

## Extraction, Purification and Characterization of Lipopolysaccharide from *Escherichia coli* and *Salmonella typhi*

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### Abstract

Lipopolysaccharide (LPS) is an important structural component of the outer cell membrane complex of gram negative microorganisms. Its causative role in gram negative bacteria-induced diseases and broad applications in different kinds of cell stimulation experiments provided a conceptual basis for studies directed at the isolation, purification, and detailed chemical characterization of LPS. The main problem with LPS purification protocols is the contamination of the end product with nucleic acids and proteins in variable proportions which could potentially interfere with downstream applications. In this study, a simple procedure for purification of LPS from *Escherichia coli* (*E.coli*) and *Salmonella typhi* (*S.typhi*) with high purity and very low contaminating nucleic acids and proteins based on the hot phenol-water extraction protocol has been introduced. The purity of extracted LPS was evaluated by silver and coomassie blue staining of SDS-PAGE gels and HPLC analysis. Limulus Amebocyte Lysate (LAL) coagulation activity and rabbit pyrogen assay were exploited to monitor the functionality of purified LPS. The results showed that DNase and RNase treatment of the sample is essential after the sonication step to eliminate nucleic acid contamination in the LPS fraction. Silver staining demonstrated ladder pattern which is characteristic of LPS. No contaminating protein was found as assessed by coomassie blue staining. HPLC fractionation revealed high degree of purity comparable with commercial LPS. Parenteral administration of purified LPS resulted in substantial increase of rabbits' body temperature (mean: 1.45 °C). LAL coagulation assay confirmed the functional activity of the purified LPS. In conclusion, the protocol presented here could be employed for isolation of LPS with high purity and functional activity.

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**Keywords:** *Escherichia coli*, Endotoxin, Extraction, Lipopolysaccharide, Purification, *Salmonella typhi*

### Introduction

Lipopolysaccharide (LPS) is the main outer membrane component of gram negative bac-

teria which constitutes about 75% of the surface<sup>(1)</sup> and 5-10% of the total dry weight

of gram negative bacteria <sup>(2)</sup>. Their basic structure consists of three parts: lipid A, core oligosaccharide and repetitive polysaccharide designated as "O" antigen. Lipid A is highly conserved and exerts the endotoxic activity, while the "O" antigen carbohydrate chain is a polymer of repeating oligosaccharides, which differs between species and is responsible for the serological specificity of bacteria <sup>(3)</sup>. LPS causes pathophysiological effects such as fever, leucopenia, leucocytosis and Shwartzman reactivity <sup>(4,5)</sup>.

The recognition of very significant causative role of LPS in gram negative bacteria-induced diseases prompted many researchers to conduct studies directed at its isolation and purification. Therefore, it is not surprising that a plenty of methods and protocols have been introduced for the isolation and purification of LPS from bacteria included among them are trichloroacetic acid extraction at 4°C <sup>(6)</sup>, aqueous butanol <sup>(7)</sup>, triton/Mg+2 <sup>(8)</sup>, cold ethanol <sup>(9)</sup> and extraction in water at 100°C <sup>(10)</sup>. Other purification protocols with phenol, chloroform, petroleum-ether <sup>(11)</sup> and methanol <sup>(12)</sup> have been described specifically for rough LPS.

Recently a combination of Westphal method based on the hot phenol extraction procedure and sized exclusion chromatography was successfully employed for the purification of LPS from *E.coli* <sup>(13)</sup>. The method proposed by Westphal <sup>(14)</sup> is still the most frequent procedure employed for LPS extraction because of its high yield. Contamination with proteins and nucleic acids are among the main disadvantages of some proposed protocols for LPS purification which hinder reliable application of the end product in such sensitive assays as molecular and immunological experiments. Although ultracentrifugation can be employed for elimination of contaminating proteins <sup>(7,14)</sup>, this usually leads to lower yields and considerable amount of nucleic acids contaminate the sedimented LPS <sup>(14)</sup>.

In this view, no single method is suited to

isolation of LPS with high purity and combination of two purification steps maybe necessary. In the present study, we have employed a modified phenol-water extraction protocol accompanied with proteinase K digestion of bacterial proteins and nuclease elimination of nucleic acids for extraction of LPS from *E.coli* and *S.typhi* with high purity.

## Materials and Methods

### *Bacterial strains and growth conditions*

*E.coli* 055:B5 (Pasteur Institute, Paris, France) and *Salmonella typhi* B-34-2 (Pasteur Institute, Tehran, Iran) were grown in Luria-Bertani broth medium (usb, Cleveland, USA) at 37°C in shaker incubator overnight. After centrifugation of culture media, sedimented bacteria were harvested and used for LPS extraction and purification.

### *LPS extraction and purification*

LPS was extracted by hot phenol-water method as described previously with some modifications <sup>(14)</sup>. In brief, bacterial suspensions ( $10^8$  colony-forming *units/mL*) were centrifuged at  $10,000\times g$  for 5 *min*. The pellets were washed twice in PBS (pH=7.2) (0.15 M) containing 0.15 mM CaCl<sub>2</sub> and 0.5 mM MgCl<sub>2</sub>. Pellets were then resuspended in 10 ml PBS and sonicated for 10 *min* on ice.

In order to eliminate contaminating protein and nucleic acids, treatment with proteinase K, DNase and RNase was performed prior to extraction step. For this purpose, proteinase K (100 µg/mL) (Roche, Mannheim, Germany) was added to the cell mixture and the tubes were kept at 65°C for an additional hour. Mixture was subsequently treated with RNase (40 µg/mL) (Roche, Mannheim, Germany) and DNase (20 µg/mL) (Roche, Mannheim, Germany) in the presence of 1 µL/mL 20% MgSO<sub>4</sub> and 4 µL/mL chloroform and incubation was continued at 37 °C overnight.

At the next step, an equal volume of hot (65-70°C) 90% phenol was added to the mixtures followed by vigorous shaking at 65-70°C for 15 *min*. Suspensions were then cooled on ice, transferred to 1.5 mL polypro-

pylene tubes and centrifuged at  $8500\times g$  for 15 *min*. Supernatants were transferred to 15 *mL* conical centrifuge tubes and phenol phases were re-extracted by 300  $\mu\text{L}$  distilled water. Sodium acetate at 0.5 *M* final concentration and 10 volumes of 95% ethanol were added to the extracts and samples were stored at  $-20^{\circ}\text{C}$  overnight in order to precipitate LPS.

Tubes were then centrifuged at  $2000\times g$   $4^{\circ}\text{C}$  for 10 *min* and the pellets were resuspended in 1 *ml* distilled water. Extensive dialysis against double distilled water at  $4^{\circ}\text{C}$  was carried out at the next step until the residual phenol in the aqueous phases was totally eliminated. Final purified LPS product was lyophilized and stored at  $4^{\circ}\text{C}$ .

#### **Silver, coomassie blue and ethidium bromide stainings**

The purified LPS was solubilized in sample buffer to the desired concentration (1 *mg/mL*), and boiled for 5 *min*. 16  $\mu\text{L}/\text{well}$  from each sample was separated on 15% SDS gel with a 4% stacking gel under reducing condition at 100 *mA* for 2 *hr* using mini-PROTEAN electrophoresis instrument (Bio-Rad Laboratories, California, USA). Silver and coomassie blue staining of the gels was performed according to the standard protocol. Staining of agarose gel with ethidium bromide was done as well in order to show if any contamination with nucleic acids persist. To do this, 10  $\mu\text{L}/\text{well}$  of reconstituted LPS from both *E.coli* and *S.typhi* and also 10  $\mu\text{L}/\text{well}$  bacterial suspensions ( $10^8$  colony-forming *units/mL*), as positive control, were loaded on agarose gel and stained with ethidium bromide.

#### **High performance liquid chromatography (HPLC)**

HPLC separations were carried out by a Knauer Smartline 1000 pump equipped with a Smartline UV detector 2500 (Berlin, Germany), and a Rheodyne 7725 injection valve (Cotati, CA, USA). The method was optimized at the 0.8 *ml/min* flow rate and 210 *nm* wavelength UV detection. Separation was carried out over  $\text{C}_{18}$  column with 4.6 *mm* diameter and 250 *mm* length from Grace Company (Munich, Germany) with a mixture

of water and acetonitrile (95:5) as mobile phase. Extra pure *E.coli* LPS (Sigma, Saint Louis, USA) was used as standard.

#### **Limulus amoebocyte lysate (LAL) assay**

The potency of LPS samples were determined by the limulus amoebocyte assay gel-clot method (LONZA, Walkersville, USA) which had a sensitivity of 0.06 *endotoxin units per milliliter (UE/mL)*, according to the protocol published elsewhere<sup>(15)</sup>.

#### **Rabbit pyrogen test**

Two white New Zealand rabbits weighing between 1.7 and 2.3 *kg* (Pasture Institute, Tehran, Iran) were used. Pyrogenicity was tested in an air conditioned room. Rabbits were given an intravenous injection of 5 *ng/kg* of body weight of purified LPS. Rectal temperatures were measured with indwelling rectal thermostats and recorded before injection and 4 *hr* after pyrogen administration. One rabbit was injected with sterile PBS as control.

## **Results**

#### **SDS-PAGE separation and silver and coomassie blue staining of LPS**

Separation over SDS-PAGE gel followed by silver staining was used to detect and visually characterize the purified LPS. Silver staining is a highly sensitive method capable of detecting as low as 1 *ng* LPS and is routinely used for visualization of the band pattern of purified LPS.

As depicted in figure 1A, LPS from both *E.coli* and *S.typhi* gave a characteristic dark staircase (ladder-like) pattern of bands. Lipid A-core LPS migrating very near the dye-front stained very intensely and appeared as black region at the bottom of gel. The results also showed different profile of LPS banding in the two strains studied. Coomassie blue staining of the gels showed no band indicating absence of contaminating proteins (Figure 1B). Staining of agarose gel with ethidium bromide and absence of any band in purified LPS products showed no contamination with nucleic acid (Figure 1C).

## Purification of Lipopolysaccharide

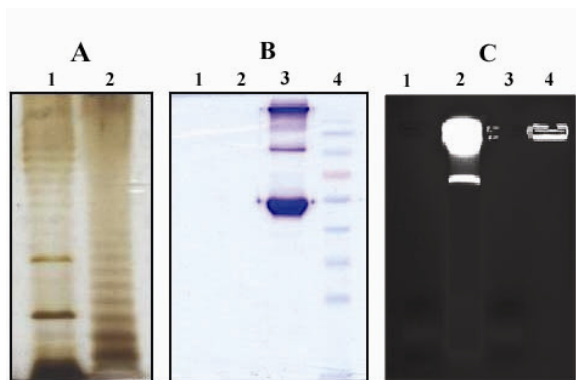


Figure 1. Silver, coomassie blue and ethidium bromide stainings of purified LPS

LPS from *E. coli* (Lanes 1A and 1B) and *S. typhi* (lanes 2A and 2B) was purified by modified hot phenol-water extraction method and fractionated by SDS-PAGE electrophoresis followed by silver (A) or coomassie blue staining (B). Ladder pattern of LPS banding which is characteristic of smooth gram negative bacteria is seen (A). The absence of band in coomassie blue staining as shown in B indicates no contamination of purified LPS with bacterial proteins. Lane 3B: Human IgG and BSA, Lane 4B: Molecular weight marker. Residual nucleic acid contamination in purified LPS products was traced by ethidium bromide staining (C). Absence of band in LPS from *E. coli* (Lane 1C) and *S. typhi* (Lane 3C) shows no contamination with nucleic acids in purified LPS products. Lane 2C and 4C: Whole *E. coli* and *S. typhi*, respectively

### HPLC analysis of purified LPS

The purity of LPS isolated from *E. coli* and *S. typhi* was assessed by HPLC. The band profile of LPS extracted from *E. coli* was analyzed and compared to that of extra pure commercial standard. As shown in figure 2, the LPS chromatogram of purified LPS from *E. coli* overlaid with that of standard indicating high purity of the product. Chromatogram of purified *S. typhi* LPS also

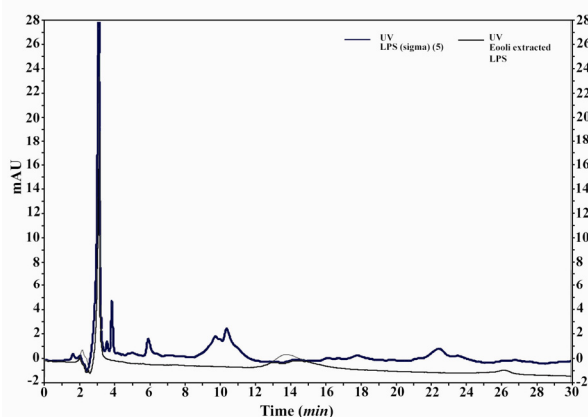


Figure 2. HPLC fractionation of LPS from *E. coli*. The gray and blue lines show chromatograms of purified *E. coli* LPS and extra pure commercial LPS from *E. coli*, respectively

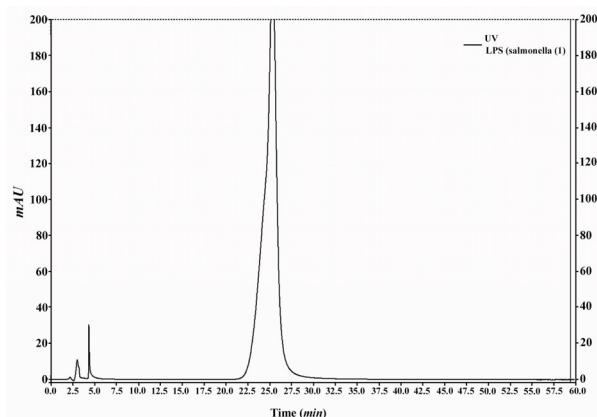


Figure 3. HPLC chromatogram of LPS extracted from *S. typhi*

showed a single major sharp band suggesting very low content of impurities (Figure 3).

### Assessment of LPS functional activity by LAL test

The principle of LAL coagulation assay is based on the fact that endotoxin activates the Limulus Amoebocyte Lysate (LAL) pro enzyme, resulting in gel formation. The result of our qualitative LAL coagulation assay indicated the functional activity of purified LPS as demonstrated by the formation of gel in vials containing the LAL.

### Evaluation of purified LPS pyrogen activity by rabbit pyrogen test

The endogenous pyrogen activity of purified LPSs was evaluated using rabbit pyrogen test. The initial rectal temperatures of the two rabbits were 38.3°C and 38.4°C. Injection of LPS from *E. coli* and *S. typhi*, caused raising the rectal temperature up to 39.7°C and 39.9°C respectively. The control rabbit which received PBS did not show considerable fluctuation of body temperature.

## Discussion

LPS is the main component of cell membrane of almost all gram negative bacteria. It is responsible for the pathological consequences of gram negative bacterial infections. Such life threatening diseases as septic shock following infection with gram negative bacteria is mediated mainly by LPS. LPS is potent activator of immune system capable of triggering cytokine release from cells of different origin. It is widely used as an inducer

of TLR-4 signaling pathway <sup>(16)</sup> and as a mediator of dendritic cell maturation <sup>(17)</sup>.

In this context, several attempts have been made so far to introduce a dozen of techniques for extraction and purification of LPS from different strains of gram negative bacteria <sup>(6-13,18-22)</sup>. Some techniques are technically or instrumentally demanding and so simple, cost-effective and at the same time reliable methods for isolation of pure LPS would be of great value.

The method introduced by Westphal <sup>(14)</sup> is still the most frequent procedure employed for LPS extraction because of its high yield. Because of the presence of contaminating substances which are introduced to the final purified LPS during extraction and purification process and their interfering effects in most downstream immunological and biological experiments, a practical approach for their elimination is necessary. Potential contaminants include capsular polysaccharide, nucleic acids and proteins, particularly outer membrane proteins that have high potential to bind LPS with high affinity <sup>(23)</sup>. Proteinase K-mediated bacterial protein digestion followed by nuclease elimination of contaminating RNA and DNA and phenol-water extraction method results in highly pure LPS free of protein and nucleic acids <sup>(24)</sup>.

In the present study, this procedure with some modifications has been employed for extraction and purification of LPS from two commonly-encountered bacterial strains; *E. coli* and *S.typhi*. In our protocol, we first disrupted bacterial cell wall by sonication followed by proteinase and nuclease treatment. Early treatment with the enzymes would allow early elimination of contaminating components which considerably enhances purity in the next steps.

Furthermore, hot phenol-mediated extraction in the next step results in elimination of residual amounts of the enzymes added in the second step. This order of treatments would conceivably yield purified LPS with the lowest amount of contaminating protein and nucleic acids.

Purified LPSs were characterized by SDS-PAGE electrophoresis followed by silver and commassie blue staining and HPLC. The results of silver staining clearly showed the ladder pattern of bands with multiple rungs which is characteristics of smooth type of gram negative bacteria due to the carbohydrate chain length variation of the O-antigen portion. LPS can be classified as either smooth or rough type based on the presence or absence of ladder like structure <sup>(25-28)</sup>. The rough form of LPS does not possess a ladder-like structure due to the lack of 'O' specific chain containing repeating units of oligopolysaccharides.

It is noteworthy that there was considerable variation in the number and band profile of LPS from *E.coli* and *S.typhi* which is due to difference in LPS structure. This variation in LPS structure forms the basis for various chemotype observed in gram negative bacteria <sup>(29)</sup>. The result of commassie blue staining of purified LPS revealed absence of contaminating bacterial proteins suggesting the effectiveness of protein elimination by proteinase treatment. The result of silver staining was confirmed by HPLC analysis which showed a sharp band of purified LPS in both cases.

### Conclusion

Although the purity of LPS is a good measure of the performance of purification system, functional activity of the final product is important as well. In this context, the results of rabbit pyrogen and LAL coagulation tests clearly proved the functional activity of the purified product. In conclusion, the protocol presented here could be employed for isolation of LPS with high purity and functional activity from different strains of smooth gram negative bacteria which have structurally different LPS.

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## References

1. Rietschel ET, Brade H. Bacterial endotoxins. *Sci Am* 1992;267(2):54-61.
2. Shnyra A, Luchi M, Morrison DC. Preparation of endotoxin from pathogenic gram negative bacteria. In: Evans TJ (eds). *Methods in Molecular Medicine* (Vol. 36 Septic Shock Methods and Protocols). Totowa: Humana press; 2000,13-25.
3. Erridge C, Bennett-Guerrero E, Poxton IR. Structure and function of lipopolysaccharides. *Microbes Infect* 2002;4(8):837-851.
4. Luderitz O, Galanos Ch, Lehmann V, Nurminen M, Rietschel ET, Rosenfelder G, et al. Lipid A: Chemical structure and biological activity. *J Infect Dis* 1973;128(Suppl 1):17-29.
5. Luderitz O, Westphal O, Staub AM, Nikaido H. Isolation and chemical and immunological characterization of bacterial lipopolysaccharides. In: Weinbaum G, Kadis S, Ajl SJ (eds). *Microbial Toxins*. New York: Academic press; 1972,145-233.
6. Staub AM. Bacterial lipido-proteinopolysaccharides (O somatic antigens). Extraction with trichloroacetic acid. In: Whistler RL, Wolfan ML (eds). *Methods in Carbohydrate Chemistry*. New York: Academic press; 1965,92-93.
7. Morrison DC, Leive L. Fractions of lipopolysaccharide from *Escherichia coli* O111:B4 prepared by two extraction procedures. *J Biol Chem* 1975;250(8):2911-2919.
8. Delahooke DM, Barclay GR, Poxton IR. A reappraisal of the biological activity of bacteroides LPS. *J Med Microbiol* 1995;42(2):102-112.
9. Sonesson A, Jantzen E, Bryn K, Larsson L, Eng J. Chemical composition of lipopolysaccharide from *Legionella pneumophila*. *Arch Microbiol* 1989;153(1):72-78.
10. Eidhin DN, Mouton C. A rapid method for preparation of rough and smooth lipopolysaccharide from bacteroides, *Porphyromonas* and *Prevotella*. *FEMS Microbiol Lett* 1993;110(2):133-138.
11. Galanos C, Luderitz O, Westphal O. A new method for the extraction of R lipopolysaccharide. *Eur J Biochem* 1969;9(2):245-249.
12. Nurminen M, Vaara M. Methanol extracts LPS from deep rough bacteria. *Biochem Biophys Res Commun* 1996;219(2):441-444.
13. Perdomo R, Montero V. Purification of *E. coli* O55: B5 lipopolysaccharides by size exclusion chromatography. *Biotecnología Aplicada* 2006;23(2):124-129.
14. Westphal O, Jann K. Bacterial lipopolysaccharides. Extraction with phenol-water and further applications of the procedure. In: Whistler RL, Wolfan ML (eds). *Methods in Carbohydrate Chemistry*. New York: Academic press; 1965,83-91.
15. Yin ET, Galanos C, Kinsky S, Bradshaw RA, Wesler S, Luderitz O, et al. Picogram-sensitive assay for endotoxin: gelation of *Limulus polyphemus* blood cell lysate induced by purified lipopolysaccharides and lipid A from gram-negative bacteria. *Biochem Biophys Acta* 1972;261(1):284-289.
16. Kawai T, Akira S. The role of pattern-recognition receptors in innate immunity: update on toll-like receptors. *Nat Immunol* 2010;11(5):373-384.
17. De Smedt T, Pajak B, Muraille E, Lespagnard L, Heinen E, De Baetselier P, et al. Regulation of dendritic cell numbers and maturation by lipopolysaccharide in vivo. *J Exp Med* 1996;184(4):1413-1424.
18. AL-Harbi AH, Austin B. The immune response of turbot, *Scophthalmus maximus* (L.), to lipopolysaccharide from a fish-pathogenic cytophaga-like bacterium. *J Fish Disease* 1992;15(5):449-452.
19. Baba T, Imamura J, Izawa K. Immune protection in carp, *Cyprinus carpio* L., after immunization with *Aeromonas hydrophila* crude lipopolysaccharide. *J Fish Disease* 1988;11(3):237-244.
20. Ingram GA, Alexander JB. The immune response of the brown trout *Salmo trutta* to lipopolysaccharide. *J Fish Biol* 1980;16(2):181-197.
21. Saeed MO, Plumb AJ. Immune response of channel catfish to lipopolysaccharides and whole cell *Edwardsiella ictaluri* vaccines. *Dis Aquat Organ* 1986;2:21-25.
22. Salati F, Ikeda Y, Kusuda R. Effect of *Edwardsiella tarda* lipopolysaccharide immunization on phagocytosis in the eel. *Nippon Suisan Gakkaishi* 1987;53(2):201-204.
23. Hitchcock PJ, Morrison DC. The protein component of bacterial endotoxin. In: Rietschel E (eds). *Handbook of Endotoxin: Chemistry of Endotoxin*. Amsterdam: Elsevier Science Publishers; 1984, 339-375.
24. Apicella MA. Isolation and characterization of lipopolysaccharides. In: Evans TJ (eds). *Methods in Molecular Medicine* (Vol. 431 Bacterial pathogenesis). Totowa: Humana press; 2000,13-25.
25. Hendrick CA, Sequeira L. Lipopolysaccharide defective mutants of the wilt pathogen *Pseudomonas solanacearum*. *Appl Environ Microbiol* 1984;48(1):94-101.
26. Hitchcock PJ, Brown TM. Morphological heterogeneity among *Salmonella* lipopolysaccharides chemotypes in silver stained polyacrylamide gels. *J Bacteriol* 1983;154(1):269-277.

27. Kido N, Ohta M, Kato N. Detection of lipopolysaccharides by ethidium bromide staining after sodium dodecylsulfate-polyacrylamide gel electrophoresis. *J Bacteriol* 1990;172(2):1145-1147.
28. Sledjeski DD, Weiner RM. *Hyphomonas* Spp., *Shewanella* Spp., and other marine bacteria lack heterogeneous (ladder like) lipopolysaccharides. *Appl Environ Microbiol* 1991;57(7):2094-2096.
29. Chart H. Lipopolysaccharide chemotyping. In: Howard J, Whitcombe DM (eds). *Methods in Molecular Biology* (Vol. 46 Diagnostic bacteriology protocols). Totowa: Humana press; 1995:41-48.