

Conjugation of Monoclonal Antibodies to Super Paramagnetic Iron Oxide Nanoparticles for Detection of her2/neu Antigen on Breast Cancer Cell Lines

Fereshteh Shamsipour¹, Amir Hassan Zarnani², Roya Ghods¹, Mahmood Chamankhah³, Flora Forouzesh³, Sedigheh Vafaei¹, Ali Ahmad Bayat¹, Mohammad Mehdi Akhondi², Mohammad Ali Oghabian⁴, and Mahmood Jeddi-Tehrani^{1*}

1. Monoclonal Antibody Research Center, Avicenna Research Institute, ACECR, Tehran, Iran

2. Department of Immunology, Reproductive Biotechnology Research Center, Avicenna Research Institute, ACECR, Tehran, Iran

3. Nanobiotechnology Research Center, Avicenna Research Institute, ACECR, Tehran, Iran

4. Research Centre for Science and Technology in Medicine, Tehran University of Medical Sciences, Tehran, Iran

Abstract

Conjugation of monoclonal antibodies to super paramagnetic nanoparticles is an effective method for cancer diagnosis and treatment. In this study the humanized anti her2/neu monoclonal antibody- Herceptin- was conjugated to super paramagnetic iron oxide (SPIO) nanoparticles using EDC method. The concentration of the conjugated antibodies was measured by Bradford assay. The antibody-nanoparticle conjugates were incubated with SKBR-3 and T47D human breast carcinoma cell lines and the presence of the conjugates on cell surface was confirmed by Prussian blue iron staining method. Conjugation of Herceptin to SPIO resulted in a precipitate-free conjugate containing 20µg antibody/mg SPIO. Prussian blue iron-staining of cells showed successful binding of the conjugates to the cell surfaces. Conjugation of monoclonal antibodies to SPIO may be a useful method for detection of tumor cells, especially by MRI techniques.

Avicenna J Med Biotech 2009; 1(1): 27-31

***Corresponding author:**
Mahmood Jeddi-Tehrani,
Ph.D., Monoclonal Antibody
Research Center, Avicenna
Research Institute, ACECR,
Tehran, Iran, P.O. Box:
19615-1177
Tel: +98 21 22432020
Fax: +98 21 22432021
E-mail:
mahjed@yahoo.com
Received: 21 Feb 2009
Accepted: 21 March 2009

Keywords: Breast cancer, Conjugation, Herceptin, Nanomagnetic particles

Introduction

Molecular probes for biomolecular recognition are of great importance in the fields of chemistry, biology, medical sciences and in biotechnology as well. These probes have been used in studies of biological functions and in ultrasensitive detection of biological factors responsible for many diseases⁽¹⁾. On the other front, the developments of nontoxic and biocompatible magnetic particles have been disclosed for biological applications since mid-1980s⁽²⁾.

Recently, magnetic particles have attracted growing interest as high performance biomaterial which is used for transport and separation of cells or cell parts^(2, 3), MRI⁽⁴⁾, hyperthermia⁽⁵⁾ and drug delivery⁽⁶⁾. Biological samples such as blood, serum, cell suspensions and cell lysates are allowed to be exposed to specific ligand-coupled particles, and the captured molecules or cells are then rapidly separated using magnetic fields^(3,7,8).

Magnetic particles conjugated with anti tumor monoclonal antibodies provide a new approach to identify tumor cells.

Antibodies labeled with magnetic nanoparticles give magnetic signals on exposure to a magnetic field. Iron oxide particles are usually coated with different organic shells including dextran, albumin or polyethylene glycol. Coated nanoparticles can be manufactured with a variety of functional groups (such as amino, aldehyde, hydroxyl, sulfate and carboxyl groups) on their surfaces. Considering these properties, we used super paramagnetic iron oxide (nanomag-D-SPIO 20nm) with COOH group on the surface, for conjugation to a humanized anti her2/neu monoclonal antibody (Herceptin) as a cancer targeting antibody.

Materials and Methods

Nanomag-D-SPIO 20 nm nanoparticles (surface COOH) and MACS separator with MS columns were purchased from Micromod (Miltenyi Biotech GmbH, Germany). The breast carcinoma cell lines SKBR-3 and T47D were obtained from Pasteur Institute of Iran. Other reagents and chemicals were obtained from Merck and Sigma.

Conjugation of anti her2 antibody (Herceptin) with nanoparticles by EDC method

N-ethyl-N-(3-dimethyl aminopropyl) carbodiimide hydrochloride (EDC, 26mM) and 10 mM N-hydroxy succinimide (NHS) were dissolved in 0.1 M 2-(N-morpholino) ethane-

sulfonic acid (MES) buffer (pH=8.3). The mixture was added to 1 ml of 5 mg/ml nanomag-D-SPIO 20 nm nanoparticles, and shaken at room temperature for 2 hours.

The particles were washed twice with phosphate buffered saline (PBS) pH=7.4 and then 0.5-1 mg/ml of Herceptin was added to the activated particles. The mixture was shaken for 3 hours and the reaction was quenched by the addition of glycine for 30 minutes^(9, 10).

The unconjugated antibodies were separated from conjugated antibodies by MACS column. The amount of immobilized antibody was estimated based on the Bradford method.

Spectrophotometric measurement of Iron

Iron concentration of conjugated samples was obtained by potassium thiocyanate method^(12,13). In brief, samples were diluted with 300 μ l 6N HCl containing %1 H₂O₂; under this condition, the iron in the samples is dissolved and oxidized to ferric state. The samples were then added to a 5% solution of potassium thiocyanate where the Fe III formed a red complex with the thiocyanate which could be measured by absorbance at 480 nm.

Cell culture

The her2/neu expressing cell lines SKBR3 and T47D were grown in RPMI 1640 medium with 10% (v/v) fetal calf serum and %1 penicillin/streptomycin. Cells were incubated at 37°C containing 5% Co₂.

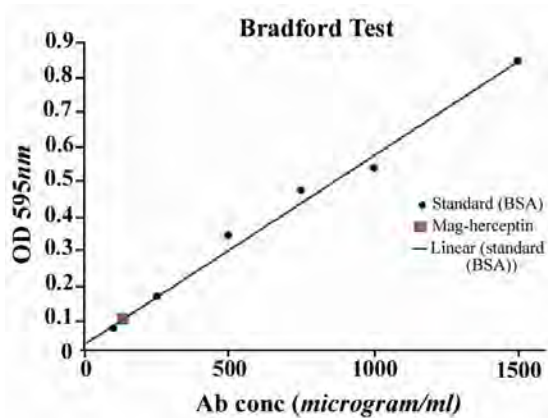


Figure 1. Antibody concentration measurement by Bradford assay (Ab conc. 100 μ g/ml)

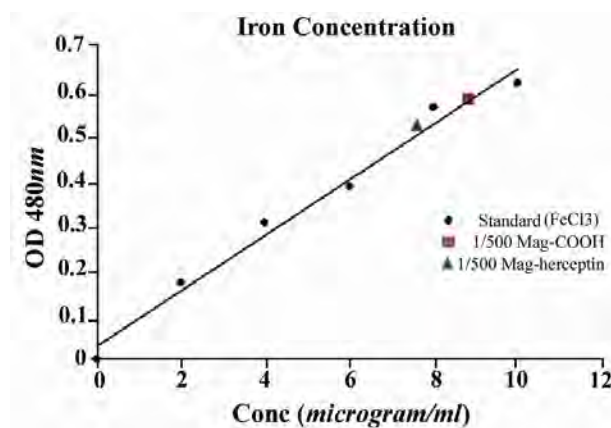


Figure 2. Magnetic nanoparticles concentration measurement by Potassium Thiocyanate method (particles conc. 4-4.5mg/ml)

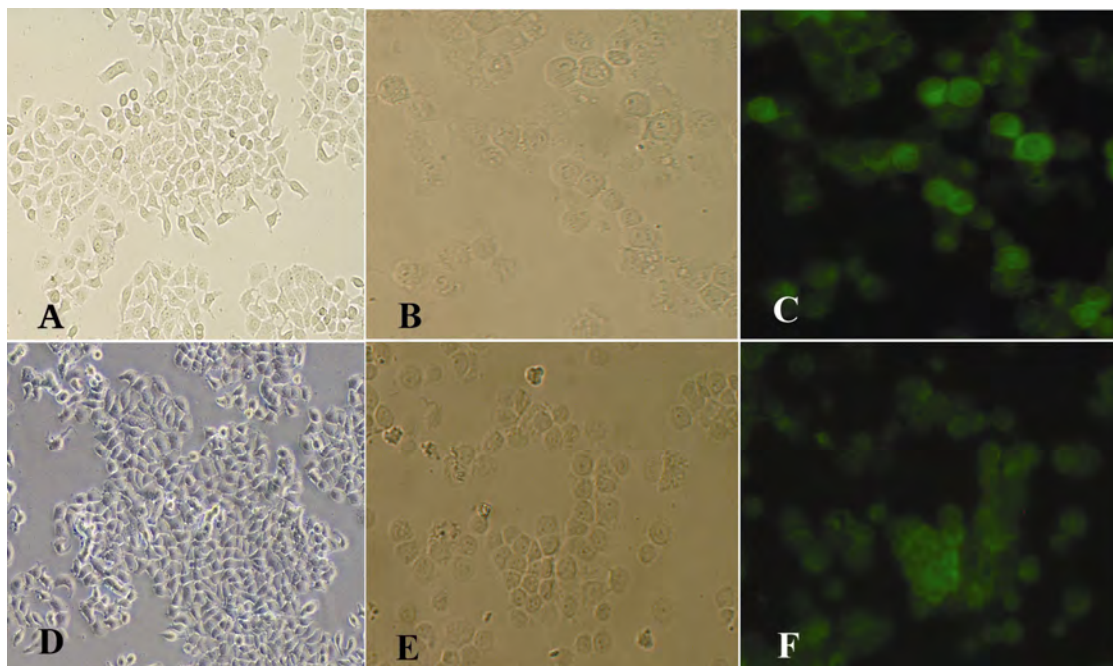


Figure 3. Light microscopy and Immunofluorescence of SKBR3 and T47D cell lines
 A: SKBR3 cells in culture, B: suspension of SKBR3 cells, C: SKBR3 cells after incubation with Herceptin-FITC, D: T47D cells in culture, E: suspension of T47D cells, F: T47D cells after incubation with Herceptin- FITC.

Immunofluorescence staining

To verify the expression of her2 proteins on the cells, the SKBR3 and T47D cells were incubated with anti her2/neu (Herceptin) at $10 \mu\text{g/ml}$ concentration for 1 hour at 37°C . After being washed in PBS, FITC-labeled anti human IgG (diluted 1/20, Avicenna Research Institute, Tehran, Iran) was added and incubated for 1 hour at room temperature. Cells were then observed directly on a fluorescence microscope (Olympus, Japan).

In vitro cell labeling

SKBR3 and T47D cells were counted and adjusted to a suspension of $4 \times 10^5 \text{ cells/ml}$; $100 \mu\text{l}$ of each cell suspension were cytopinned on microscope slides (Shandon cyto-spin 4, Thermo, Germany). The cells were incubated with $100 \mu\text{l}$ magnetic nanoparticles (with or without antibody; $5 \mu\text{g Ab}$ and 0.2 mg iron) for 1 hour at 37°C . Then cells were washed extensively with PBS to remove unbound particles.

The nanoparticles that bounded on the cell surface were detected by iron staining with Prussian blue staining method^(14, 15).

Results

We used 20 nm nanoparticles (a magnetic core covered with dextran) with carboxyl group for conjugation to Herceptin as a cancer targeting antibody. The final products of conjugation were suspensions without precipitate and the amount of immobilized antibody was $20\text{-}36 \mu\text{g Ab/mg}$ nanoparticles (Figure 1).

The amount of iron conjugated was determined by potassium thiocyanate method using $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ as a reference (Figure 2).

The SKBR3 and T47D cell lines showed high levels of her2/neu expression, after incubation with Herceptin-FITC (Figure 3), but not with the isotype control antibody (data not shown).

These results confirmed the presence of antigen on the cell surface. Specific binding of the conjugates to the cells can be determined by using electron microscopy, MRI imaging and iron staining^(13,18,19).

In this study after incubation of the cells with conjugated nanoparticles, the blue stain induced by iron staining showed the accumulation of conjugated nanoparticles on the cell surface (Figure 4).

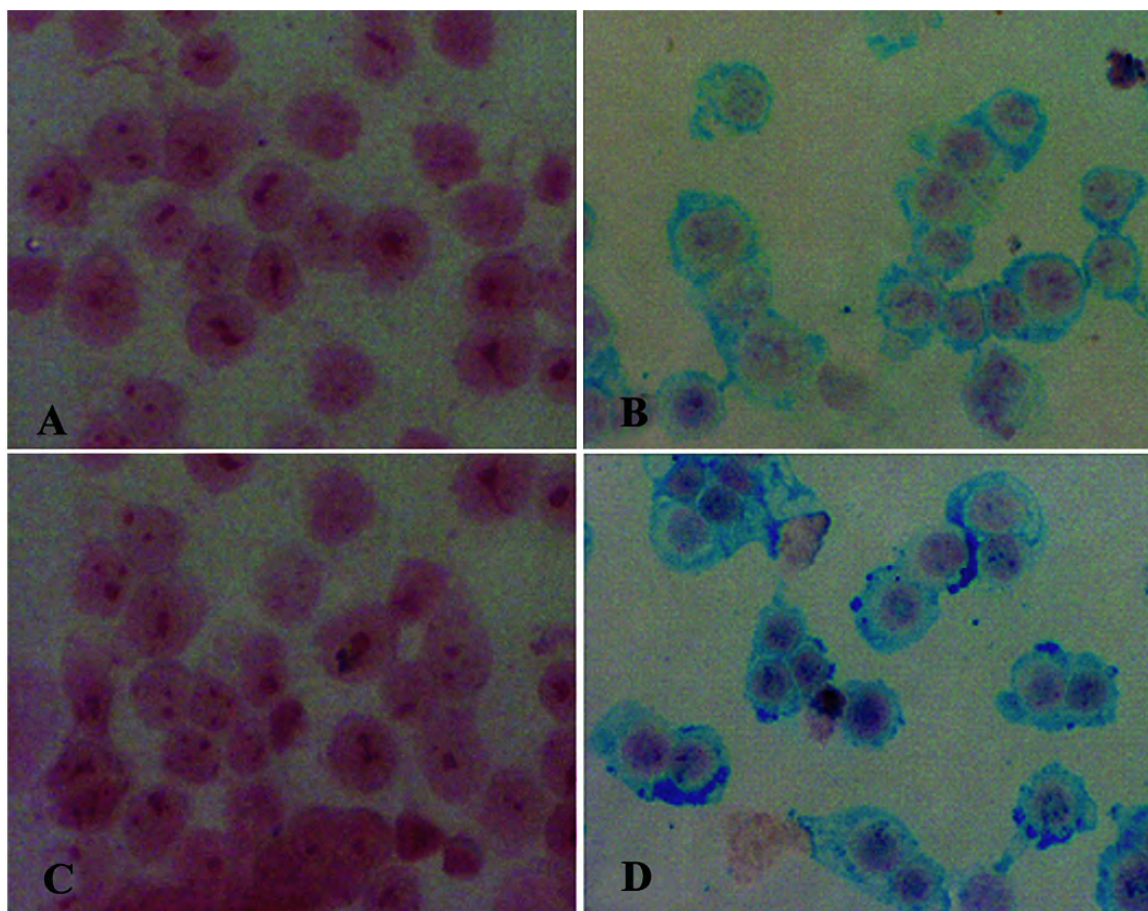


Figure 4. Iron staining of cells after incubation with conjugated nanoparticles
A: SKBR3 cells, B: KBR3 with iron staining, C: T47D cells, D: with iron staining.

Discussion

Polymer-coated magnetic particles with particle sizes 5-500 nm have been employed in medicine or biotechnology for many years⁽¹⁶⁾. In this study the 20 nm nanoparticles were coupled via their surface carboxyl group to the amino groups on the Herceptin antibody using the EDC method^(10, 17).

After conjugation, the amount of immobilized antibody was approximately 20 µg/mg magnetite. However by increasing concentration of antibody during the process, the efficiency of the conjugation did not improve. In other studies the efficiency of conjugation has been reported as 5-20 µg Ab/mg particles^(9,18). Conjugated nanoparticles bound specifically to the her2/neu antigen.

Iron staining, confirmed the presence of nanoparticles on the cell surface. In the present study we showed specific binding of Herceptin-nanomagnetic particle conjugates

to her2/neu over expressing cells, suggesting a future application of Herceptin-magnetite for MR imaging of breast cancer.

Acknowledgment

This work was supported by a grant from the Nanotechnology Committee of Iran's Ministry of Health and Medical Education.

References

1. Santra S, Zhang P, Wang K, Tapeç R, Tan W. Conjugation of biomolecules with luminophore-doped silica nanoparticles for photostable biomarkers. *Anal Chem* 2001;73(20):4988-4993.
2. Gruettner C, Teller J, Westphal F, Ivkov R. Magnetic nanoparticle compositions and methods related thereto. Patent application publication 2005. Pub No: US 2005 /0271745 A1 Pub Date: Dec 8, 2005.
3. Nagasaki Y, Kobayashi H, Katsuyama Y, Jomura T, Sakura T. Enhanced immunoresponse of

- antibody mixed-PEG coimmobilized surface construction of high performance immuno-magnetic ELISA system. *J Colloid Interface Sci* 2007;309(2):524-530.
4. Baio G, Fabbi M, d.Totero D, Ferrini S, Cilli M, Derchi LE, Neumaier CE. Magnetic resonance imaging at 1.5T with immunospecific contrast agent in vitro and in vivo in a xeno-transplant model. *Mag Reson Mater Phy* 2006;19:313-320.
 5. Itoa A, Kugaa Y, Hondaa H, Kikkawab H, Horiuchib A, Watanabeb Y, et al. Magnetite nanoparticle-loaded anti-her2 immunoliposomes for combination of antibody therapy with hyperthermia. *Cancer Lett* 2004;212(2):167-175.
 6. Kawasaki ES, Player A. Nanotechnology, nanomedicine, and the development of new, effective therapies for cancer. *Nanomedicine* 2005;1(2): 101-109.
 7. Lawson EL, Clifton JG, Huang F, Li X, Hixson DC, Josic D. Use of magnetic beads with immobilized monoclonal antibodies for isolation of highly pure plasma membranes. *Electrophoresis* 2006;27(13):2747-2758.
 8. Lee JH, Huh YM, Jun YW, Seo JW, Jang JT, Song HT, et al. Artificially engineered magnetic nanoparticles for ultra-sensitive molecular imaging. *Nat Med* 2007;13(1):95-99.
 9. Gruttnera C, Mullera K, Tellera J, Westphala F, Foremanb A, Ivkovb R. Synthesis and antibody conjugation of magnetic nanoparticles with improved specific power absorption rates for alternating magnetic field cancer therapy. *J Magn Mater* 2007;311:181-186.
 10. Denardo SJ, Denardo G, Miers L, Natarajan A, Foreman A, Gruettner C, et al. Development of tumor targeting bio probes (¹¹¹In-Chimeric L6 Monoclonal Antibody nanoparticles) for alternating magnetic field cancer therapy. *Clin Cancer Res* 2005;11(19 Pt2):7087s-7092s.
 11. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 1976;72:248-254.
 12. Owen CS, Sykes NL. Magnetic labeling and cell sorting. *J Immunol Methods* 1984;73(1):41-48.
 13. Funovicsa MA, Kapellerb B, Hoellerc C, Sud HS, Kunstfeldc R, Puiga S, et al. MR imaging of the her2/neu and 9.2.27 tumor antigens using immunospecific contrast agents. *Magn Reson Imaging* 2004;22(6):843-850.
 14. Bancroft JD, Stevens A. *Theory and Practice of Histological Techniques*, 4th ed. New York: Churchill Livingstone Inc;1996.
 15. Luna LG. *Manual of Histological Staining Methods of the AFIP*. 3rd ed. New York: McGraw Hill; 1968.
 16. Wagner K, Kautz A, Ro" der M, Schwalbe M, Pachmann K, Clement JH, et al. Synthesis of oligonucleotide-functionalized magnetic nanoparticles and study on their in vitro cell uptake. *Appl Organometal Chem* 2004;18(10):514-519.
 17. Prestvik WS, Berge A, Mork PC, Stenstad PM, Ugelstad J. Preparation and application of monosized particles in selective cell separation. In: Hafeli U, Schutt W, Teller J, Zborowski M (eds). *Scientific and clinical applications of magnetic carriers*. New York: Springer;1997,11-35.
 18. Hilger I, Trost R, Reichenbach JR, et al. MR imaging of her2/neu protein using magnetic nanoparticles. *Nanotechnology* 2007;18(13):135103-135111.
 19. Zhang C, Jugold M, Woenne EC, Lammers T, Morgenstern B, et al. Specific targeting of tumor angiogenesis by RGD-conjugated ultra-small superparamagnetic Iron oxide particles using a clinical 1.5-T magnetic resonance scanner. *Cancer Res* 2007;67(4):1555-1562.