (Invitrogen, 11668) according to manufacturer's protocol. For this purpose, one day prior to transfection, the cells were seeded in a 6-well plate at a cell density of 2x10^5 cells per well until they reached ~50% confluence. For plasmid DNA transfection, 2.5 μg DNA (1.9 puromycin: sgRNA ratio) was added to 150 μl of Opti-MEM medium (Invitrogen, 31985), followed by addition of 150 μl of Opti-MEM containing 5 μl of Lipofectamine 2000. The sample was mixed by gently flicking the tubes a few times and then the mixture was incubated at room temperature for 20 min. The entire solution was added to the cells in a 6-well plate and mixed by gently swirling the plate. The plate was incubated at 37°C for 48 hr in a 5% CO₂ incubator. Medium was changed 48 hr after transfection with a fresh medium containing 7.5 μg puromycin for each well. Puromycin selection was done over 3-5 days. After selection, transfected cells were assayed. To monitor transfection efficiency, transfection control wells (GFP plasmid) were used.

**T7E1 mismatch-detecting assay**

Genomic DNA from transfected and control NIH/3T3 cells was isolated by DNA extraction kit (Qiagen). Fragments containing Rag1 and Rag2 genes were amplified from extracted DNA using Rag1 and Rag2 primers that had been designed in the vicinity of the target sites in Rag1 and Rag2 genes, respectively (Table 2). Purified PCR products (100 ng) were denatured and renatured under the following thermocycler conditions: 95°C for 5 min, 95 to 85°C at -2°C/s, 85 to 25°C at -0.1°C/s, and held at 4°C. Then, 2 μl of NEB buffer 2 (New England Biolabs, Ipswich, MA, USA), 1 μl (5 U) of T7 endonuclease I (New England Biolabs, M-0302S), and H₂O were added to total volume of 20 μl and the mixture was incubated at 37°C for 30 min. Finally, treated PCR products were analyzed by 2% agarose gel to visualize the cleavage bands. The non-transfected control sample was also treated by T7E1 and used as a negative control, since sometimes minor background bands may be present even in the negative control sample due to the unspecific cleavage of T7E1 nuclease.

**Sequencing analysis**

The PCR products were sequenced by Sanger sequencing with desired Rag1 and Rag2 primers and mutations were confirmed by sequencing (Macrogen, Korea).

**Results**

**Target selection, design and characterization of Cas9 and sgRNAs expressing vector**

To design effective sgRNAs for editing the mouse Rag1/Rag2 genes, the sequence within coding regions of Rag1/Rag2 genes was analyzed through the online CRISPR RGEN Tools program (https://www.rgenome.net).

We targeted two regions in each Rag1/Rag2 gene in which two sgRNAs separated by 213 bp for Rag1 and 2 sgRNAs separated by 12 bp for Rag2 (Figure 1A). Potential target sites were selected based on predictive rules to avoid off-179 targets (see Materials and Methods). We targeted two regions in each Rag1/Rag2 gene in which two 180 sgRNAs were designed.

We assembled sgRNAs in separate expression vectors using cloning steps as described in materials and methods. Only plasmid DNA was used to transfected, since RNA preparation, handling and storage take laborious steps. Four sgRNAs were constructed into vectors with sgRNA scaffolds driven by a U6 promoter and Cas9 under CBh and CMV promoter in pX330 and

| Table 2. Primers used to amplify fragments of RAG1 and RAG2 genes |
|-----------------|------------------|
| Primer names    | Sequence 5’ to 3’ |
| RAG1-primer F   | GAA GAA GCA CAG AAG GAG AAG |
| RAG1-primer R   | ATC GGC AGG AGC GAC AAT AGC |
| RAG2-primer F   | ATTCCTCCTGCAAGACT |
| RAG2-primer R   | GCATAAGACTGCAAGCA |
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To verify ligation, sequencing and diagnostic restriction digestion were performed (Figure 1C). pX330 has been designed so that when the plasmid is cloned with ad hoc sgRNA, BbsI restriction site is lost. Conversely, cloning the guide sequence into the pLenti vector generates an additional EcoRI restriction site. The results of sequencing and diagnostic digestion demonstrated that sgRNAs have been successfully constructed in CRISPR/Cas9 vectors. For constructing RAG1-F1R1/pX330, among several clones only two clones (clone 2 and 4), for constructing RAG1-F2R2/pLenti, clone 1 and 2, for constructing RAG2-F1R1/pX330, clone 4 and 5 had the correct ligation based on the results of restriction enzyme digestion and sequencing. For constructing RAG1-F2R2/pX330, clone 1 and 2 had the correct ligation based on the results of restriction enzyme digestion but pLenti-Cas-Guide, respectively (Figure 1B).
only clone 2 had the correct insert direction based on the results of sequencing.

**sgRNA-Cas9 guided genome editing in Rag genes**

Rag1 and Rag2 sgRNAs targeted two sites simultaneously in each gene. To analyze genome editing by sgRNA-Cas9, genomic DNA was isolated from cells harvested 3-5 days after transient transfection. The rate of transfection was visualized about 50% based on EGFP fluorescence using fluorescent microscopy (Figure 2). Extracted DNA was analyzed for the presence of site-specific gene modification by PCR amplification of regions surrounding the target sites as well as T7EN1 cleavage assay. Fragments resulted from Rag1 PCR amplification showed that before T7EN1 cleavage assay, there were two bands of Rag1 fragment on agarose gel in transfected lane compared with wild type lane. This smaller amplicon was the result of the expected deletion that occurred in the Rag1 gene in a large number of cells (Figure 3A). Since gel electrophoresis after cleavage activity showed the occurrence of the fragment deletion in Rag1 gene, there was no need to subject the PCR products of Rag1 to T7EN1 cleavage assay. Deleted fragment ~213 bp in size could rapidly be detected by PCR without requirement for T7EN1 cleavage assay since deletion has occurred in many cells. Evidence of nuclease cleavage of Rag2 gene could not be detected by PCR amplification and only was visible after T7EN1 cleavage assay (Figure 3B).

**Sequencing of PCR product**

To confirm the results of cleavage assay, Sanger sequencing was used and showed the indels were detected at target sites (Figure 4).

**Discussion**

In the present study, it was shown that genome editing by CRISPR/Cas9 system is efficient with highly active sgRNAs to generate novel mutations of Rag1/Rag2 genes in the NIH/3T3 mouse cell line.

Amazing result for Rag1 gene showed that before T7EN1 cleavage assay, only by a simple PCR reaction an obvious deletion was detected in Rag1 gene in a large number of cells. These results of mutation detection in Rag1/Rag2 genes demonstrate that CRISPR/Cas9 system represents an effective and potential genome editing tool in NIH/3T3 cell line. It was also shown that deletion of a gene fragment with high effi-
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ciency can occur in Rag genes by simultaneous cleavage at two targeted sites in one gene. Such results have been reported in other cases \(^ {8,30-33}\) but in this report significant deletion has been achieved in about 50% of cellular population. Taken together, these results showed that CRISPR/Cas9 system with highly active and correctly-designed sgRNAs generate mutations in desired genes and efficient deletions can be achieved using two exonic sgRNAs targeting one gene. Compared to frame shift mutations, genomic deletion can be useful for generation of specific gene knockouts.

The sequencing results demonstrated that the selected sgRNAs worked effectively with Cas9 on Rag1 and Rag2 genes in NIH/3T3 cell lines. Rag1/Rag2 gene targeting by CRISPR/Cas9 system makes deletions one-step in about 50% of cell lines. CRISPR/Cas9 system was applied with highly active sgRNAs for targeting the Rag1/Rag2 genes to make mutant cell lines. The CRISPR/Cas9 system is much easier than other systems like ZFN and TALEN to genome engineering. High efficiency editing by CRISPR/Cas9 system can be achieved in mouse cell line genomes at targeted locations with efficient and well-designed sgRNAs. Genome editing results indicated that CRISPR/Cas9 system with correctly-designed sgRNAs generate mutations in desired genes and significant deletions can be achieved in the large number of cells using two exonic sgRNAs targeting one gene.

Conclusion

It was shown that CRISPR/Cas9 system can be highly efficient when gRNAs are designed rationally and provides a powerful approach for genetic engineering of cells and model animals.

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